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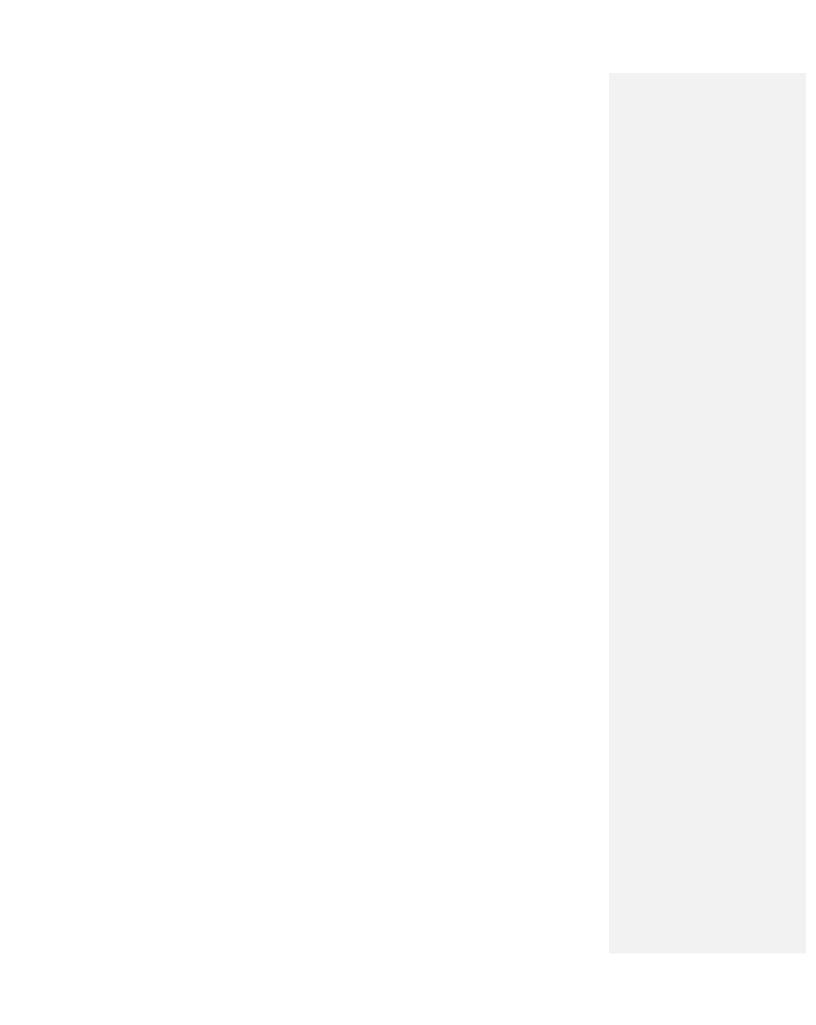
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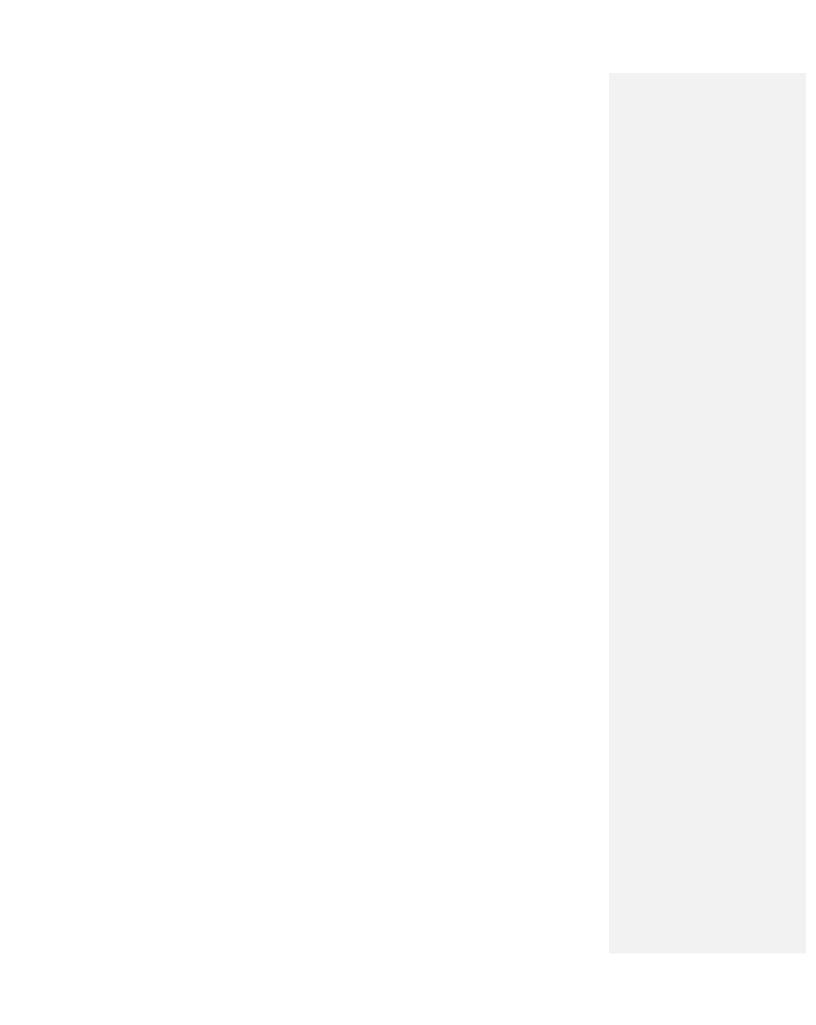
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# <sup>1</sup> The ocean's ultimate trashcan: Hadal trenches as

<sup>2</sup> major depositories for plastic pollution

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11 Keywords: microplastic; marine debris; sediment; hadal trench; ecosystems

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Abstract: Plastic debris and marine microplastics are being discharged into the ocean at an alarming scale and have been observed throughout the marine environment. Here we report microplastic in sediments of the Challenger Deep, the deepest known region on the planet, abyssal plains and hadal trenches located in the Pacific Ocean (4900 m to 10890 m). Microplastic abundance reached 71.1 items per kg dry weight sediment. That high concentrations are found at such remote depths, knowing the very slow sinking speed of microplastics, suggests

that supporting mechanisms must be at-play. We discuss cascading processes that transport microplastics on their journey from land and oceanic gyres through intermediate waters to the deepest corners of the ocean. We propose that hadal trenches will be the ultimate sink for a significant proportion of the microplastics disposed in the ocean. The build-up of microplastics in hadal trenches could have large consequences for fragile deep-sea ecosystems.

