



Monitoring photolysis and (solar photo)-Fenton of enrofloxacin by a methodology involving EEM-PARAFAC and bioassays: Role of pH and water matrix

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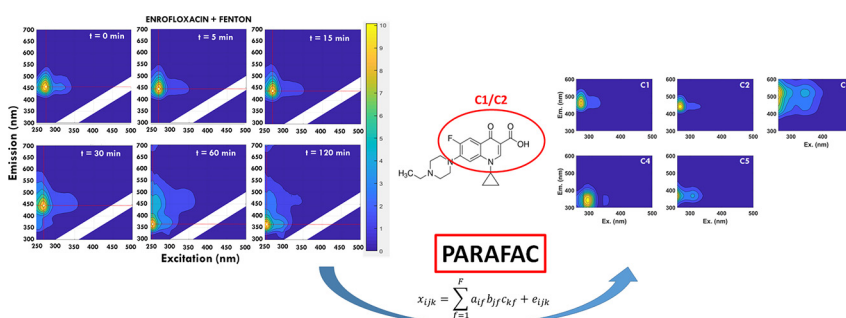
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HIGHLIGHTS

- Enrofloxacin degradation has been studied by photolysis and (photo)-Fenton.
- The effect of water constituents, pH and iron chelation has been determined.
- There is a relationship between antibacterial activity and fluoroquinolone core.
- EEM-PARAFAC is a simple and fast methodology to study a fluoroquinolone degradation.

GRAPHICAL ABSTRACT



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ABSTRACT

The degradation of enrofloxacin (ENR) by direct photolysis, Fenton and solar photo-Fenton processes has been studied in different water matrices, such as ultra-pure water (MQ), tap water (TW) and highly saline water (SW). Reactions have been conducted at initial pH 2.8 and 5.0. At pH = 2.8, HPLC analyses showed a fast removal of ENR by (solar photo)-Fenton treatments in all studied water matrices, whereas a 40% removal was observed after 120 min of photolysis. However, TOC measurements showed that only solar photo-Fenton was able to produce significant mineralization (80% after 120 min of treatment); differences between ENR removal and mineralization can be attributed to the release of important amounts of reaction by-products. Excitation-emission matrices (EEMs) combined with parallel factor analysis (PARAFAC) were employed to gain further insight into the nature of these by-products and their time-course profile, obtaining a 5-component model. EEM-PARAFAC results indicated that photolysis is not able to produce important changes in the fluoroquinolone core, in sharp contrast with (solar photo)-Fenton, where decrease of the components associated with fluoroquinolone core was observed. Agar diffusion tests employing *E. coli* and *S. aureus* showed that the antibacterial activity decreased in parallel with the destruction of the fluoroquinolone core.

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Abbreviations: ENR, enrofloxacin; FQs, fluoroquinolones; MQ, MilliQ water; TW, tap water; SW, saline water; EEMs, fluorescence excitation-emission matrices; PARAFAC, parallel factor analysis.

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1. Introduction

Fluoroquinolones (FQs) are broad-spectrum antibiotics extensively used in veterinary and human medicine (Ebert et al., 2011). In fact, according to B. Hamad (2010), FQs are the third-largest group of antibiotics, accounting for 17% of the global market share. These drugs might enter the environment through different sources such as pharmaceutical industries, hospital and urban wastewaters as well as fish or animal farms (Van Doorslaer et al., 2014). In addition to this, the amount of these antibiotics that are excreted unmetabolized can be estimated in a 70% (Van Doorslaer et al., 2014). In fact, these chemicals are commonly found in wastewater treatment plants (WWTPs) in concentrations that are usually in the ng/L to µg/L range, but can also be found at higher concentrations (up to 50 mg/L) in effluents of drug manufacturing plants (Rakshit et al., 2013). Conventional WWTPs are not able to remove efficiently this family of substances, as percentages of removal of 50–80% have been reported depending on the molecule (Van Doorslaer et al., 2014), thus constituting an additional route for their incorporation to natural aquatic ecosystems (Guinea et al., 2009). Besides, the FQ core is relatively stable to hydrolysis and thermal decomposition, providing these compounds of large environmental persistence (Babić et al., 2013).

One of the major concerns associated with the presence of antibiotics in the environment is the development of bacterial resistances. The European Food Security Agency has identified FQs among the substances which can generate this risk (ECDC/EFSA/EMA, 2017). Additionally, FQs or their decomposition by-products have shown toxicity towards aquatic organisms if they are not properly eliminated from water bodies (Aristilde et al., 2010; Ebert et al., 2011; Robinson et al., 2005).

In sharp contrast with the low efficiency of the chemical and biotic pathways for FQ removal, photolysis has been described as a major route for the self-remediation of ecosystems containing these substances. Important work has been devoted to study the photodegradation of FQ in different aqueous media (Ge et al., 2018; Zhang et al., 2019). Its photochemical reactivity is attributable to the presence of chromophores which can absorb wavelengths that are within the range of solar radiation reaching the Earth's surface ($300\text{ nm} < \lambda$) (Babić et al., 2013). Different mechanisms, namely direct and indirect photolysis as well as self-sensitized processes, have been described for FQs (Ge et al., 2010; Li et al., 2011).

