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1 Can benthic foraminifera serve as proxies for changes in benthic macrofaunal community  
2 structure? Implications for the definition of reference conditions

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24 macrofauna – Community structure - Indicator species – Fjords

25

26 **Abstract**

27 Benthic macrofauna is one of the most widely used biological groups to assess the ecological  
28 status of marine systems. Lately, attention has been paid to similar use of benthic  
29 foraminifera. In this study, distribution patterns of benthic foraminiferal and macrofaunal  
30 species were investigated simultaneously in 11 fjords in southeastern Norway in order to  
31 assess correlations and responses to environmental conditions. Selected fjords allowed to  
32 investigate contrasted environmental conditions from low total organic carbon (TOC)  
33 content (sediment TOC < 2.7%) in normoxia (bottom-water [O<sub>2</sub>] > 2 mL O<sub>2</sub>.L<sup>-1</sup>) up to high  
34 TOC content (> 3.4%) in severe hypoxia (< 0.5 mL O<sub>2</sub>.L<sup>-1</sup>). Environmental parameters  
35 comprised bottom-water dissolved oxygen, grain size, total organic carbon, total nitrogen  
36 (TN), pigments and depth below threshold (DBT). Foraminiferal and macrofaunal community  
37 data were significantly correlated (Procrustes analysis  $m^2 = 0.66$ ,  $p = 0.001$ ). Hence, benthic  
38 foraminiferal distribution patterns mirror those of benthic macrofauna. However, as  
39 opposed to the foraminifera, macrofauna was not recorded at the most oxygen-depleted  
40 stations and, hence, was more sensitive to severe oxygen depletion. With regard to  
41 assigning species to ecological groups for ecological quality status assessment, the results  
42 suggest that species, *e.g.* *Spiroplectammina biformis* (foraminifera), *Scalibregma inflatum*  
43 (macrofauna), may exhibit different ecological requirements depending on their habitat.  
44 Considering the observed congruent patterns of benthic foraminifera and macrofauna,  
45 palaeo-communities of benthic foraminifera could be used as indicators of reference

46 conditions for benthic macrofaunal community structure. This would however need further  
47 developments of algorithms to perform such a translation.

## 48 **1. Introduction**

49

50 Most benthic macro-invertebrate species are sedentary, making them dependent on  
51 the local environmental conditions. Living at the sediment-water interface, they integrate  
52 aquatic as well as sediment conditions. Their communities show dramatic changes in their  
53 composition in response to organic matter loads (Pearson & Rosenberg 1978, Bouchet &  
54 Sauriau 2008), oxygen depletion (Rosenberg et al. 2002), oil spills (Gray et al. 1990), sewage  
55 (Glémarec & Hily 1981), heavy metals (Olsgard & Gray 1995) and physical disturbance  
56 (Lavesque et al. 2009). Tolerant species thrive at the early stages of the perturbation,  
57 followed by highly tolerant opportunistic species when the perturbation becomes severe  
58 (Glémarec & Hily 1981). Hence, macrofauna are routinely used in environmental bio-  
59 monitoring studies (e.g. Warwick 1986, Grall & Glémarec 1997, Borja et al. 2003, Bouchet &  
60 Sauriau 2008). In response to the implementation of marine legislation such as the Water  
61 Framework Directive (WFD), the Clean Water Act, and the Marine Strategy Framework  
62 Directive (MSFD), numerous indices based on macrofauna have been established for  
63 assessing the ecological quality status (EcoQ, see review in Pinto et al. 2009). Most of the  
64 indices developed to assess the EcoQ of marine systems are based on the indicative value of  
65 macrofaunal species. They have been classified into 5 ecological groups of sensitivity to  
66 disturbances from sensitive to first-order opportunistic species (Glémarec & Hily 1981, Borja  
67 2000). Lately, concerns arose about these indices (see review in Spilmont 2013). One of the  
68 main concerns is the definition of reliable reference conditions for benthic macrofaunal  
69 communities. This is a crucial point since in the European Water Framework Directive  
70 (WFD), reference conditions are mandatory to assess the EcoQ. According to the WFD, the  
71 reference conditions (also called “high” status) are defined as “for any surface water body

72 type reference conditions or high ecological status is a state in the present or in the past  
73 where there are no, or only very minor, changes to the values of the hydromorphological,  
74 physico-chemical, and biological quality elements which would be found in the absence of  
75 anthropogenic disturbance” (Common implementation strategy for the Water Framework  
76 Directive (2000/60/EC), Guidance Document No.5, transitional and coastal waters –  
77 typology, reference conditions and classification systems p. 106). Assessment of  
78 environmental quality is based on the extent of deviation from these reference conditions,  
79 following the definitions in the WFD ([www.ec.europa.eu/environment/water/marine.htm](http://www.ec.europa.eu/environment/water/marine.htm)).  
80 Most of the coastal areas are modified or have been modified by human activities (Diaz and  
81 Rosenberg 2008). Hence, EcoQ assessment within the WFD is facing issues to determine  
82 true reference condition (Nielsen et al. 2003, Elliott and Quintino 2007). As for benthic  
83 macrofauna, since there is almost no data from pre-impact time, reference conditions have  
84 to be determine using today’s communities, which is an issue considering the  
85 aforementioned degradation of coastal areas.

86         There is an increasing interest in the use of benthic foraminifera to characterize the  
87 health of marine systems (e.g. Alve 1995, Scott et al. 2001, Mojtahid et al. 2006, Bouchet et  
88 al. 2007, Frontalini et al. 2009). Lately, indices to use benthic foraminifera routinely to  
89 assess EcoQs have been proposed (Hallock et al. 2003, Alve et al. 2009, Bouchet et al. 2012,  
90 Barras et al. 2014, Dimiza et al. 2016). Specifically, benthic foraminiferal species were  
91 assigned to ecological groups (EGs) according to their sensitivity/tolerance to increasing  
92 organic matter enrichment (Alve et al. 2016, Jorissen et al. 2018), inspired by work on  
93 benthic macrofauna to develop the AMBI index (Glémarec & Hily 1981, Borja et al. 2000). It  
94 led to the development of the Foram-AMBI to assess the ecological quality status of marine  
95 systems. In these studies, the response of species along a gradient of enrichment in organic

96 matter allowed their classification into groups of different sensitivity. Five ecological groups  
97 were defined: sensitive (EG1), indifferent (EG2), tolerant (EG3), second-order opportunistic  
98 (EG4) and first-order (most) opportunistic (EG5). The AMBI and the Foram-AMBI are  
99 computed based on the proportion of the different ecological groups in the species  
100 community, allowing assessment of ecological quality status.