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### 25 1 Introduction

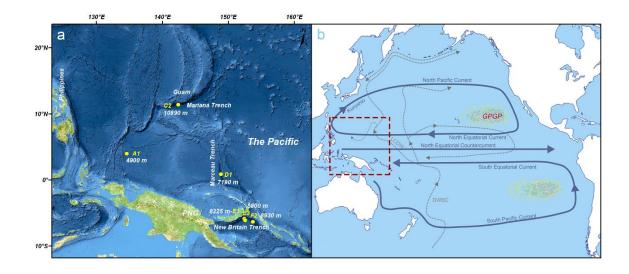
26 The Earth has experienced drastic changes since the beginning of Anthropocene. A particular 27 period, the so-called Plastic Age, has attracted mass attention (Carpenter and Smith, 1972). 28 Considering their transient hundred-year history (Crawford and Quinn, 2017), plastics have the 29 potential to challenge the productivity, diversity and function of marine ecosystems. The extent 30 that plastic pollution has intruded the ocean is still unknown. An increasing number of 31 observations in different marine environments suggest it is becoming ubiquitous, and 32 accumulating in the ocean. Thompson et al. (2004) found plastic fragments in estuarine 33 sediments around UK, followed by discoveries of microplastics in oceans around the world, 34 including the Polar Regions (Obbard et al. 2014) and the deep sea (Van Cauwenberghe et al. 35 2013). Extensive studies on the abundance of microplastics in coastal beaches, estuaries, 36 offshore waters and marginal seas have been conducted globally (Lusher 2015; Browne et al. 37 2015; Bosker et al. 2017). Since the discovery of the Great Pacific Garbage Patch (GPGP) 38 (Moore et al. 2001) in the center of the North Pacific gyre, the GPGP has been found to 39 accumulate plastics rapidly during the past two decades – four to sixteen times the concentration 40 previously reported, with microplastics consisting 94% of floating pieces (Lebreton et al. 2018). 41 Nevertheless, theoretical estimates of the amount of microplastics in the surface ocean from 42 input estimates heavily outweigh the measured concentrations (Eriksen et al. 2014). This begs 43 the question "where has all the plastics gone".

Investigation into the vertical distribution of microplastics in the water columns (La Daana et al. 2018) suggests that the fate for microplastics is not only the surface ocean, but also the deep sea (Woodall et al. 2014), including the most remote trenches (Jamieson et al. 2019). Although the density of certain types of plastic debris is lighter than seawater, which leads to their initial 48 accumulation in surface water, it is likely that the breakdown of plastic debris causes the 49 formation of microplastics and sinking within the water columns. Sinking factors include not 50 only density of plastic polymers, but also biofouling that leads to the sinking of almost every 51 category of commercial plastics to the seafloor (Engler 2012). Photodegradation may cause 52 plastics to progressively break down, and a loss of microplastics smaller than 1 mm at the open 53 ocean surface was observed (Cozar et al. 2014) compared to model estimation (Lebreton et al. 54 2012), indicating mechanisms quickly transferring microplastics from surface to the deep must 55 be at-play. Even buoyant polymers like polyethylene, when incorporated into marine snow, have been shown to sink at a rate of 818 m day<sup>-1</sup> (Porter et al. 2018). However, the vertical change of 56 57 microplastics in water column from sea surface to the ocean floor remains unknown.

58 The hadal zone is the deepest part of the ocean (6000-11000 m), which accounts for 45% of 59 the depth range of the ocean (Jamieson et al. 2010). While most studies on microplastics in 60 seabed sediments have mostly been undertaken in coastal areas and continental shelves, very few 61 studies have been done on sediments in the deep-sea or the hadal zone. It has been shown that 62 marine litter is widespread on the deep sea floor, with plastics often being the most common type 63 of marine litter encountered (Mordecai et al. 2011). Compared to microplastics, larger marine 64 debris in deep-sea environments is more frequently reported in deep diving expeditions. 65 Deep-sea trawls in the Gulf of Mexico (at a depth between 250-3650 m) revealed that most 66 man-made waste on the seabed were plastics (Wei et al. 2012). The majority of marine debris in 67 the Ryukyu Trench were plastics, especially fragmented plastic bags (Shimanaga and Yanagi 68 2016). Based on a video database over 22 years, Schlining et al. (2013) found that anthropic 69 marine debris accumulated on the steepest slopes of the Monterey canyons was mostly plastics. 70 By reviewing images and video clips from Deep-sea Debris Database developed by JAMSTEC,

71 Chiba et al., (2018) found increasing marine debris in the abyss over the past 30 years, with the 72 deepest record of one plastic bag at 10890 m from Mariana Trench. Miyake et al. (2011) 73 reviewed video surveys of submersible dives in Japan Trench and Ryukyu Trench down to 7216 74 m and noted plastics were the most common marine debris. Using photographic time-series 75 surveys to investigate the change in concentration of marine litter on seafloor from 2002-2014 at 76 depth of 2500 m in the Arctic, there were more litter on the seafloor than at the surface (Tekman 77 et al., 2017). Those findings all support the theory that portions of plastic debris at the sea 78 surface eventually sink to the seafloor, increasing the likelihood of deposition at the deepest and 79 remotest corner of the ocean.

To investigate to what degree microplastics have transported to the hadal zone, and to understand the fate of missing microplastics from sea surface, this study investigated microplastics in sediment samples collected from abyssal plain and some of the deepest trenches in the hadal zone, reviewed the potential cascading processes that transport microplastics to remote hadal trenches, and proposed hadal trenches will be the ultimate sink for a significant proportion of the microplastics.