Natural processes are not always strong enough to treat effluents polluted with antibiotics, but some photochemical advanced oxidation processes (AOPs) have been shown to be useful to treat FQ (Guinea et al., 2009; Feng et al., 2018; Feng et al., 2016; Van Doorslaer et al., 2013; Sun et al., 2019). AOPs are able to generate powerful transitory reactive species, mainly hydroxyl radicals ($\bullet\text{OH}$) (Glaze et al., 1987; Legrini et al., 1993), but other species can also be involved (An et al., 2010). In particular, Fenton process is able to produce reactive species, such as $\bullet\text{OH}$ and highly oxidized iron species, using iron salts as catalysts to improve the oxidizing ability of H_2O_2 , following a rather complex mechanism that is still to be completely elucidated (Pignatello et al., 2006). The photo-Fenton process consists in an enhancement of the Fenton reaction upon irradiation with UV or visible light, and hence sunlight can be employed with the economic and ecological advantages that they involve (Malato et al., 2009). Some examples are available in literature on the treatment of FQs by Fenton (Gupta and Garg, 2018; Villegas-Guzman et al., 2017) and photo-Fenton (Michael et al., 2013; Villegas-Guzman et al., 2017).

However, some research is still needed on the use of (photo)-Fenton to FQs removal. In particular, the effect of operational variables such as pH and or salinity has not been deeply evaluated. As FQs are commonly employed in marine fish farms (Aufartová et al., 2017; Buschmann et al., 2012; He et al., 2012; Robinson et al., 2005), the presence of high amounts of chlorides can drive undesired secondary reactions, decreasing the efficiency of the process (Pignatello, 1992; Buxton et al., 1988)

and/or leading to the formation of chlorinated by-products (Calza et al., 2005; Liu et al., 2009). Hence, further research is required to determine the extent of such reactions and to rule out possible undesirable side effects. On the other hand, exploring the performance of FQs degradation at mild pHs becomes of paramount relevance in real-case scenarios (Santos-Juanes et al., 2017). In addition to this, complete mineralization of FQs is a challenging goal to be reached (Gupta and Garg, 2018), since reaction by-products remain in the system, and their nature has to be unveiled and their biocompatibility assessed.

In this work, we report on the degradation of enrofloxacin (ENR), a common FQ developed to be employed in animals (Idowu and Peggins, 2004), also related to ciprofloxacin (a FQ used in human medicine) (Aresta et al., 2019; Ebert et al., 2011; García Ovando et al., 1999). Photolysis and (solar photo)-Fenton have been tested under simulated sunlight at different pHs (2.8 and 5.0) and using three aqueous matrices, namely Milli-Q (MQ), tap water (TW) and highly saline water (SW), following ENR removal by HPLC and mineralization by TOC analysis. However, these tools provide only partial information on the behaviour of the organic matter along the process and on the nature of the by-products that are released. Hence, implementation of a simple methodology to obtain major trends on by-products formation is needed. In this context, chemometric tools are gaining momentum, as they can extract valuable information from raw complex data (Alves and Poppi, 2013; Boughattas et al., 2019; Karoui and De Baeremaeker, 2007; Trawiński and Skibiński, 2018). In this sense, and taking advantage of the photophysical properties of FQ-type molecules, fluorescence excitation-emission matrices (EEMs) with the multi-way method, parallel factor analysis (PARAFAC), have been selected to study the ENR degradation. EEM-PARAFAC is based on the decomposition of the measured EEMs into their underlying chemical components, which can be associated with families of compounds bearing similar chromophores (García et al., 2019). Due to this reason, it has been mainly employed in the organic fraction characterization of complex water samples containing dissolved organic matter (Cory and McKnight, 2005; Stedmon et al., 2003; Tang et al., 2020) and also to monitor its changes (Li et al., 2014; Ly et al., 2018). Although it cannot reach the detail of sophisticated HPLC-HRMS techniques, it is easy, relatively cheap and non-time consuming. As far as we know, this is the first time that a FQ degradation is followed in time-course experiments using EEM-PARAFAC as until now, only a few examples of application of this technique on FQs can be found in chemometrics (De la Peña et al., 2003; Vosough et al., 2015) or to describe chemical interactions of these substances (Ni et al., 2009). In addition, there are very few examples on the application of this technique to gain further insight into the efficiency of advanced oxidation processes (Carabajal et al., 2017; García et al., 2019; García-Ballesteros et al., 2016).

Finally, bioassays based on the inhibition halo of *E. coli* and *S. aureus* have been used to estimate the antibacterial activity of the effluents derived from the different treatments, relating the results with all the above mentioned methodologies.

2. Experimental section

2.1. Reagents

High purity ENR (>99%), purchased from Sigma-Aldrich, was used in all the experiments. Na_2SO_3 anhydride, $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$, H_2O_2 (33% w/v), ascorbic acid, 1,10-phenantroline 1-hydrate, NH_4VO_3 , NaCl, H_2SO_4 96%, and UHPLC grade methanol and acetonitrile were obtained from AppliChem-Panreac. 80% formic acid was purchased from VWR Chemicals. Ultra-pure water was prepared with a Millipore system (MQ). Catalase from bovine liver (lyophilized powder 2000–5000 units/mg protein) was purchased from Sigma-Aldrich. Tap water employed in the experiments was obtained from the urban supply network of Alcoy (Spain) and had the following characteristics: pH = 7.4, inorganic carbon = 52 mg/L, conductivity = 510 µS/cm, $[\text{Cl}^-]$ = 27.8 mg/L,

$[\text{SO}_4^{2-}] = 33.3 \text{ mg/L}$, water hardness = 142 mg/L of CaCO_3 . Saline water was prepared by dissolving 30 g of NaCl in 1 L of tap water.