101           Benthic foraminifera leave an easily accessible and abundant fossil record, which  
102 allows reconstruction of the characteristics and timing of historical environmental variations  
103 (e.g. Alve, 1991; Hayward et al., 2004, Dolven et al. 2013, Francescangeli et al. 2016).  
104 Consequently, it is possible to trace the record of human-induced disturbance over decades  
105 or centuries. Indeed, in a pilot study, Alve et al. (2009) suggested that *in situ* reference  
106 conditions can be established using fossil benthic foraminiferal assemblages from dated  
107 sediment cores. For example, by comparing the “background” fossil foraminiferal  
108 assemblages to the modern living foraminiferal assemblages at the same site, it would be  
109 possible to determine if a site is naturally anoxic or has become anoxic with recent human  
110 influence. This kind of temporal, *in situ* monitoring is not possible with soft-bottom  
111 sediment macrofauna because they do not leave abundant or, for most species, any fossil  
112 records. Benthic foraminifera may thus provide a powerful tool for defining habitat-specific,  
113 *in situ* reference conditions for soft-bottom coastal and transitional waters. Although the  
114 response of macrofauna and foraminifera species to environmental gradients may partly  
115 differ (Mojtahid et al. 2008), the few studies comparing the two groups have indicated that  
116 their responses to environmental changes are basically similar (Schafer et al. 1975, Schafer  
117 et al. 1995, Klitgaard-Kristensen & Buhl-Mortensen 1999, Bouchet 2007, Denoyelle et al.  
118 2010, Dolven et al. 2013, Hess et al. 2013, Wlodarska-Kowalczyk et al. 2013). For instance,  
119 benthic foraminiferal and macrofaunal communities showed similar response to oil-based

120 drill mud disposal with tolerant and opportunistic species dominating in the vicinity of the  
121 disposal area (Denoyelle et al. 2010). Furthermore, calcareous foraminifera can be reliable  
122 indicators for variability in density, diversity and species composition of benthic macrofauna  
123 in the Arctic at Svalbard (Wlodarska-Koawalczuk et al. 2013). Hence, it is reasonable to  
124 suggest that macrofaunal and foraminiferal species with similar sensitivity to environmental  
125 changes may show co-occurrence. This would be of great interest when it comes to the  
126 definition of reference conditions. It is reasonable to think that fossil benthic foraminifera  
127 could be used as proxies to define reference conditions for benthic macrofauna  
128 communities. Further baseline studies are however necessary to validate such a hypothesis.  
129 It is thus urgent to quantify the level of correlation between the distribution patterns of  
130 these two groups in other environments.

131 In the present study, living benthic foraminifera and macrofauna were investigated  
132 in fjordic systems along the Norwegian Skagerrak coast to assess to which degree their  
133 responses to major driving environmental factors correspond. This work is part of a  
134 comprehensive project (PES), which includes both foraminifera and macrofauna collected at  
135 the same sites at the same time. An understanding of the driving forces of the benthic  
136 foraminifera communities, expressed as species diversity, was partly achieved in a previous  
137 analysis of the same data set (Bouchet et al. 2012, 2013). The present paper aims (i) to  
138 determine the driving environmental factors of both the benthic foraminiferal and  
139 macrofaunal community structures, (ii) to identify indicator species of both normal and poor  
140 environmental conditions, (iii) to compare species assignment in EG from this study to the  
141 existing Foram-AMBI and AMBI species lists, and (iv) to assess the correlation between  
142 benthic foraminiferal and macrofaunal communities patterns to identify the potential of



143 benthic foraminifera to serve as a proxy for benthic macrofauna in environmental  
144 assessments.

145

## 146 **2. Material and methods**

147

### 148 **2.1 Study area and sampling sites**

149

150 In August 2008, 27 stations from 11 silled basins along the Norwegian Skagerrak  
151 coast, NE North Sea (Figure 1), were sampled for bottom water, sedimentological,  
152 biogeochemical and faunal analyses. Stations between 23 and 204 m water depth were  
153 selected to provide an oxygen gradient with stable temperature (5-6°C; occasionally 8-9°C at  
154 <40 m depth) and salinity (33-34) conditions (Table 1; see Bouchet et al. 2012 for further  
155 details) using information from previous studies (Buhl-Mortensen et al. 2009 and  
156 unpublished data from the Norwegian Institute for Water Research (NIVA) and Institute of  
157 Marine Research (IMR)). The study basins commonly experience partial to complete deep-  
158 water renewals during winter. In addition to measured variables, the depth below threshold  
159 (DBT) *i.e.* the difference between the water depth at a station and the sill depth of the  
160 fjord, has been characterized for stations. DBT is not a factor *sensu stricto*, but a measure  
161 that reflects the degree or duration of water stagnation in the deep areas of the fjord.

162

### 163 **2.2 Field sampling**

164

165 Four sediment cores (8 cm diameter) were collected at each of the 27 stations with a  
166 Gemini gravity corer, a modified Niemistö corer (Niemistö 1974). Three replicate cores were

167 used for benthic foraminifera and one for pigments, total organic carbon and total nitrogen  
168 analyses. Once on deck, bottom water samples from just above the sediment-water  
169 interface in two cores were immediately transferred to Winkler bottles for subsequent  
170 dissolved oxygen analysis. All sediment cores were sectioned on board and, for the present  
171 study, the top 0-1 and 1-2 cm slices were analysed. Sediment samples for pigments, total  
172 organic carbon and total nitrogen analyses were frozen immediately after sectioning, and  
173 kept away from the light. Foraminiferal samples were preserved in rose Bengal-stained 70%  
174 ethanol ( $1 \text{ g l}^{-1}$ ) to avoid protoplasm degradation and to distinguish living (stained) from  
175 dead specimens (Murray & Bowser 2000). At each station, macrofauna was sampled using a  
176  $0.1 \text{ m}^2$  van Veen grab (4 replicates) and a sub-sample of the top 0-1 cm from one grab per  
177 station was collected for grain size analyses. The grab sediments were sieved on board  
178 through 1 mm screens and fixed in 4% buffered formaldehyde in seawater to preserve  
179 macrofauna specimens.