86

87 Figure 1. Map of the six sampling sites and a schematic of ocean circulation in surface and deep 88 water in the Pacific Ocean (modified from Talley et al., 2011). (a) Sampling sites are marked 89 with yellow dots. C1: Challenger Deep, Mariana Trench; A1: north of Papua New Guinea 90 (PNG), east of the Philippines, abyssal plain; D1: Marceau Trench; E1, E2 and F2 are located in 91 the New Britain Trench. (b) Red dotted square shows the location of (a). Surface circulation are 92 marked by blue solid lines and abyssal circulation are marked by grey dotted lines. GPGP: Great 93 Pacific Garbage Patch; DWBC: Deep Western Boundary Current; LCDW: Lower Circumpolar 94 Deep Water

95

### 96 2 Materials and Methods

### 97 2.1 Sampling

98 Sediment samples from abyssal plain and hadal trenches were collected by Lander (sites 99 except E3) or a box corer (site E3) during sea trials of Autonomous and Remotely-operated 100 Vehicle (ARV) Rainbowfish in December, 2016 (Figure 1). The 4800-tonnes Scientific Research 101 Ship Zhang Jian is the main supporting vessel for Rainbowfish. Samples were collected from one 102 abyssal plain near the Philippines (4800 m), the Challenger Deep in the Mariana Trench (10890 103 m), Marceau Trench (7190 m), and the New Britain Trench (5800 m, 8225 m, 8930 m). Detailed 104 information on geographical coordinates, sediment description, and grain size distribution of 105 sediment samples can be found in Table 1. Landers were equipped with a multitude of deep-sea 106 apparatuses, which took samples of hadal settings including water, sediment and macro 107 organisms. Tubes are equipped on the Lander to collect hadal sediment samples. Only top layers 108 of samples from each sampling sites were taken for microplastic analysis (refer to Figure S1 for

photos of sampling tools). Back in the Mobile Hadal Science Laboratory, sediment samples were
carefully wrapped with aluminum foil, after which they were put into clean lunch boxes and
stored at -80 °C in the fridge.

112	Table 1. Detailed information on geographical coordinates, sediment description, and grain size
113	distribution of sediment samples from six sampling sites.

Sampling site	Location	Depth [m]	Longitude [degrees_east]	Latitude [degrees_north]	Sampler	Sediment Description	Sand [%]	Silt [%]	Clay [%]
A1	Abyssal plain east of Philippines	4900	134.6021	3.9954	Lander	Brown, Soft	9.4	47.8	42.8
E3	New Britain Trench	5800	152.5609	-6.1006	Box corer	Deep Brown, Soft	7.7	58.0	34.3
D1	Marceau Trench	7190	148.8887	0.8975	Lander	Deep Brown, Soft	2.7	51.5	45.8
E1	New Britain Trench	8225	152.4252	-5.8662	Lander	Deep Brown, Soft	3.5	64.4	32.1
F2	New Britain Trench	8930	153.7430	-6.3258	Lander	Yellow, Soft, with tephros	9.9	74.3	25.8
C2	Challenger Deep, Mariana Trench	10890	142.4041	11.3537	Lander	Deep Brown, Soft	4.3	66.4	29.3

114

# 115 **2.2 Density separation and numeration**

For each site, about 500 g wet weight sediment were used for the analysis of microplastics.
Samples were firstly weighted using analytical balance (TB-2002, Denver Instrument, USA),
freeze-dried at -80°C using Christ Alpha 1-4 LDplus (Germany) and dry weight was determined.

119 An aliquot of 25 g dry weight sediment per sample from six sites (3 replicates for each site) were 120 utilized for density separation (18 samples in total). Microplastics were extracted from the sediments according to Peng et al. (2017). Saturated sodium iodide solution (NaI, 1.8 g mL<sup>-1</sup>) 121 122 was applied for density separation to improve extraction efficiency. The solution was added to a 123 beaker with 25 g dry sediment, stirred for 2 min with a clean glass rod and settled down for 24 h. 124 The supernatant was filtered using a vacuum pump, and microplastic particles were collected onto a filter membrane (RAWP04700-MF-Millipore<sup>™</sup> Membrane Filter, 1.2 µm pore size). The 125 126 filter was dried in room temperature until constant weight. The filter was then observed under 127 dissecting microscopes (Leica M165 FC, Germany) for numeration. Shape, size and color of 128 microplastics were categorized.

#### 129 **2.3 Microplastic identification**

130 Because only thin particles can be compressed in the diamond cell for the µ-FT-IR instrument, 131 one fragment was too big, and was tested using the FT-IR instrument (Figure 2c). All the other particles were identified by µ-FT-IR. ATR mode of µ-FT-IR instrument (Thermo Fisher Nicolet 132 133 iN10) were used to identify microplastic polymer types and Transmittance mode of FT-IR 134 instrument (Thermo Fisher Nicolet iS5) was applied in one big particle based on different size 135 limit for the two types of FT-IR instruments. Grain size distribution was performed using laser 136 particle analyzer (Coulter LS13 320). Field-emission Scanning Electron Microscopy (SEM, FEI 137 Sirion 200) was used to characterize morphology of microplastics.

# 138 2.4 QA/QC

Quality assurance and quality control must follow the strictest procedures to prevent anycontamination, especially airborne contamination (Woodall et al., 2014). All analyses were

141 conducted in a laminar flow cabinet (Airtech SW-CJ-1FD). All the glassware and metal utensils 142 were muffled at 450 °C for 3 h before usage to prevent microfiber contamination. Chemicals and 143 Milli Q water were filtered prior to usage. Filter membranes were examined under microscopes 144 to check for possible contamination before usage. Procedural blanks were performed at each step 145 of density separation of each single sample. Blank 1 was set during the preparation of saturated 146 NaI solution. Blank 2 was set during the filtration of each sediment sample. Blank 3 was set for 147 the same volume of Milli Q water during density separation. In all the 24 procedural blanks 148 during the experiment, no fiber or other contamination was found under microscopic inspection.

## 149 **2.5 Recovery rate and statistical analysis**

Recovery rate experiments were conducted using commercial PP resin pellets (4.37±0.29 mm), lab-made PP fibers (1.66±0.59 mm) and lab-made PET fibers (1.07±0.52 mm). This consideration of particle shape and size used for recovery test took environmental abundance, microplastic size and actual polymer composition into consideration. PP resin pellets, lab-made PP and PET fibers were spiked into soft hadal sediments collected where microplastics had been removed using density separation. Statistical analysis was done using IBM SPSS Statistics 22.