Agar diffusion tests were made using *Escherichia coli* CECT 101 strain for MQ and TW assays; because of the incompatibility of *E. coli* with highly saline media, the halotolerant *Staphylococcus aureus* CECT 435 for the SW ones. Tryptone soya broth and Mueller-Hinton broth were both provided by Scharlau.

2.2. Reactions

Reactions were carried out in cylindrical open glass reactors (total volume of 500 mL). For each experiment, the reactor was loaded with 250 mL of ENR solution (initial concentration of 25 mg/L), also containing 5 mg/L of Fe(II) (using $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ as the source of iron), prepared with the desired water matrix, MQ, TW or SW. Initial pH was adjusted to 2.8 or 5.0 by dropwise addition of H_2SO_4 0.5 M and/or NaOH 1 M. For the (solar photo)-Fenton experiments, the stoichiometric amount of H_2O_2 to mineralize ENR was added (125 mg/L); this is a useful procedure commonly employed to normalize the amount of hydrogen peroxide added to the reaction medium and to ensure that the process is not stopped because of the exhaustion of hydrogen peroxide (Bernabeu et al., 2011; Gomis et al., 2015). When necessary, irradiations were performed with a solar simulator (Oriol Instrument) equipped with a high-pressure Xe short arc lamp (Ushio UXL-302-0).

Experiments were run for 120 min and samples were taken periodically and processed differently depending on the type of analysis. In the case of HPLC and EEMs measurements, 5 mL of each sample were taken from the solution, and 50 μL of methanol were added to quench the excess of hydrogen peroxide, thus stopping the reaction (Gomis et al., 2014; Pignatello et al., 2006). Then 50 μL of H_2SO_4 5 M were added to each sample in order to ensure the acidic pH, due to the pH dependence of the absorption and fluorescence spectra of ENR (Lizondo, 1997). For TOC analysis, Na_2SO_3 was used instead of methanol to deal with the excess of H_2O_2 in order to avoid interferences caused by the addition of extra organic substances. Finally, in the case of the bioassays, samples were previously adjusted to pH 7.0 (± 0.5) with NaOH 1 M, and the remaining hydrogen peroxide was consumed by addition of catalase enzyme.

Dark controls were performed with and without iron observing no changes in the concentration of ENR. Controls with H_2O_2 without iron, and with and without irradiation, were also performed. H_2O_2 in the dark was not able to degrade ENR.

2.3. Chemical analysis

ENR concentration was determined using HPLC. The equipment consisted of a Hitachi Chromaster, equipped with a pump model 5110, a UV/Vis detector (model 5410), and oven (model 5310) and an autosampler (5210 model). A C18 Machery-Nagel column Nucleodur- π^2 5 μm was used as stationary phase and an isocratic flow of 0.25 mL/min of formic acid 0.1 M and acetonitrile (80:20) was used as eluent. The oven was set at 40 $^\circ\text{C}$ and the detection was performed at 275 nm. Mineralization analyses were performed in a Shimadzu TOC-V equipment with ASI-V autosampler.

EEMs were measured with a fluorimeter Photon Technology International (PTI) QuantaMaster 400, equipped with a Xe arc lamp. The explored excitation range was 250–500 nm (at 5 nm intervals), whereas the emission range was 300–700 nm (also at 5 nm intervals). A Hitachi-UH5300 spectrophotometer was employed to correct the inner filter effect (250–700 nm). The same spectrophotometer was used to measure the dissolved total iron concentration according to the ISO 6332:1988 standardized method and also to monitor H_2O_2 following a modification of the metavanadate method reported by Nogueira et al. (2005).

For the agar diffusion tests, bacteria (*E. coli* for MQ and TW, and *S. aureus* for SW) were previously grown up in a nutrient media for

18 h. Then, colonies were inoculated in a Tryptone soya broth, having a suspension of 0.5 units in McFarland scale. A sterile cotton swab was used to pick bacterial suspension and spread it on the surface of the Mueller-Hinton agar in the Petri dish. Afterwards, 0.85 cm wells were made and filled with 100 μL of the sample. Finally, the Petri dishes were incubated at 37 $^\circ\text{C}$ for 24 h, and then the inhibition halos were read.

2.4. Mathematical calculations

PARAFAC was applied in order to analyze the underlying signals present in the obtained EEM. For this purpose, 110 EEMs, obtained from sampling ENR solutions after different periods of treatment (described in Section 2.2) were recorded. PARAFAC was conducted using MATLAB 2018b with the graphical user interface EEMlab (Micó et al., 2019), which employs the routines of drEEM toolbox for the pre-treatment and analysis of the data (Murphy et al., 2013).