180

### 181 **2.3 Biogeochemical analyses**

182

183 Bottom-water dissolved oxygen concentrations at the time of sampling were  
184 analysed using Winkler titration. Classification of stations as normoxic (bottom-water  $[\text{O}_2] >$   
185  $2 \text{ mL O}_2\cdot\text{L}^{-1}$ ), hypoxic ( $0.5 - 2 \text{ mL O}_2\cdot\text{L}^{-1}$ ) or severe hypoxic ( $< 0.5 \text{ mL O}_2\cdot\text{L}^{-1}$ ) follows Diaz and  
186 Rosenberg (1995). Sediments were freeze-dried prior to sedimentological and geochemical  
187 analyses. For grain size analyses, the dried sediment was weighed, soaked in tap water, and  
188 washed on a  $63 \mu\text{m}$  Endecote-sieve. The  $>63 \mu\text{m}$  fraction was dried and weighed and the  
189  $<63 \mu\text{m}$ -fraction was calculated based on the dry mass. Total organic carbon (TOC) and total  
190 nitrogen (TN) were analysed following acid treatment (10% HCl) to remove inorganic C using

191 a CHN analyser (Carlo Erba Elemental Analyzer 1106). Classification of stations as low  
192 (<2.7%), medium (2.7-3.4%) and high (>3.4%) TOC content follows that of Molvaer et al.  
193 (1997). For pigment analyses, homogenised freeze dried sediment (0.3-0.6 g) was extracted  
194 in 5 ml 90% acetone in water. The mixture was sonicated for 30s before overnight  
195 extraction. The extract was centrifuged (15 min. at 3500 rpm), and 100 µL of the  
196 supernatant was injected on the HPLC for pigment analysis. The analytical system was based  
197 on a method described earlier (Wright et al. 1991, Jeffrey et al. 1997). Pigments were  
198 identified by comparison of retention time and absorption spectra of authentic standards  
199 (DHI Water and Environment, Denmark) and the literature (Jeffrey et al. 1997). Individual  
200 pigments were quantified at 436 nm for chlorophylls and pheopigments, and 450 nm for  
201 carotenoids. Response factors (RF) for each pigment were obtained by single standard runs.  
202 When no standard was available, RF for similar pigments was used with a correction for  
203 differences in mass. Pigment concentrations were standardised by the total organic carbon  
204 content (Reuss et al. 2010).

205

#### 206 **2.4 Faunal analyses**

207

208 Foraminiferal samples were washed through 500 and 63 µm mesh sieves, and the  
209 63-500 µm fraction was split into 8 using a modified Elmgren wet splitter (Elmgren 1973).  
210 One eighth of each sample was re-sieved and all live (stained) foraminifera in the 63-125  
211 and 125-500 µm fractions were identified to species level and counted in the wet state. The  
212 number of individuals >500 µm relative to smaller ones was trivial (<0.1%) so including them  
213 would not influence the results. Wet-sample analysis allows preservation of all species,

214 including fragile organic-walled and loosely cemented agglutinated foraminifera. This study  
215 is based on the 63-500  $\mu\text{m}$  fraction *i.e.* the sum of 63-125 and 125-500  $\mu\text{m}$  fractions.

216 In the laboratory, macrofaunal samples were rinsed on 1 mm-mesh sieves and  
217 transferred to 70% ethanol. All macrofaunal specimens were counted and identified to  
218 species level, or the lowest identifiable taxonomic unit.

219 Assignments of foraminiferal and macrofaunal species into ecological groups (EG)  
220 were made according to Alve et al. (2016) and Borja et al. (2000), respectively. The concept  
221 is based on the classification of species (or groups of species) into five EGs (sensitive,  
222 indifferent, tolerant, 2<sup>nd</sup> order opportunist and 1<sup>st</sup> order opportunist) representing specific  
223 sensitivity levels to an increasing gradient of organic matter (Borja et al. 2000). The groups  
224 allow the assessment of the ecological quality status of marine systems.

225

## 226 **2.5 Numerical analyses**

227

228 Principal components analysis (PCA) was applied on the set of environmental  
229 variables to characterize the main natural gradients and describe relationships between  
230 sediments, oxygen and fjord topography. For the species community data, initial detrended  
231 correspondence analyses indicated that the gradient is short (1.6 SD) in the foraminiferal  
232 data and long (3.3 SD) in the macrofaunal data. Consequently, redundancy analysis (RDA)  
233 and canonical correspondence analysis (CCA), respectively, were applied to find which  
234 environmental variables were most clearly related to the species communities. Analyses  
235 were done on the subset of 15 stations with all chemistry data and at least 30 individuals  
236 in the pooled macrofaunal sample (all pooled foraminiferal samples had many more than 30  
237 individuals). Community data were  $\log(x + 1)$  transformed prior to analysis; taxa occurring at

238 only one station were removed. Environmental variables were selected using forward  
239 selection with 999 permutations. Procrustes analysis (Peres-Neto & Jackson 2001) was used  
240 to compare unconstrained ordinations of the foraminiferal (PCA) and  
241 macrofaunal (correspondence analysis) community data. The  $m^2$  statistics, which is  
242 analogous with the  $r^2$  of a correlation, is reported. Co-correspondence analysis (Co-CA, Ter  
243 Braak & Schaffers 2004) was used to relate the foraminiferal and the macrofaunal  
244 communities in a direct way.

245 All calculations were performed using the statistical language R version 3.4.1 (R Core  
246 Team, 2017). Ordinations were run with the vegan version 2.4-3 (Oksanen et al. 2010). R  
247 code is archived at <https://github.com/richardjtelford/PES>.