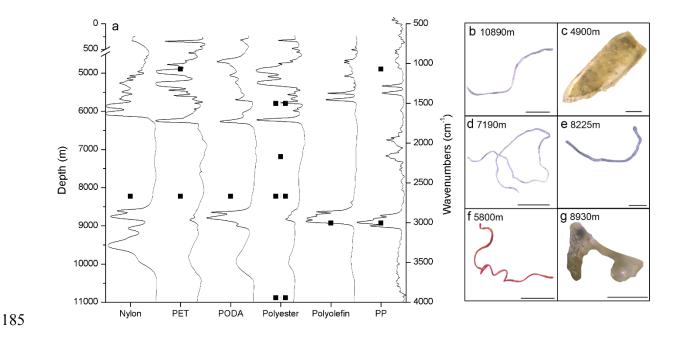
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#### 157 **3 Results and Discussion**

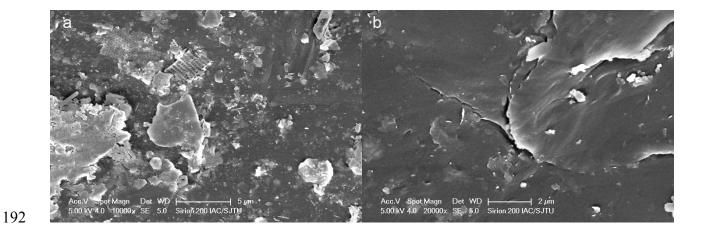
### 158 **3.1 Characteristics of microplastics in hadal trenches**

Microplastics were found at all six sampling sites in hadal trenches and abyssal plain. In total, all 32 microplastic particles were extracted from the 3 kg sediment samples (wet weight) and the average abundance of microplastics in sediments was 1.78 items per 25 g dry weight sediment,

162 i.e., 71.1 items per kg dry weight sediment. Assuming the density of sediments from the Mariana 163 Trench to be 1.26 g cm<sup>-3</sup> (Glud et al., 2013), the average abundance by volume was 89.6 items 164 per L dry weight sediment. The highest abundance was found at E1 in the New Britain Trench, 165 with 3.33 items per 25 g dry weight sediment (i.e., 133.3 items per kg dry weight sediment). The 166 lowest abundance was 0.67 items per 25 g dry weight at station D1 in the Marceau Trench (i.e., 167 26.7 items per kg dry weight sediment). The properties of the microplastics (excluding rayon) 168 from six sites are shown in Table 2. Rayon fibers and fragments from six sites were also 169 identified (Table S1), and because procedural blanks indicated no microfiber contamination 170 occurred, rayon fibers are listed in the result as a reference recognizing the debate on whether 171 rayon should be classified as microplastics (Peeken et al., 2018). Transparent particles 172 (excluding rayon) constituted 64.3% of total particles, followed by red particles (21.4%) and blue 173 particles (14.3%). Fibers (excluding rayon) were the major shape of microplastics, constituting 174 of 57.1% of all the particles, while fragments constituted 42.8% of total microplastics, a 175 significant higher percentage than those in coastal sediments (Frias et al., 2016). Size of 176 microplastics ranged from  $73-12376 \mu m$  (Table 2). The most prevalent type of polymers was 177 polyester (50%). Only one polypropylene (PP) fragment (Figure 2c) from A1 had to be identified 178 by FT-IR due to its relatively larger size, which led to the slightly different spectrum compared 179 to other polymer spectra obtained by µ-FT-IR (Figure 2a). The surface texture of this PP 180 fragment was characterized by SEM analysis (Figure 3) and shows attached organic matter and 181 apparent signs of degradation, suggesting that plastic particles may sink due to biofouling and 182 continue to fragment into microplastics through ingestion (Dawson et al., 2018) even in the 183 deep-sea floor. In this study, recovery rates for PP pellets, PP fibers and PET fibers were 100%, 184  $94.4 \pm 4.8\%$  and  $89.8 \pm 5.8\%$  (mean  $\pm$  SD, n=3, CI=95%), respectively (Table S2).



**Figure 2.** Microplastics in sediments in the abyssal plain and hadal trenches, and imaged examples of microplastics from each site at various depth. (a) Distribution of microplastics categorized by polymer type (excluding rayon, marked as black squares) and infrared spectra of six kinds of polymers identified using FT-IR instruments. (b) Fiber (ID: C2-2-2, polyester), (c) fragment (A1-1-2, PP), (d) fiber (D1-2-1, polyester), (e) fiber (E1-2-3, nylon), (f) fiber (E3-3-1, polyester), and (g) film (F2-1-3, polyolefin).



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Figure 3. SEM images of the surface texture of one PP fragment (ID: A1-1-2). (a) showing
organic matter attached to the surface of microplastic particle with magnification of 10000×, and
(b) showing signs of degradation on the surface with magnification of 20000×.

196 **3.2 Microplastic abundance** 

197 This study found that microplastics were present in sediments from some of the deepest 198 corners on earth, including the Mariana Trench, Marceau Trench and New Britain Trench at 199 depths from 4900 to 10890 m. The highest abundance of microplastics was 133.3 items per kg 200 dry weight sediment. The high concentration of microplastics in three deepest hadal trenches of 201 the planet suggests that microplastics pollution has reached the full ocean depth and deposit at a 202 high concentration, making hadal trenches the major depositories and ultimate sink for 203 microplastics.

Table 2 A total of 14 identified microplastics (excluding rayon) in six sampling sites in abyssal
 plain and hadal trenches, illustrating the characteristics of microplastics.

Samplin g site	Location	Dept h [m]	Abundance [items/ L dry weight]	Abundanc e [items/kg dry weight]	Microplasti c ID	Shape	Color	Size [µm]	Polymer [Match %]	Polyme r Density [g/cm <sup>3</sup> ]
A1	Abyssal plain east of	4900	67.2	53.3	A1-1-1	Fragmen t	Red	115	PET (94.73)	1.30-1.5 0
	Philippine s				A1-1-2	Fragmen t	Transpare nt	6068	PP (82.10)	0.88-1.2 3
E3	New Britain Trench	5800	67.2	53.3	E3-2-2	Fiber	Blue	1760	Polyester (96.49)	1.30-1.5 0
					E3-3-1	Fiber	Red	2895	Polyester (91.83)	1.30-1.5 0
D1	Marceau Trench	7190	33.6	26.7	D1-2-1	Fiber	Transpare nt	8298	Polyester (97.09)	1.30-1.5 0
E1	New Britain	8225	168.0	133.3	E1-2-1	Fragmen t	Blue	105	Polyester (82.62)	1.30-1.5 0