Briefly, the procedure, which is explained in detail in the works of Bro (1997) and Bro and Kiers (2003), consists in the following key steps: EEMs were first normalized by water Raman first order scattering signal (350 nm excitation wavelength and 365–450 nm for emission) in order to reduce error of the power lamp decay over time due to usage. Inner-filter effects were handled by uploading the absorbance spectra of each sample. MQ, TW and SW in the presence of 50 μL of methanol and 50 μL of 5 M H_2SO_4 solution were also recorded to perform the corresponding blank-subtractions to each sample. Regions affected by Rayleigh and Raman scattering were removed and data in those domains were estimated by interpolation from either sides of the bands. The correct number of components was assessed according to the chemical consistency of the obtained data, the evaluation of the distribution of the residuals and the core consistency diagnostic (CCD).

3. Results and discussion

3.1. ENR removals in different media

The degradation of ENR was studied under photolysis, Fenton and solar photo-Fenton in MQ water at pH 2.8 and 5.0. Plots of the normalized concentration of ENR vs. time are presented in Fig. 1. It can be observed that, at the same pH, ENR removal was faster in photo-Fenton, followed by Fenton and finally photolysis. The pH effect was not important for photolysis, but (photo)-Fenton was more efficient at pH = 2.8 than 5.0. This should be attributed to precipitation of iron (III) at pH above 3 by formation of hydroxides that are not able to participate in the Fenton process (Pignatello et al., 2006). However, it is remarkable that ENR removal was still fast at pH = 5.0 by means of solar photo-Fenton (complete elimination of 25 mg/L of ENR in <60 min). In fact,

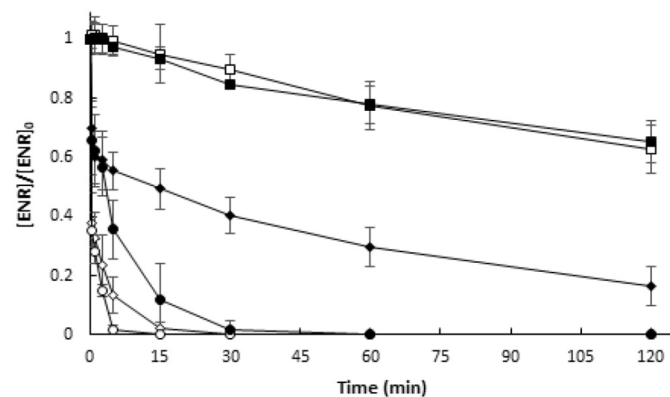


Fig. 1. ENR degradation profiles in Milli-Q water: photolysis at pH = 5 (■); photolysis at pH = 2.8 (□); Fenton at pH = 5 (◆); Fenton at pH = 2.8 (◇), photo-Fenton at pH = 5 (●) and photo-Fenton at pH = 2.8 (○).

complexation of iron is one of the strategies followed to keep iron active at mild pH (Santos-Juanes et al., 2017; Pignatello et al., 2006; De Luca et al., 2014). This effect has been observed in some compounds such as phenols, where complexation of iron by these substances shifted the optimum pH to values close to 4 (García-Ballesteros et al., 2016). Indeed, formation of complexes between FQs and iron has been observed and described in literature (Cuprys et al., 2018; Efthimiadou et al., 2008; Turel, 2002) and their role in Fenton and related processes has been reported (Chen et al., 2017; Giri and Golder, 2019). Hence, it can be proposed that ENR forms photoactive complexes with iron to explain the good efficiency of photo-Fenton at pH = 5.0. An indirect evidence is that the amount of iron remaining in the solution after photolysis remained nearly constant at 5 mg/L because of the formation of the ENR-Fe complex, in sharp contrast with the photo-Fenton experiment, where a continuous decrease in iron concentration was observed, in agreement with the destruction of ENR (in the absence of the complexant, precipitation of iron occurs).

Water matrix effect in the ENR degradation was also studied, performing the experiments also in TW and SW. Obtained removal percentages in these conditions at both studied pHs, 2.8 and 5.0, for direct photolysis presented no significant differences with the ones obtained in MQ, being systematically in the range 20–40% after 120 min; therefore, water constituents do not have big influence regarding direct irradiation of the compound. On the contrary, a matrix effect can be observed for Fenton and photo-Fenton, as shown in Fig. 2a and b. In those cases the fastest ENR removals were obtained in SW followed by TW and then in MQ. This primary disappearance of the parent product could be attributed to some reactivity of chlorinated radicals. As a matter of fact, in solutions containing high concentrations of chlorides, Cl^\bullet and $\text{Cl}_2^{\bullet-}$ can play an important role, as they can be formed either by direct reaction between $\bullet\text{OH}$ and Cl^- , or by photolysis of iron-chloride complexes, such as FeCl^+ , FeCl_2^+ and FeCl_2^+ (De Laat and Le, 2006; Machulek Amilcar et al., 2007; Soler et al., 2009). This is in agreement

with the lower H_2O_2 consumption observed in SW in comparison with the other two water matrices (see Fig. S1). As a matter of fact, higher consumptions have been detected at pH = 2.8 than at 5.0, in agreement with the better performance of photo-Fenton at former conditions.