248

### 249 **3. Results**

250

#### 251 **3.1 Environmental conditions at sampling stations**

252

253 Depth-below threshold (DBT), bottom-water dissolved oxygen concentration,  
254 sediment grain size and sediment organics were rather different among the stations (Table  
255 1). Sandnesfjord, Eidangerfjord, Risørbassenget (R60), Topdalsfjord, Goosefjord, Frierfjord  
256 (except F70) and Indre Hvaler have rather shallow DBT below 50 m, whereas the other  
257 fjords have DBT up to 169 m. In Kristiansandsfjord there is no sill. High (normoxic) bottom-  
258 water oxygen concentrations ( $>2 \text{ mL O}_2 \cdot \text{L}^{-1}$ ) characterized stations in the Indre Hvaler basin,  
259 Kragerøfjord, Topdalsfjord, the shallower stations in Goosefjord, Frierfjord (F30 and F50),  
260 Ærøydypet, Sandnesfjord, Eidangerfjord and Kristiansandsfjord as opposed to stations in  
261 Risørbassenget (except R60), the deeper stations in Frierfjord (F70 and F90) and Håøyfjord

262 where hypoxic ( $0.5\text{-}2\text{ mL O}_2\cdot\text{L}^{-1}$ ) to severe hypoxic ( $<0.5\text{ mL O}_2\cdot\text{L}^{-1}$ ) conditions are recorded.  
263 Sediments at stations in Risørbassenget (except R160 and R180) and Kristiansandsfjord, as  
264 well as station G50 were sandier than the others, with less than 90% of the sediment mass <  
265  $63\text{ }\mu\text{m}$  (Table 1). Total organic carbon concentrations were higher than 3.4% in Ærøydypet,  
266 Kragerøfjord, Sandnesfjord, Eidangerfjord, Risørbassenget, Topdalsfjord, Frierfjord (F90) and  
267 Goosefjord (Table 1).

268 In the PCA of environmental variables, the first two axes explained 75.2% of the  
269 variance (Fig. 2). Two main gradients appear; one related to sediment components (particle  
270 fine fraction, TOC) and one to oxygen and fjord basin topography ( $\text{O}_2$ , DBT). The sediment  
271 gradient separates stations with very high fine fraction and moderate organic content  
272 (lower left: Indre Hvaler IH, Frierfjord F) from stations with lower fine fraction and high  
273 organic contents (upper right: Goosefjord G, Risørbassenget R). The oxygen – fjord  
274 topography gradient separates stations in deep fjord basins with shallow sill and low oxygen  
275 content (lower right: Risør basin R, Håøyfjord 102, deep Frierfjord F) from stations with  
276 either shallow water depths or in fjords with deep sills (upper left). The pigments correlate  
277 with DBT and characterized the deep basins, particularly the Risør basin. Most pigments  
278 were intercorrelated, except allo-xanthin that appeared to be related also to the sediment  
279 fine fraction.

280

### 281 **3.2 Environmental variables and fauna**

282

283 In total, 116 foraminiferal taxa and 290 macrofaunal taxa were identified. Benthic  
284 foraminifera occurred in all samples, whereas macrofauna were absent from station F90  
285 (replicate 1 and 2), station R120 (replicate 4), station R160 (all replicates) and station R180

286 (all but replicate 1). Based on RDA and CCA, respectively, benthic foraminifera and  
287 macrofauna species were significantly related to the same subset of environmental  
288 parameters, viz. TOC and DBT (Figs 3 and 4). Together, the selected parameters explained  
289 43% and 29% of the variation for foraminiferal and macrofaunal species assemblages,  
290 respectively.

291 The distribution of foraminiferal taxa in relation to TOC and oxygen, the latter being  
292 related to DBT (Fig. 2), is illustrated in Fig. 5. The species *Micrometula hyalostriata*,  
293 *Phainogullmia aurata* and *Spiroplectammia biformis* mainly occur at low TOC (<2.7%) and  
294 high bottom-water oxygen concentrations. Highest abundances of *Cassidulina laevigata* and  
295 *Bulimina marginata* were observed at stations with low to high TOC (0.8-9%) and high  
296 bottom-water oxygen concentrations. *Cylindrogullmia alba* and *Leptohalysis scottii* occur at  
297 similar TOC levels but at stations with high to low bottom-water oxygen concentrations.  
298 *Liebusella goësi*, *Textularia earlandi* and *Recurvoides trochamminiforme* show highest  
299 abundances under TOC concentrations between 3.0 and 4.3% and well-oxygenated  
300 conditions. *Stainforthia fusiformis*, *Discorbinella bertheloti*, *Fissurina* sp. and *Bolivinellina*  
301 *pseudopunctata* thrive under high TOC concentrations (>3.4%) and hypoxic to severe  
302 hypoxic conditions.

303 Compared to the assignments of foraminifera in Alve et al. (2016) four new species  
304 (*Micrometula hyalostriata*, *Phainogullmia aurata*, *Cylindrogullmia alba*, *Leptohalysis scottii*)  
305 were assigned and three (*Cassidulina laevigata*, *Liebusella goësi*, *Bolivinellina*  
306 *pseudopunctata*) were re-assigned based on their abundance relative to sediment TOC in  
307 this new data set (Table 2).

308 The distribution of dominant macrofaunal species on TOC and oxygen is illustrated in  
309 Fig. 6. *Pseupopolydora* sp., *Chaetozone setosa*, *Capitella capitata*, *Thyasira* cf. *sarsii* occurred

310 at hypoxic stations with high TOC. *Mediomastus fragilis* was overabundant in  
311 Kristiansandfjord that is contaminated from trace elements. *Thyasira equalis* and  
312 *Spiophanes kroeyeri* occurred at well-oxygenated stations with medium TOC content.  
313 *Amphiura chiajei*, *A. filiformis* and *Scalibregma inflatum* occurred at normoxic stations (Fig.  
314 6B) with low organic matter concentrations (Fig. 6A).

315 Compared to the assignments of the AMBI list (Borja et al. 2000, last update: June  
316 2017), two species (*Scalibregma inflatum* and *Thyasira cf. sarsi*) were re-assigned based on  
317 their abundance relative to sediment TOC in this new data set (Table 2).

318

319

### 320 **3.3 Foraminiferal and macrofaunal community relationships**

321

322 Foraminiferal and macrofaunal community data were significantly correlated  
323 (Procrustes analysis  $m^2 = 0.66$ ,  $p = 0.001$ ). Using a co-correspondence analysis on  
324 foraminiferal and macrofaunal community, 24% of the variance in the macrofaunal  
325 community was explained by the foraminiferal community (Fig. 7).