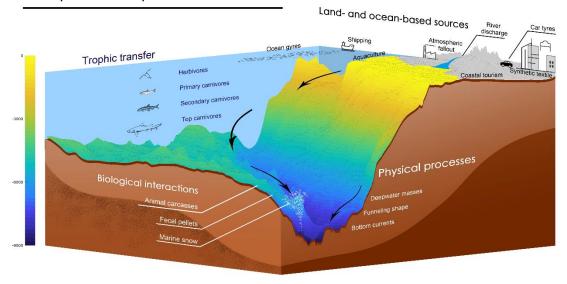
	Trench									
					E1-2-3	Fiber	Transpare nt	1204	Nylon (98.38)	1.13-1.3 8
					E1-3-2	Fiber	Transpare nt	1009	Polyester (95.67)	1.30-1.5 0
					E1-3-4	Fragmen t	Red	73	PET (99.01)	1.30-1.5 0
					E1-3-5	Film	Transpare nt	491	Poly(octadec yl acrylate) (98.32)	0.90
F2	New Britain Trench	8930	84.0	66.7	F2-1-3	Film	Transpare nt	422	Polyolefin (85.51)	0.86
					F2-1-4	Fiber	Transpare nt	774	PP (97.25)	0.88-1.2 3
C2	Challenge r Deep, Mariana	1089 0	117.6	93.3	C2-2-2	Fiber	Transpare nt	1058	Polyester (83.40)	1.30-1.5 0
	Trench				C2-2-5	Fiber	Transpare nt	1237 6	Polyester (98.30)	1.30-1.5 0

206

207 High hydrostatic pressure and extreme environments make hadal trenches out of reach by 208 most human activities except anthropogenic pollutants. While some deep-sea surveys have visually identified marine litter on the deep-sea floor (Miyake et al., 2011), few have 209 210 successfully sampled and quantified microplastics due to the technical challenges of collection 211 and retrieval of samples from the hadal zone below 6500 m (Cui et al., 2014), e.g., the 212 development of full ocean depth ROV/ARV/HOV, and the collection and preservation of hadal 213 samples under 1100 MPa. Therefore, sinking mechanisms for microplastics entering the hadal 214 environment remain largely unknown due to limited research on microplastic distribution in the 215 deep-sea and hadal environments and lack of knowledge on hadal science. Here we propose 216 sources and potential pathways for microplastics to enter the hadal trenches from land and 217 sea-based sources, which contributing to microplastics export to the seafloor, making hadal 218 trenches as major depositories and ultimate sink for microplastics.

219 Our results illustrate that microplastics in sediments from hadal trenches are even higher than 220 those identified from the conventional "deep-sea". Microplastic particles have been found in 221 sediments in the Atlantic Ocean at depth from 1100-5000m with an average abundance of 0.5 items per 25 cm<sup>2</sup> (Van Cauwenberghe et al., 2013). In the Kuril-Kamchatka trench in the 222 223 North-West Pacific, fiber was the dominant type of microplastics in sediment samples at depth 224 from 4869 and 5766 m (Fischer et al., 2015). Deep-sea cores from the Mediterranean, 225 South-West Indian Ocean and North-East Atlantic Ocean at a depth down to 3500 m revealed 226 that the abundance of microplastic fibers in sediments was several orders of magnitude higher 227 than that in surface waters (Woodall et al., 2014).

228 Recently, microplastics were also found in water and sediment samples in the Mariana Trench. According to Peng et al. (2018), microplastic abundance reached as high as 13.51 pieces  $L^{-1}$  in 229 bottom waters, and 2200 pieces  $L^{-1}$  in sediments. They compared microplastic abundance at 230 231 10903 m with 8 other studies, and found that the abundance is four times higher than that in the 232 offshore waters near Vancouver, and twenty times higher than deep-sea sediments from Atlantic 233 (Peng et al., 2018). This may be an example of hadal trenches as the sink for microplastics, 234 however, compared to the abundance in our study which collected samples from several hadal 235 trenches with larger sample quantity (500 g wet weight from each site), such high concentration 236 may attribute to direct conversion from 5 mL to 1000 mL for each sediment sample in their study 237 (Peng et al., 2018). While studies attempt to make comparisons among microplastic abundance 238 worldwide, comparison should be made among studies with same sampling method and 239 reporting units, as stressed in GESAMP WG40 report (GESAMP, 2016).





240

Figure 4. A schematic of pathways of microplastics from various sources entering the hadal trenches (taking the New Britain Trench as an example), making the deepest spots on Earth the ultimate sink and depository for microplastics. Microplastics from land and ocean sources may enter the marine environments and transfer through the food chains. Through biological interactions (e.g., marine snow), microplastics increase density and sink to deeper layers. Once entering the hadal zone, physical processes facilitate microplastics to deposit in the hadal trenches and bioavailable for benthic organisms.

### 248 **3.3 Sources and sinks**

We propose cascading processes that transport microplastics on their journey from land and oceanic gyres through intermediate waters to the deepest corners of the ocean (Figure 4), making hadal trenches the ultimate sink for plastic pollution. The most significant source of all sizes of plastic debris is the input from land-based sources, especially river discharge, and concentration

253 of microplastics positively correlates with the amount of mismanaged plastic waste in river 254 catchments (Lebreton et al., 2017). Papua New Guinea in the West Pacific, a region regarded as 255 major plastic waste input into the ocean in the world (Jambeck et al., 2015), was more populated 256 in recent years, putting pressure on the waste treatment facilities in growing informal settlements 257 in coastal cities (Smith, 2012). This may explain the higher abundance of microplastics found in 258 New Britain Trench in this study. A recently identified source of marine microplastics is 259 atmospheric fallout (Dris et al., 2016). Through wind transfer, airborne microfibers may account 260 for 7% of marine microplastic pollution (Boucher and Friot, 2017). The heavily populated NW 261 Pacific and the rapidly expanding Great Pacific Garbage Patch (GPGP) may contribute to the 262 quantity of microplastics at the ocean surface (Moore et al., 2001; Lebreton et al., 2018). 263 Ocean-based sources, including maritime activities, aquaculture and fishery, still contribute to a 264 great portion of plastic marine debris, although dumping at sea has been banned by MARPOL 265 Annex V since 1988 (Borrelle et al., 2017).