3.2. Fluorescence spectroscopy analysis

The removal of ENR should not be related straightforward with a detoxification of the solution, as other products (even more toxic than parent compounds) might be released into the solution. To evaluate the amount of organics in the solution, total organic carbon (TOC) was determined after 120 min of solar photo-Fenton in all three matrices at pH = 2.8 and 5.0. Interestingly, higher mineralization was reached for solar photo-Fenton at pH = 2.8 for both MQ and TW with values of ca. 80%, followed by the same matrices at pH = 5.0 (mineralization slightly above 50%), whereas for SW, mineralization was below 20% at both pHs. Finally, in the case of the control of H_2O_2 with irradiation in MQ water, the observed ENR removal did not result in any mineralization.

The next step is gaining further insight into the nature of the organic matter present in the solution. However, for such complex samples conventional chromatography cannot be applied and gross parameters such as total organic carbon or chemical oxygen demand provide scarce information. For this reason, EEMs were measured, as they can give the fingerprints of major chromophores present in the reaction mixture. Fig. 3 shows the EEMs recorded for the photolysis, Fenton and solar photo-Fenton of ENR in MQ at pH = 2.8. It can be observed that photolysis was not able to produce significant changes in EEMs, which could be attributed to the persistence of the FQ core. On the contrary, changes in the EEMs can be observed for (photo)-Fenton, being faster for photo-Fenton than for Fenton. These observations might indicate that those processes are able to destroy the FQ moiety.

In order to gain further insight into the behaviour of the FQs along the process, PARAFAC was applied to the data set containing all the measured EEMs obtained in this work, namely photolysis, Fenton and photo-Fenton, in all three water matrices and both pH values (see Section 2.4 for details). A model consisting in 5 components was chosen, as in comparison with the models with fewer components, it was the only one able to resolve all the overlapped fluorescence signals appearing in the measured EEMs and, on the other hand, showed a higher chemical consistency when compared to those considering more components (see Fig. S2), that shows the CCD behaviour vs. the number of modelled components, were an abrupt change in the tendency of the curve was observed beyond the 5-model (Bro and Kiers, 2003).

In Fig. 4 the fingerprints for the 5-component model are shown. C1 can be related with the parent pollutant, ENR, although other by-products containing the FQ core can contribute. C2 presents very similar fingerprint as C1, but with a small shift in the maximum position to lower values in the emission axis; thus C2 is also supposed to belong to FQ type molecules. On the other hand, C3, C4 and C5 appear only in those experiments where the FQ structure is destroyed (Fenton and solar photo-Fenton, at pH = 2.8 with MQ and TW), and hence, they should be associated to different families of by-products generated by the strong oxidation of FQ.

Certainly, one of the most important subjects to be addressed is the removal of the FQ moiety, since the antibiotic properties could be associated to this functional group. Hence, the remaining amount of FQ might be considered as a preferred method to assess the efficiency of the photochemical process than just the disappearance of ENR followed by HPLC. Fig. 5 shows the plot of C1 + C2 vs. time; as C1 and C2 are assigned to compounds with the FQ moiety, the sum of their scores could be associated with the remaining amount of FQ in the medium.

Comparison of Figs. 1 and 5 shows that, in general, similar trends can be found between C1 + C2 and ENR relative amount shown by HPLC: a) No destruction of the FQ in photolysis experiments, in line with the

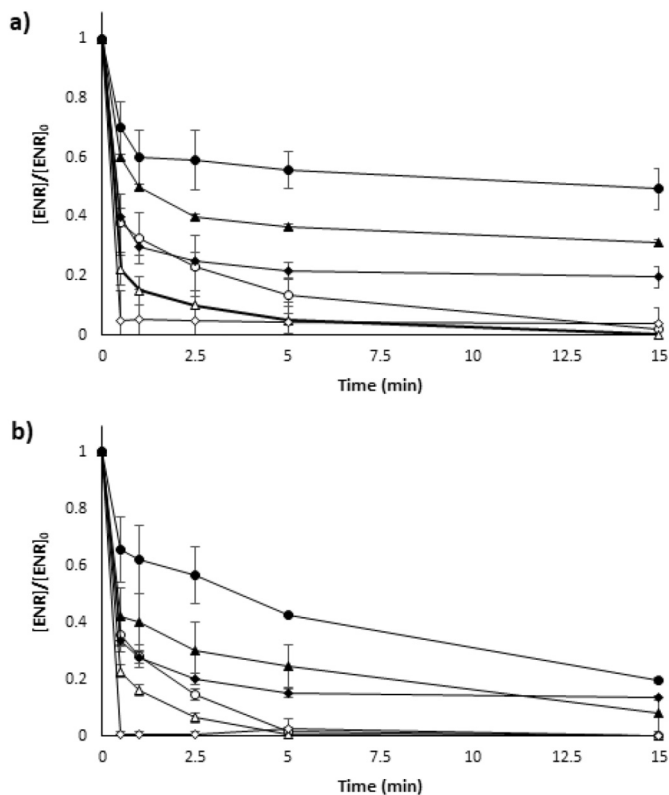


Fig. 2. ENR removal in different aqueous media for a) Fenton, b) solar photo-Fenton. SW at pH = 5 (◆); SW at pH = 2.8 (◇); TW at pH = 5 (▲); TW at pH = 2.8 (△); MQ at pH = 5 (●) and MQ at pH = 2.8 (○).