326

## 327 **4. Discussion**

328

329 In the present study, patterns of distribution of foraminiferal and macrofaunal  
330 species are best explained by the variables of TOC and DBT. Along the same lines, deep-  
331 water renewal and oxygenation in a fjord depend on DBT as well as on the supply of organic  
332 matter (*i.e.*, impacts oxygen consumption). DBT will, however, also represent the influence  
333 of other variables, e.g. pigments, different organic components and temperature, that



334 increase or decrease due to the gradually changing conditions with depth. In the following  
335 discussion, however, it will be made reference to oxygen rather than of DBT *sensu stricto*  
336 that may incorporate several unknown factors. This is further supported by Bouchet et al.  
337 (2012) who showed on the same dataset that benthic foraminiferal diversity is correlated to  
338 bottom-water dissolved water oxygen. These two variables (oxygen and TOC) are indeed  
339 known to be drivers of both diversity and species composition of benthic foraminifera and  
340 macrofaunal communities (Alve 1990, Riera et al. 1999, Gustafsson & Nordberg 2000,  
341 Rosenberg et al. 2002, Bouchet et al. 2012). In the studied Norwegian fjords, benthic  
342 foraminiferal and macrofaunal species responded in the same way to different  
343 environmental conditions (Procrustes analysis), except that macrofauna was less tolerant to  
344 the oxygen depleted conditions. This is because, unlike macrofauna, some foraminiferal  
345 species can perform anaerobic metabolism (e.g., Risgaard-Petersen et al. 2006). Our findings  
346 confirmed the results from the few studies assessing and comparing the response of benthic  
347 foraminifera and macrofauna to different environmental conditions, mentioned in the  
348 introduction.

349

#### 350 **4.1 Foraminiferal indicator species**

351

352 In this study, *Micrometula hyalostriata*, *Phainogullmia aurata* and *Spiroplectammina*  
353 *biformis* are sensitive (EG1) to organic enrichment, occurring primarily in normoxic  
354 conditions. *Phainogullmia aurata* has been described as a pioneer species showing  
355 maximum abundance in the colonization phase on uncontaminated clay and almost  
356 disappearing with time as the amount of sediment organic material increased (Hess et al.

2014). The ecology of the soft-walled species *M. hyalostrata* is poorly known. In this study, it was common only at normoxic, low-TOC sites (Fig. 5). *Spiroplectammina biformis* was the second-most common species in the most hypoxic parts of the organic matter enriched Drammensfjord, southern Norway (Alve 1995) but it was not able to survive persistent severe hypoxia below 0.5 mL O<sub>2</sub>.L<sup>-1</sup> (Alve 1991, 1995). It was also reported to tolerate organic enrichment in several eastern Canadian estuaries and embayments and in the North-east Atlantic (Alve et al. 2016, Schafer et al. 1991). In the present study, its highest abundance was at a low-TOC site. It was also sensitive to TOC in the Saguenay fjord and in a study in Canadian fish aquaculture sites (Schafer et al. 1991, 1995).

*Cassidulina laevigata* and *Cylindrogullmia alba* show the same distribution pattern along the TOC gradient as for species from the indifferent EG2. *Cassidulina laevigata* is a typical species of the Skagerrak-Kattegat fauna (*sensu* Nordberg et al. 2000), which used to flourish in several Scandinavian fjords until it was replaced by species associated with frequent coastal hypoxia (e.g., Alve 1991, Polovodova Asteman & Nordberg 2013, Polovodova Asteman et al. 2015). In the present study, *C. laevigata* is never abundant, it only occurs at stations with normoxic conditions and mainly, but not solely, where TOC is low to moderate (Fig. 5). Hence, *C. laevigata* may belong in the indifferent group EG2, although it has been classified as a sensitive species (Alve et al. 2016). The present results show that *C. alba* can survive under a wide range of TOC concentrations.

*Bulimina marginata*, *Recurvoides trochamminiforme*, *Liebusella goësi* and *Textularia earlandi* are all tolerant species (EG3) to organic enrichment in the studied fjords, occurring in normoxic conditions. *Bulimina marginata* is tolerant to oxygen-depleted conditions (Alve 1991, Barmawidjaja et al. 1992). However, it is not able to survive persistent severe hypoxia below 0.5 mL O<sub>2</sub>.L<sup>-1</sup> (Filipsson & Nordberg 2004). In this study, it occurred over a broad

381 range of TOC concentrations, but only under normoxic conditions. As for *R.*  
382 *trochamminiforme*, Alve et al. (2016) mentioned it as a tolerant species. *Liebusella goësi* is  
383 known not to survive severe hypoxia (Nordberg et al. 2000, Filipsson & Nordberg 2004).  
384 Tolerance to TOC enrichment under appropriate oxygen concentrations has already been  
385 observed for *T. earlandi* (Schafer et al 1995). It is however sensitive to oxygen depletion  
386 (Gustafsson & Nordberg 2000, Polovodova Asteman et al. 2015). Its abundance decreased at  
387 dissolved-oxygen concentrations below 1.0 ml O<sub>2</sub>.L<sup>-1</sup> (Gustafsson & Nordberg 2000).  
388 *Bolivinellina pseudopunctata*, *Stainforthia fusiformis*, *Leptohalysis scottii* and  
389 *Fissurina* sp. are assigned in the opportunistic EG4/5, thriving well in sediment with high  
390 TOC content based on this study results, occurring at hypoxic to severe hypoxic stations. The  
391 two first species are known to be able to survive at high TOC and hypoxic to severe hypoxic  
392 conditions (Alve 1994, Gustafsson & Nordberg 2000, Alve 2003). These species  
393 outcompeted the typical Skagerrak-Kattegat species throughout the degradation of the  
394 environmental conditions in the area during the 1900s (Gustafsson & Nordberg 2001, Alve  
395 et al. 2009, Dolven et al. 2013). *Bolivinellina pseudopunctata* is an opportunistic species able  
396 to tolerate low oxygen conditions (Gustafsson & Nordberg 2001), although it has been  
397 reported to disappear after periodic severe hypoxia or anoxia (Gustafsson & Nordberg  
398 2000). The observation of this species in the most organic rich stations supports that it is an  
399 opportunistic species, which is not in agreement with the classification of Alve et al. (2016)  
400 in the indifferent EG2. Our results suggest that *L. scottii* is able to survive under hypoxic  
401 conditions as opposed to what has been reported in Havstensfjord in Sweden (Gustafsson &  
402 Nordberg 2000). Furthermore, it occurs in this study in fjords with a wide range of TOC  
403 concentrations from 2 to 9%. It is a species known to tolerate environmental disturbances  
404 such as paper mill and fish farming effluents (Alve & Nagy 1986, Scott et al. 2001,