266 Although an increasing amount of plastics was found accumulating at the surface (Lebreton et 267 al., 2018), the eventual fate of microplastics, however, still lacks investigation. The mechanisms 268 that assist vertical transport of microplastics from the surface to the deep ocean has only been 269 investigated under lab conditions (Long et al., 2015) or in the shallow seas (Katija et al., 2017). It 270 is likely the high concentrations of microplastics in this study indicate that the deep ocean 271 (Woodall et al., 2014), especially the hadal trenches (Jamieson et al., 2019), may be a significant 272 resting ground for microplastics pouring into the ocean. But how do they get there? Direct 273 sinking of microplastic particles alone may take hundreds or thousands of years to reach the 274 hadal trenches. The presence of microplastics in the hadal zone requires there to be mechanisms 275 that promote the rapid descent from surface to full ocean depth. It cannot be gravitational sinking

276 - spherical microplastics may spend 10-15 years in the euphotic zone before sinking, while for 277 fibers it is 6-8 months (Chubarenko et al., 2016). Biofouling will increase the density of 278 microplastics, leading to loss of buoyancy and vertical transportation to the seafloor 279 (Chubarenko et al., 2016). Marine snow, or particulate organic matter (POM) synthesized in the 280 euphotic zone, is the main food supply of life in hadal environments (Long et al., 2015; Taylor et 281 al., 2016), and an important component of biological pump (Miyake et al., 2011). As 282 microplastics are detected in marine snow (Zhao et al., 2017), the pathways and fluxes of marine 283 snow from ocean surface to the deeper waters can predict possible mechanisms of microplastic 284 occurrence even in the deepest hadal trenches (Ichino et al., 2015). Based on the equations from 285 laboratory tests (Long et al., 2015), we calculated the time for microplastics incorporated into 286 marine aggregates to reach the hadal sediments in our study (Figure S2). A minimum of 14 days 287 is enough for a polypropylene fragment to reach 4900 m if aggregated in phytoplankton 288 aggregates. Once marine snow is removed to the deep, its quantity and quality is reduced by 289 large extent (Jamieson et al., 2010). A video showcasing marine snow at 8930 m in the New 290 Britain Trench was recorded during sample collection (Movie S1). A possible mechanism for 291 microplastics to reach the seafloor may be the formation of "ecocorona", which increases the 292 density of microplastics (Galloway et al., 2017). The formation of ecocorona on the surface of 293 microplastics by macromolecules or microorganisms alters the size, hydrophobicity and other 294 chemical properties of microplastics (Rillig et al., 2017). This has been proven by the high 295 concentration of nitrogen (N) on microplastics collected from surface waters, as N is not a 296 content for consumer plastics, but crucial proxy for biomass (Morét-Ferguson et al., 2010). 297 Using SEM analysis, we found organic matter attached to the surface of microplastics (Figure 3).

298 Microplastic and plastic debris have been shown to be ingested and entangled by 395 marine 299 species (Gall and Thompson, 2015), from herbivores (e.g., zooplankton, Moore et al., 2001; Cole 300 et al., 2013; Frias et al., 2014; Hall et al., 2015; Sun et al., 2017; Frydkjær et al., 2017), primary 301 carnivores (e.g., bivalves, Taylor et al., 2016; Murray and Cowie, 2011; von Moos et al., 2012), 302 secondary carnivores (e.g., fish, Dantas et al., 2012; Bråte et al., 2016; Jabeen et al., 2017; 303 Anastasopoulou et al., 2013) to top carnivores (e.g., large shark, Lusher et al., 2015; Alomar and 304 Deudero, 2017). Transfer of microplastics in food webs had been proven in both field and lab 305 experiments, including the transfer within planktonic food web from mesozooplankton to 306 macrozooplankton (Setälä et al., 2014), natural transfer from mussels to crabs (Farrell and 307 Nelson, 2013), and from fish to top predators (Nelms et al., 2018). Microplastics were found in 308 stranded whales (Lusher et al., 2015), and whale fall could support significant food source in 309 deep-sea floor (Smith and Baco, 2003). Giant larvaceans transport microplastics and carbon into deep sea either by sinking of fecal pellets or discarded mucus houses at rates of 300 m day<sup>-1</sup> and 310 800 m day<sup>-1</sup>, respectively (Katija et al., 2017). Microplastic particles with increased density due 311 312 to biofouling will sink to deeper layers in the ocean. By transferring through marine food webs, 313 aggregating into marine snows, fecal pellets, or animal carcasses, microplastics are vertically 314 transported to the deeper layers.

But export from the surface and intermediate ocean does not explain fully why the microplastics are focused in the trenches. Microplastic abundance in different layers has distinct distribution patterns, with the surface layer and the deep water at higher concentrations (La Daana et al., 2018), indicating vertical change in microplastics in water columns. Trenches are well ventilated due to the overflow of dense water currents (Johnson, 1998), which brings in oxygen and nutrition (Ichino et al., 2015), as well as some pollutants (Jamieson et al., 2017). The 321 unique topography of trenches increases the likelihood of microplastic accumulation. Similar to 322 submarine canyons, funneling mechanism of trenches tend to accumulate organic matter and 323 other substances in the hadal ecosystem with little opportunity for dispersal. Steep slope of 324 trenches and occasional seismic activity accelerate the accumulation of substances along the 325 trench axis, or "trench resource accumulation depth" (Jamieson et al., 2010). The Atacama 326 trench at 7800 m functions as a deep oceanic trap, or a "depocentre", for organic matter, which 327 accelerates benthic microbial processes in hadal environment (Danovaro et al., 2003). 328 Microplastics that undergo the funneling effects will eventually sink to the bottom along the axis 329 of a trench. Compared to gravity-driven transport of marine snow, bottom currents play a greater 330 role in the transport of organic matter, and leads to higher biomass along the axis of a trench (Ichino et al., 2015). The maximum of current velocity at 10890 m can reach 8.1 cm s<sup>-1</sup> (Johnson, 331 332 1998), making the lateral transport of sediments, dissolved oxygen and other substances possible 333 within the trench. In this study, microplastics showed a relatively high abundance in surface 334 sediments in hadal trenches, which may reflect accumulation along the axis of hadal trenches, 335 thus more sampling efforts should be focused in this area.

#### 336 **3.4 Ecological effects**

Ecological theory for littoral zones is applicable for characteristics in trench communities, albeit the elevated hydrostatic pressure (Jamieson et al., 2010). The ecological effects of microplastics include acting as "Trojan horse" for hydrophobic organic chemicals (HOCs) in food webs (Diepens and Koelmans, 2018), and as carrier for foreign species (Miyake et al., 2011). Different accumulation patterns for biomagnification of PCBs and PAHs were revealed by feeding zooplankton, fish and top predators in Arctic food web with HOCs contaminated microplastics (Diepens and Koelmans, 2018). In six deepest trenches including the Mariana Trench, three species of amphipod were found to ingest microplastics, indicating microplastics are bioavailable in hadal ecosystems (Jamieson et al., 2019). With the discovery of POPs in hadal amphipod species (Jamieson et al., 2017), similar "Trojan horse" effect may apply to hadal food webs.