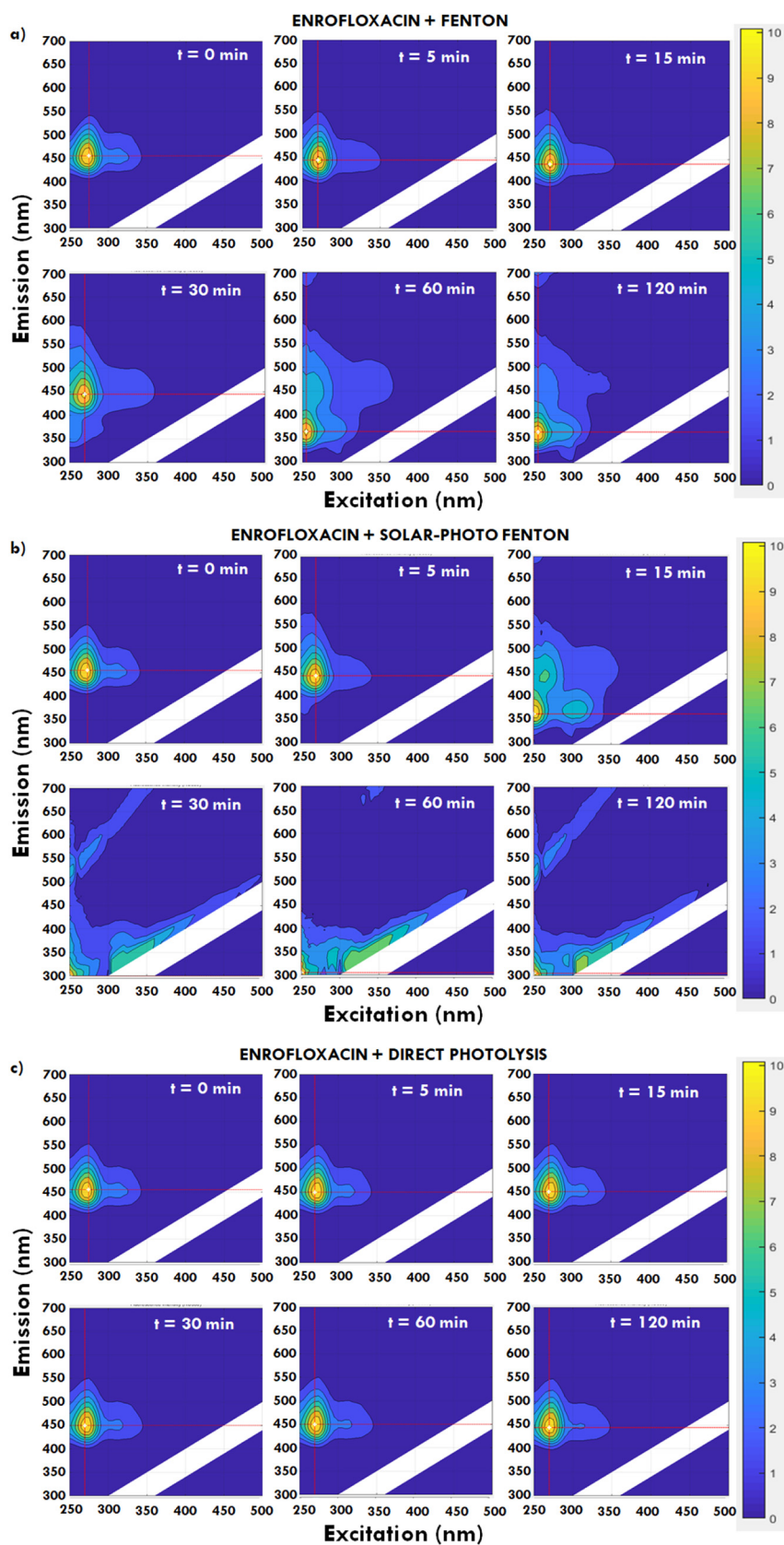


Fig. 3. Normalized fluorescence excitation-emission matrices obtained at different stages of ENR degradation in MQ at initial pH = 2.8 (0, 5, 15, 30, 60 and 120 min) by: a) Fenton, b) solar photo-Fenton and c) direct photolysis.