405 Polovodova Asteman et al. 2015) and it is common in eutrophic environments (Scott et al.,  
406 2005; Sabbatini et al., 2012). *Stainforthia fusiformis* is a characteristic species of heavily  
407 disturbed environments (see review in Alve 2003). Except that species of *Fissurina* seem to  
408 be ectoparasites on *Discorbis vilardeboanus* (d'Orbigny) and *Rosalina bradyi* (Cushman) (see  
409 Collen & Newell, 1999, and references therein), little is known about the ecology of  
410 *Fissurina* spp. In the present material, *Fissurina* sp always co-occurs with *Discorbinella*  
411 *bertheloti*, perhaps as a parasite?

412 *Discorbinella bertheloti* (also found under the name *Cibicides bertheloti*) was found  
413 alive (stained) at hypoxic and severe hypoxic stations. With their epifaunal mode of life  
414 (Murray 2006), *Cibicides* spp. have been considered as sensitive species not directly linked  
415 to sedimentary TOC (Alve et al. 2016). They rather reflect the impact of bottom-near  
416 currents (Linke & Lutze 1993). Hence, occurrence of *D. bertheloti* at the most oxygen-  
417 depleted stations with high TOC content was probably due to recent transport of living  
418 specimens. It is thus difficult to conclude on the indicative value of this species.

419

## 420 **4.2 Macrofaunal indicator species**

421

422 In this study, *Amphiura chiajei*, *A. filiformis*, *Scalibregma inflatum* are sensitive  
423 species (EG1) to TOC, occurring at normoxic stations. They are known to not tolerate  
424 oxygen-depleted conditions (Aschan & Skullerud 1990, Nilsson & Rosenberg 1994, 2000,  
425 Rosenberg et al. 2001). The three species have been reported to solely occur at unpolluted  
426 sites in the Oslofjord (Mirza & Gray 1981). A high abundance and biomass of *A. filiformis* and  
427 *A. chiajei* has been observed at well oxygenated sites in the Skagerrak (Josefson 1990).  
428 According to AMBI's ecological groups (EG) of sensitivity to disturbances (Borja et al. 2000),

429 *A. chiajei* and *A. filiformis* are indifferent species (EG2) and *S. inflatum* is a tolerant species  
430 (EG3). *Amphiura filiformis* has been reported to be sensitive to severe disturbances around  
431 oil platforms (Olsgard & Gray 1995). Note that the tolerance level of *S. inflatum* is rather  
432 uncertain since it has been reported to be either sensitive (Mirza & Gray 1981, Rygg 1985b,  
433 Nilsson & Rosenberg 1994, 2000, Rosenberg et al. 2001, Pabis & Sobczyk 2015) or tolerant  
434 (Rygg 1985a, Aschan & Skullerud 1990, Borja et al. 2000, Levin et al. 2009) to disturbances.

435 *Thyasira equalis* and *Spiophanes kroeyeri* dominate at well-oxygenated stations  
436 moderately rich in organic matter. They are not present at the hypoxic/severe hypoxic  
437 stations, suggesting that neither of the species would tolerate low oxygen concentrations.  
438 This contradicts other works on these species (see review in Levin et al. 2009). For instance,  
439 *T. equalis* has been reported to survive low-oxygen concentrations (Josefson & Widbom  
440 1988, Nilsson & Rosenberg 2000). It usually occurs in moderately disturbed areas (Pearson  
441 1975, Mirza & Gray 1981, Rygg 1985a), although it has been reported not to tolerate copper  
442 pollution (Rygg 1985b). The results of the present study are in accordance with the  
443 classification of *T. equalis* in the tolerant EG 3 (Borja et al. 2000). Conversely, *S. kroeyeri* has  
444 been mostly reported to disappear in the early stage along a gradient of increasing  
445 disturbances *i.e.* oil platform activities, sewage outfall, copper, organic enrichment, oxygen  
446 depletion (Mirza & Gray 1981, Rygg 1985b, Aschan & Skullerud 1990, Nilsson & Rosenberg  
447 2000, Mojtahid et al. 2008, Oug et al. 2012). Few studies nevertheless reported the  
448 presence of this species in low disturbed sites (Pearson 1975, Gray & Pearson 1982, Rygg  
449 1985a, Moore & Rodger 1991). The classification of *S. kroeyeri* in the tolerant EG3 (Borja et  
450 al. 2000) is at least questionable.

451 *Pseupopolydora* sp., *Chaetozone setosa*, *Capitella capitata* and *Thyasira* cf. *sarsii*  
452 occur at stations with hypoxic conditions and high TOC contents. *Mediomastus fragilis*

453 dominates in the heavily polluted with trace metals Kristiansandfjord. It belongs together  
454 with *T. cf. sarsii* to the tolerant EG3, *C. setosa* and *P. sp.* belong to the second-order  
455 opportunistic EG4 and *C. capitata* to the first-order opportunistic EG5 (AMBI-list, Borja et al.  
456 2000). They are all characterizing low-oxygenated environments (Levin et al. 2009).  
457 *Mediomastus fragilis* can tolerate pollution such as oil spills, disturbances associated to oil  
458 platforms, metals and high level of organic enrichment (Pearson & Rosenberg 1978, Mirza &  
459 Gray 1981, Dauvin 2000, Mojtahid et al. 2008, Oug et al. 2012). *Thyasira cf. sarsii* is sensitive  
460 to low oxygen concentrations (Josefson & Widbom 1988, Nilsson & Rosenberg 2000) but it  
461 can colonize sediments polluted by copper, organic matter, and dredging materials from oil  
462 platform activities (Pearson 1975, Rygg 1985b, a, Olsgard & Gray 1995). *Capitella capitata*  
463 and *P. spp.* are known colonizers of completely anoxic sediments in fjord systems  
464 (Rosenberg et al. 2001). They are often the last species to survive these depleted conditions  
465 (Pearson & Rosenberg 1978, Mirza & Gray 1981, Oug et al. 2012). *Chaetozone setosa* and *C.*  
466 *capitata* are typical indicators of severe effects around oil platforms of the Norwegian  
467 continental shelf (Olsgard & Gray 1995). *Chaetozone setosa* usually thrives in depleted  
468 conditions (Rygg 1985a, Olsgard & Hasle 1993, Pabis & Sobczyk 2015). It is a common  
469 species in organic enriched areas (Gray et al. 1990).