348 Once entering the hadal food webs, microplastics might be perpetually locked in the trophic 349 cycling due to both surface-derived carcasses and hadal-derived amphipod as food sources by 350 amphipods (Jamieson et al., 2010). At the deepest corner on earth, amphipods are hungry 351 necrophages, cannibals, detritivores and carnivores, which ingest nutrient-poor POM from above 352 (Jamieson et al., 2010). Dawson et al. (2018) found a new pathway for plastics to transfer in 353 biogeochemical cycling. Ingested microplastics may fragment into nanoparticles and pass 354 physical barriers of krill or other marine species. It is very likely that hadal species facilitate 355 microplastic fragmentation through ingestion and translocation, thus securing microplastics and 356 nanoplastics in the hadal zone, making hadal trenches the ultimate sink for plastics. However, 357 even comprehensive understanding of ecological impact of plastic pollution in coastal area is still 358 lacking. More research efforts are required for the risk assessment of microplastics in the hadal 359 zone to inform policy makers.

# 360 4 Conclusions

Microplastics found in sediments in the three deepest hadal trenches show that microplastics are already accumulating at the bottom of deepest corners on the planet, with an average abundance of 71.1 items per kg dry weight sediment (i.e., 89.6 items per L dry weight). Our study provides essential baseline data on microplastic pollution in hadal environments (down to 10890 m), which confirms that microplastics have reached full-ocean depth. The potential 366 cascading processes that transport microplastics to hadal trenches provides an answer to how 367 microplastics are transported from sea surface to seafloor, including land and ocean-based input 368 sources (e.g., maritime activities), ingestion by marine biota and trophic transfer, biological 369 interactions with microplastics (e.g., marine snow) and physical processes (e.g., deep water 370 currents) in the hadal trenches. These processes make the hadal trenches major depositories and 371 the ultimate sink for microplastics. The understanding of sinking mechanisms and ecological 372 effects of microplastics in the hadal zone will hopefully enhance the understanding for the 373 remediation and prevention measures of microplastic pollution for relevant stakeholders.

374

# 375 Supporting Information:

- 376 Additional results
- 377 Figure S1
- 378 Figure S2
- 379 Table S1 S2
- 380 Movie S1
- 381

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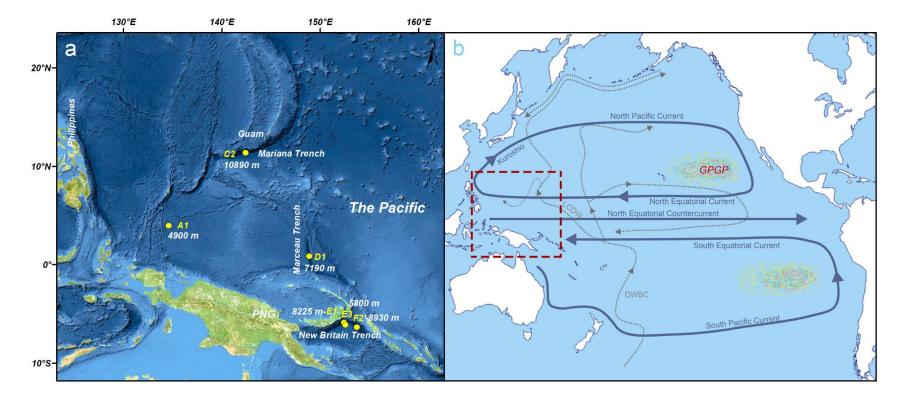
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Sampling	Location	Depth	Longitude	Latitude	Sampler	Sediment	Sand	Silt	Clay
site		[m]	[degrees_east]	[degrees_north]		Description	[%]	[%]	[%]
A1	Abyssal	4900	134.6021	3.9954	Lander	Brown, Soft	9.4	47.8	42.8
	plain east								
	of								
	Philippines								
E3	New	5800	152.5609	-6.1006	Box	Deep	7.7	58.0	34.3
	Britain				corer	Brown, Soft			
	Trench								
D1	Marceau	7190	148.8887	0.8975	Lander	Deep	2.7	51.5	45.8
	Trench					Brown, Soft			
E1	New	8225	152.4252	-5.8662	Lander	Deep	3.5	64.4	32.1
	Britain					Brown, Soft			
	Trench								
F2	New	8930	153.7430	-6.3258	Lander	Yellow,	9.9	74.3	25.8
	Britain					Soft, with			
	Trench					tephros			
C2	Challenger	10890	142.4041	11.3537	Lander	Deep	4.3	66.4	29.3
	Deep,					Brown, Soft			
	Mariana								
	Trench								

Table 1. Detailed information on geographical coordinates, sediment description, and grain size distribution of sediment samples from six sampling sites.

Sampli Locatio Dept Abunda Abundan Micropla Shape Color Size Polymer Polyme stic ID ng site n h nce ce [µm [Match %] r [items/kg ] Density [m] [items/L dry  $[g/cm^3]^3$ dry 8-40 weight] weig ht] A1 67.2 A1-1-1 115 PET 1.30-1.5 Abyssal 4900 53.3 Fragm Red (94.73) 0 plain ent A1-1-2 PP (82.10) 0.88-1.2 east of Fragm Transpar 606 Philippi 3 8 ent ent nes E3 New 5800 67.2 53.3 E3-2-2 Fiber Blue 176 Polyester 1.30-1.5 Britain 0 (96.49) 0 Trench E3-3-1 Fiber Red 289 Polyester 1.30-1.5 5 (91.83) 0 D1 7190 33.6 26.7 D1-2-1 Fiber Transpar 829 Polyester 1.30-1.5 Marceau Trench 8 (97.09) ent 0 E1 1.30-1.5 8225 168.0 133.3 E1-2-1 105 New Fragm Blue Polyester Britain (82.62) 0 ent 1.13-1.3 Trench E1-2-3 Fiber Transpar 120 Nylon 4 (98.38) 8 ent E1-3-2 1.30-1.5 Fiber 100 Transpar Polyester ent 9 (95.67) 0 E1-3-4 73 PET 1.30-1.5 Red Fragm (99.01) 0 ent E1-3-5 Film Transpar 491 Poly(octad 0.90 ecyl ent acrylate) (98.32) F2 New 8930 84.0 66.7 F2-1-3 Film Transpar 422 Polyolefin 0.86 Britain (85.51) ent Trench F2-1-4 Fiber Transpar 774 PP (97.25) 0.88-1.2 3 ent C2Challen 1089 117.6 93.3 C2-2-2 Fiber Transpar 105 Polyester 1.30-1.5 0 8 (83.40) 0 ger ent C2-2-5 1.30-1.5 Deep, Fiber 123 Polyester Transpar 0 Mariana 76 (98.30) ent Trench