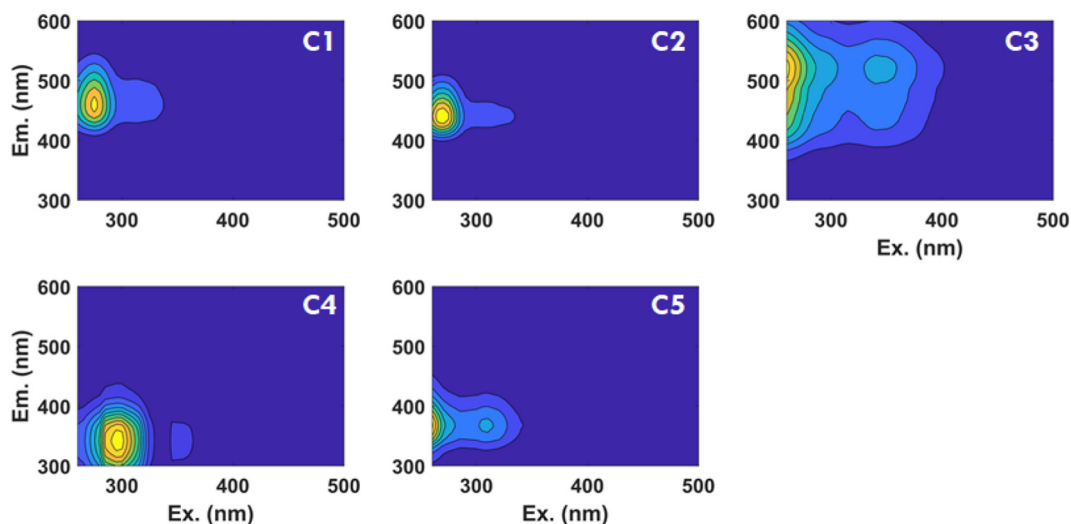


Fig. 4. Fingerprints of the 5 components obtained after the mathematical treatment by PARAFAC of a set of 110 EEMs obtained along photolysis, Fenton and solar photo-Fenton in all three water matrices and at pH 2.8 and 5.0.

low removal of the parent pollutant; b) some decrease in the C1 + C2 components was reached in the other cases following the trend: Fenton at pH = 5.0 < photo-Fenton at pH = 5.0 = Fenton at pH = 2.8 < photo-Fenton at pH = 2.8. This behaviour can be understood by considering that although photolysis is able to remove ENR, it cannot produce important changes in the fluoroquinolone structure (Babić et al., 2013; Ge et al., 2018; Zhang et al., 2019) and only stronger oxidation agents (e.g. $\cdot\text{OH}$) efficiently attack the fluoroquinolone core (An et al., 2010).

It is also interesting to test according to this parameter the effect of the water matrix. Results obtained for photo-Fenton can be observed in Fig. 6. A comparison of Figs. 2 and 6 shows significant differences, involving that the removal of ENR might not involve the elimination of the FQ moiety. Reactions carried out in SW resulted in a very fast removal of ENR but the presence of C1 and C2 after 120 min, indicates that the reaction resulted in the release of by-products still having the FQ structure. On the contrary, in MQ and TW, important changes in the structure of the molecule have occurred and all the fluoroquinolonic intermediates were removed after 15 min at pH = 2.8 and at 30 min at pH = 5.0. This is in line with TOC measurements (where the lowest mineralization percentage was obtained in the case of SW at both studied pHs) and can be explained if (photo)-Fenton in the presence of high concentration of chlorides results in the formation of less reactive species, that can destroy ENR but not the FQ core.

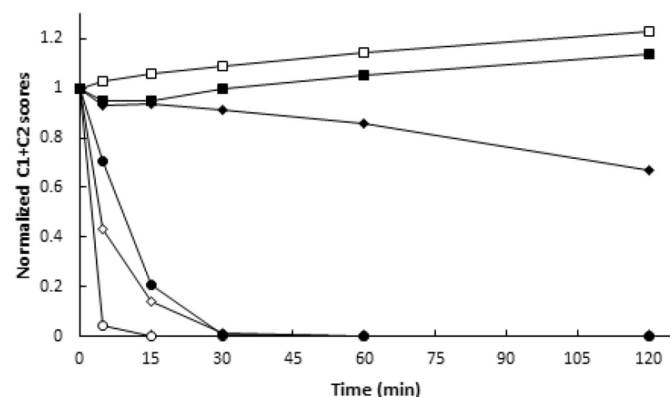


Fig. 5. Plot of the fluoroquinolonic components obtained in the EEM-PARAFAC model (C1 and C2) vs. time in MQ water. Photolysis at pH = 5 (■); photolysis at pH = 2.8 (□); Fenton at pH = 5 (◆); Fenton at pH = 2.8 (◇), photo-Fenton at pH = 5 (●) and photo-Fenton at pH = 2.8 (○).

3.3. Agar diffusion tests

In order to check the residual antibiotic properties of the FQ treated solution, inhibition halo tests were performed. In line with expectations, these bioassays showed an almost identical behaviour as the fluoroquinolonic compounds, as shown by the C1 + C2 components decay in EEM-PARAFAC. Fig. 7 shows results in MQ water for photolysis and (photo)-Fenton at pH = 2.8 and 5.0. The inhibition halo disappeared after 15 and 30 min for solar photo-Fenton and Fenton processes respectively in MQ at pH 2.8, whereas in photolysis the inhibition halo remained constant even after 120 min of irradiation and not notable decrease of inhibition halos for Fenton performed at pH 5.0 was seen. This might indicate that the destruction of the FQ structure is needed to eliminate the antibiotic properties (Ge et al., 2018, 2015; Rosen et al., 1989).