470

#### 471 **4.3 Concerns about the indicative value of classified species**

472

473 The use of the presence of certain species to assess the state of marine systems has  
474 a long history. The concept is based on an *a priori* apprehension that some species have  
475 narrow ecological requirements. Hence, species are meant to be indicative of the prevailing  
476 conditions where they are found. Based on numerous works (e.g., Pearson & Rosenberg

477 1978, Glémarec & Hily 1981, Hily 1983), lists of benthic macrofaunal species, assigned into  
478 groups of sensitivity to disturbances, have been established; the most widely used being the  
479 species associated to the AMBI index (Borja et al. 2000). Lately, following the suggestions of  
480 the FOBIMO working group, the same approach was used to assign benthic foraminiferal  
481 species to EGs of sensitivity/tolerance to conditions along an increasing stress (organic  
482 matter enrichment) gradient (Alve et al. 2016, Jorissen et al. 2018). So far, assignment of  
483 foraminiferal species has only been done for the North-East Atlantic and Arctic fjords,  
484 continental shelves, and slopes and for the Mediterranean (Alve et al. 2016, Jorissen et al.  
485 2018). The use of a single species list classification means that the indicative value of a  
486 species is a static concept *i.e.* a species is expected to have a similar sensitivity or tolerance  
487 wherever it occurs and regardless of the adaptation ability of this species. However, it is  
488 known that some species are plastic enough to adapt to their environment and could  
489 change their autecology requirements along environmental gradients (see review in Zettler  
490 et al. 2013 and references therein). Hence, a species tolerance spectrum might be wider  
491 than a single category, *i.e.* “sensitive species” or “tolerant species”. Species even exhibit  
492 different responses to disturbance depending on their habitat and the source of  
493 disturbances; they behave as sensitive species while they can be tolerant or opportunistic  
494 somewhere else or against a different pollution source (Zettler et al. 2013). The  
495 biogeographical/ecosystem-type restricted approach chosen to assign foraminiferal species  
496 tend to support the ability of species to adapt their sensitivity/tolerance level to their  
497 environment.

498         In this study, we reported discrepancies in the assignment in EG with the study of  
499 Alve et al. (2016) for *S. biformis*, *C. laevigata*, *L. goësi* and *B. pseudopunctata*. For *C.*  
500 *laevigata*, our study results suggest a classification in the indifferent EG2 instead of the

501 sensitive EG1 (Alve et al. 2016). These two EGs are rather close conceptually since they  
502 include species that cannot tolerate any excess in organic matter. Some biotic indices, *i.e.*  
503 Bentix (Simboura & Zenetos 2002), group these two EGs in one EG. This might be something  
504 to consider in a further implementation of the Foram-AMBI list. *Spiroplectammina biformis*  
505 was assigned to the tolerant EG in the Foram-AMBI list of species from the North-East  
506 Atlantic (Alve et al. 2016). In several eastern Canadian estuaries and embayments, *S.*  
507 *biformis* has been reported in moderately polluted local environments (Schafer et al. 1991).  
508 This study results however suggest that it is a sensitive species to TOC enrichment. In the  
509 Saguenay fjord, *S. biformis* abundances have also been reported to decrease with increasing  
510 organic matter load from a pulp mill effluent (Schafer et al. 1991). In a study in four  
511 Canadian aquaculture sites, it showed a significant negative correlation to TOC enrichment  
512 due to the presence of fish cages (Schafer et al. 1995). These evidences suggest that it is  
513 thus not sure whether this species belongs to the sensitive EG1 or the tolerant EG3.  
514 Discrepancies in the response of species to TOC enrichment have been found between this  
515 study results and previous work for both benthic foraminifera and macrofauna. As for  
516 benthic macrofauna, there are eight studies, including this one, reporting a sensitive  
517 behavior of the polychaete *S. inflatum* towards disturbances and at least four reporting a  
518 tolerant one (see aforementioned references). In the AMBI species list, this species is  
519 assigned to the tolerant EG. The assignment of *S. inflatum* within the AMBI list is rather  
520 uncertain. The same concerns arose about *T. cf. sarsii* which behave like an opportunistic  
521 species in this study and is assigned to tolerant EG in the AMBI-list. Occurrences of *T.*  
522 *equalis*, *S. kroeyeri* and *S. inflatum* in several Norwegian fjords and coastal areas (Rygg &  
523 Norling 2013) showed tolerance levels corresponding to EG3 (tolerant), thus supporting the  
524 classification in the AMBI system. These discrepancies may be due to the fact that indices'



525 scores come from large dataset collections covering large geographical regions and  
526 subregions and not local scales (Borja et al. 2000, Rosenberg et al. 2004). Hence, they  
527 integrate the "experience" from a species behavior over a large span of data or literature  
528 data. Local adaptation of species ecological requirement may lead to wrong interpretation of  
529 the species indicative value (Dauvin et al. 2010, Zettler et al. 2013).

530 Zettler et al. (2013) reported that the response of macrofaunal species towards  
531 organic enrichment would change along a gradient of salinity. They concluded that the  
532 interaction between environmental variables should not be neglected when using static  
533 indicative value of a species. This study results tend to confirm the latest concerns about the  
534 concept of species lists of tolerance/sensitivity. Furthermore, this study shows that these  
535 concerns may apply to benthic foraminifera. At a large biogeographical scale, the indicative  
536 value may be reliable, but it is rather uncertain at a local geographical scale. This study  
537 results definitively support cautions (Dauvin et al. 2010, Zettler et al. 2013) when it comes to  
538 the use of indicator species lists. It seems obvious that more studies are needed to constrain  
539 these biases. Supplementary approaches should be considered, for instance rank-frequency  
540 diagrams (Frontier 1976, Bouchet et al. 2007, Seuront 2013) and diversity indices (Bouchet  
541 et al. 2012, Dolven et al. 2013, Francescangeli et al. 2016, Bouchet et al. 2018).