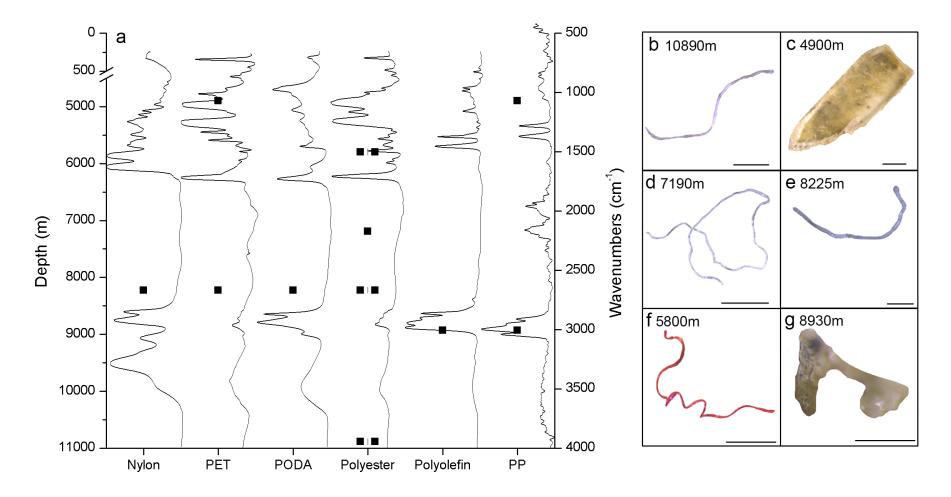
Table 2 A total of 14 identified microplastics (excluding rayon) in six sampling sites in abyssal plain and hadal trenches, illustrating the characteristics of microplastics.



**Figure 1.** Map of the six sampling sites and a schematic of ocean circulation in surface and deep water in the Pacific Ocean (modified from Talley et al., 2011). (a) Sampling sites are marked with yellow dots. C1: Challenger Deep, Mariana Trench; A1: north of Papua New Guinea (PNG), east of the Philippines, abyssal plain; D1: Marceau Trench; E1, E2 and F2 are located in the New Britain Trench. (b) Red dotted square

shows the location of (a). Surface circulation are marked by blue solid lines and abyssal circulation are marked by grey dotted lines. GPGP:

Great Pacific Garbage Patch; DWBC: Deep Western Boundary Current; LCDW: Lower Circumpolar Deep Water



**Figure 2.** Microplastics in sediments in the abyssal plain and hadal trenches, and imaged examples of microplastics from each site at various depth. (a) Distribution of microplastics categorized by polymer type (excluding rayon, marked as black squares) and infrared spectra of six kinds

of polymers identified using FT-IR instruments. (b) Fiber (ID: C2-2-2, polyester), (c) fragment (A1-1-2, PP), (d) fiber (D1-2-1, polyester), (e)

fiber (E1-2-3, nylon), (f) fiber (E3-3-1, polyester), and (g) film (F2-1-3, polyolefin).

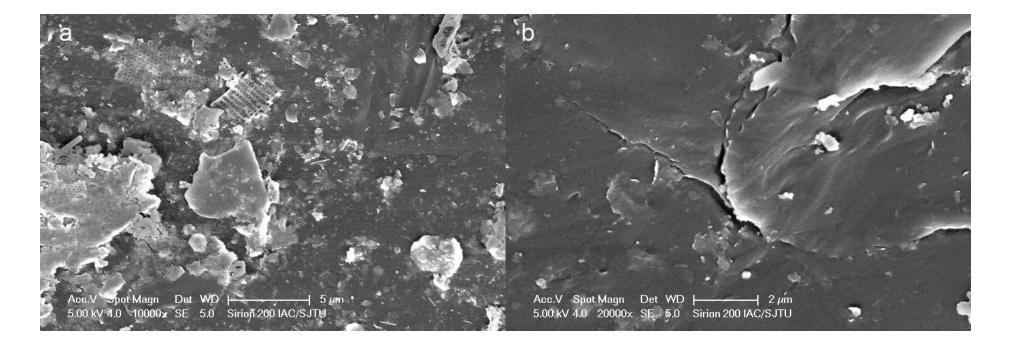
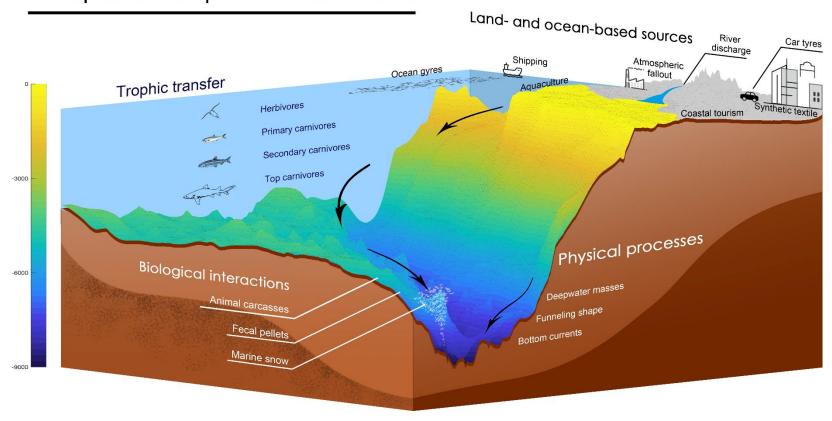


Figure 3. SEM images of the surface texture of one PP fragment (ID: A1-1-2). (a) showing organic matter attached to the surface of microplastic

particle with magnification of 10000×, and (b) showing signs of degradation on the surface with magnification of 20000×.



# Microplastics transport to hadal trenches

**Figure 4.** A schematic of pathways of microplastics from various sources entering the hadal trenches (taking the New Britain Trench as an example), making the deepest spots on Earth the ultimate sink and depository for microplastics. Microplastics from land and ocean sources may enter the marine environments and transfer through the food chains. Through biological interactions (e.g., marine snow), microplastics increase density and sink to deeper layers. Once entering the hadal zone, physical processes facilitate microplastics to deposit in the hadal trenches and bioavailable for benthic organisms.

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