The same trend can be found in the photo-Fenton treatment under different experimental conditions (Fig. 8). The decay in the antibacterial activity is in agreement with the data obtained with the EEM-PARAFAC analysis of the FQ components: the degradation was faster at pH = 2.8 than at pH = 5.0 and the process driven at MQ water was more efficient than when salts were present. Finally, to verify if the possible chlorinated by-products formed in SW matrices were even more toxic than the original ENR, additional *Aliivibrio fischeri* experiments were

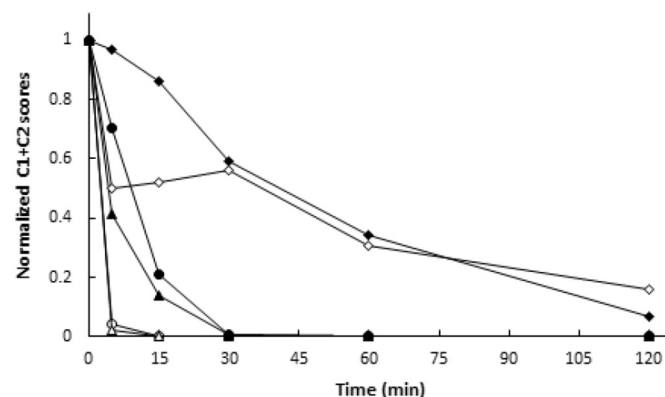


Fig. 6. Elimination of fluoroquinolonic components (C1 and C2) obtained in the EEM-PARAFAC model vs. time with solar photo-Fenton at different water matrices. SW at pH = 5 (◆); SW at pH = 2.8 (◇); TW at pH = 5 (▲); TW at pH = 2.8 (△); MQ at pH = 5 (●) and MQ at pH = 2.8 (○).

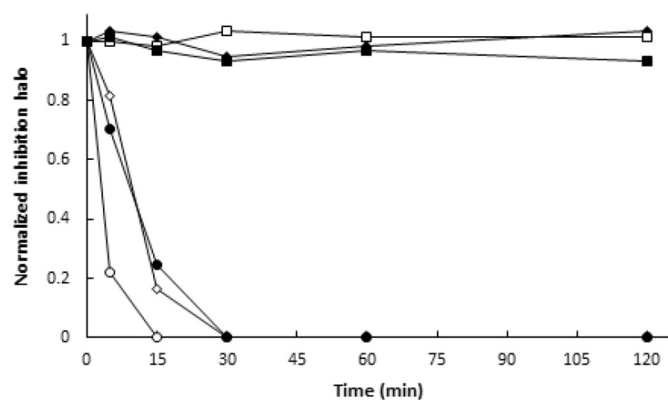


Fig. 7. Antibacterial activity behaviour obtained with *E. coli* for the different tested processes in MQ at both studied pHs. Photolysis at pH = 5 (■); photolysis at pH = 2.8 (□); Fenton at pH = 5 (◆); Fenton at pH = 2.8 (◇), photo-Fenton at pH = 5 (●) and photo-Fenton at pH = 2.8 (○).

performed, showing also a decay of toxicity with the reaction time (approximately 50% decrease in the toxicity measured according this assay after 120 min of photochemical process).

4. Conclusions

Solar photo-Fenton process resulted to be more efficient than photolysis and Fenton (according to ENR removal, mineralization percentage and inhibition halo decrease). Regarding the effect of the initial pH, higher removals were obtained at pH 2.8 rather than at pH 5.0. However, the good performance of solar photo-Fenton at pH 5, probably due to iron complexation, requires further research. Similar performances have been observed in MQ and TW, with fast total removal and high mineralization achieved in both cases. On the contrary, in SW, the fast ENR removal did not result in concomitant mineralization or destruction of the FQ core.

According to bioassays and EEM-PARAFAC results, photolysis of ENR does not result in destruction of the FQ core, therefore keeping its antibacterial activity. In contrast, (solar photo)-Fenton processes are able to destroy the FQ core, what results in a decrease of the inhibition halo.

Finally, EEM-PARAFAC study has demonstrated to be a simple methodology to provide supplementary information on the behaviour of organic matter and can be more easily correlated than HPLC analysis with changes in toxicity, and could be an alternative to sophisticated analytical techniques when identification of reaction by-products is not required.

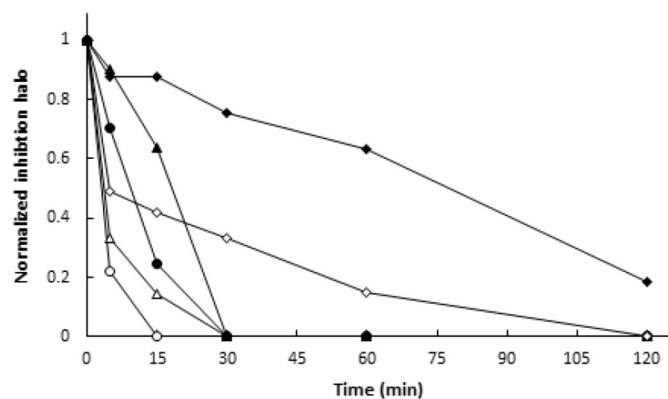


Fig. 8. Antibacterial activity behaviour obtained with solar photo-Fenton within the different tested water matrices. SW at pH = 5 (◆); SW at pH = 2.8 (◇); TW at pH = 5 (▲); TW at pH = 2.8 (△); MQ at pH = 5 (●) and MQ at pH = 2.8 (○).

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.scitotenv.2020.137331>.

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