542

#### 543 **4.4 Benthic foraminifera as indicator of macrofaunal community structure**

544

545 The assessment and quantification of cross-taxon congruence *i.e.* similar response  
546 along an environmental gradient of species belonging to different taxa, in space and time,  
547 represents a critical step in the identification of suitable indicator taxa for biodiversity. In  
548 this study, Procrustes rotation and co-correspondence analysis (Co-CA) were used to

549 quantify the strength of the congruence between foraminiferal and macrofaunal  
550 communities along the studied environmental parameters gradient. Procrustes rotation and  
551 Co-CA are powerful and robust methods to evaluate the degree of congruence between two  
552 community datasets (Peres-Neto & Jackson 2001, Gioria et al. 2011). Patterns of  
553 foraminiferal species distribution closely resembled those of the macrofauna, as is showed  
554 by significant correlation between the groups. The strength of the congruence in species  
555 composition between benthic foraminifera and macrofauna along the environmental  
556 gradient reflects the similarity in their response to TOC enrichment. Interactions can explain  
557 congruence in species between these two groups. First, there may be a trophic link between  
558 the two groups; benthic macrofauna species being able to feed on benthic foraminifera  
559 (Lipps 1983). Secondly, benthic macrofauna bioturbating activities enhance the vertical  
560 distribution of foraminifera in deeper sedimentary layers (Bouchet et al. 2009). Last but not  
561 least, there is an evident similarity in the patterns of distribution of species of both groups in  
562 response to environmental gradients.

563         In this study, benthic foraminiferal and macrofaunal species composition appear to  
564 respond to the same environmental factors, in particular bottom-water dissolved oxygen  
565 and organic matter content. Changes in the composition of foraminiferal and macrofaunal  
566 assemblages are expressed in increased dominance of tolerant/opportunistic species and  
567 the progressive disappearance of sensitive species under degraded conditions *i.e.* increasing  
568 TOC content and decreasing bottom-water oxygen concentrations. It supports other studies,  
569 although there are few, showing similar response of the two groups to adverse  
570 environmental conditions. Patterns of foraminiferal species distribution closely resembled  
571 those of macrofauna in response to disturbances of a glacier (Wlodarska-Kowalczyk et al.  
572 2013). Similar succession from sensitive to tolerant species has been reported for both

573 groups in response to different pollution sources such as sewage outfalls, industry, oil-based  
574 drill mud disposal and aquaculture (Schafer et al. 1975, Schafer et al. 1995, Mojtahid et al.  
575 2008, Denoyelle et al. 2010). However, some studies suggested that benthic foraminifera  
576 could be more indicative than benthic macrofauna along a gradient of disturbances  
577 (Mojtahid et al. 2008, Denoyelle et al. 2010). In this study, benthic foraminifera exhibited  
578 more tolerance to the most depleted conditions (high TOC and anoxic conditions), by  
579 maintaining higher densities than did the macrofauna. This confirms previous results  
580 showing that benthic macrofauna is more sensitive than benthic foraminifera to depleted  
581 oxygen conditions (Josefson & Widbom 1988) and, as previously mentioned, is directly  
582 linked to the fact that some foraminiferal species perform anaerobic metabolism (*e.g.*,  
583 Risgaard-Petersen et al. 2006). It implies that benthic foraminifera are able to occur along  
584 the whole environmental gradient, which is an asset compared to benthic macrofauna in  
585 monitoring studies when approaching the “bad” end of the environmental gradient. The  
586 strength of the congruence between benthic foraminifera and macrofauna suggests that  
587 foraminifera could be reliable indicators of benthic macrofauna community structure. This  
588 has interesting implications, in particular with regard to defining reference conditions for  
589 areas where the environmental conditions may have changed over time, as suggested by  
590 Alve et al. (2009). Defining reference conditions are mandatory according to the WFD and  
591 complying to MSFD when it comes to assessing the health of marine systems. Hindcasting is  
592 one method that can be used to determine previous conditions. This is hardly possible with  
593 benthic macrofauna since historical data are scarce. Mostly the molluscs can fossilise and  
594 allow a reconstruction of palaeo-environments (Poirier et al. 2009), meaning that most of  
595 the biological signal is lost in the fossil sediment. Conversely, most of the foraminiferal  
596 community can fossilise and be preserved in the fossil sediment. It allows reconstruction of

597 palaeo-environments (e.g., Alve 1991, Alve et al. 2009, Dolven et al. 2013, Polovodova  
598 Asteman et al. 2015, Francescangeli et al. 2016). Using fossil benthic foraminifera, it is  
599 possible to determine objective and reliable reference conditions. Hence, fossil benthic  
600 foraminifera could be used as proxies to assess the deviation from reference conditions for  
601 benthic macrofauna communities. However, this requires an algorithm for translating  
602 foraminiferal community data into macrofaunal community data (specified macrofaunal  
603 species and their abundances). More conceivably, the ecological quality status has to be  
604 estimated directly from the foraminiferal data. This requires an environmental classification  
605 system using foraminiferal indices. The development of Foram-AMBI (Alve et al. 2016,  
606 Jorissen et al. 2018) is one step towards such a system.

607

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898 **Figure Captions**

899

900 Figure 1: Map of the sampling stations

901

902 Figure 2: Principal components analysis (PCA) of environmental variables: biplot of stations  
903 and variables for axes 1 and 2 (75 % of variation). Stations with one or more missing values  
904 have been omitted from analysis.

905

906 Figure 3: Redundancy analysis (RDA) of foraminiferal species assemblages: triplot of species  
907 (crosses), stations (dots) and significant environmental variables (vectors). Most dominant  
908 species are indicated by abbreviated names. Stations with missing data for environmental  
909 variables or strongly impoverished fauna have been omitted from analysis.

910

911 Figure 4: Canonical correspondence analysis (CCA) of macrofaunal species assemblages:  
912 triplot of species (crosses), stations (dots) and significant environmental variables (vectors).  
913 Most dominant species are indicated by abbreviated names. Stations with missing data for  
914 environmental variables or strongly impoverished fauna have been omitted from analysis.

915

916 Figure 5: Abundances (ind.m<sup>-2</sup>) of the main foraminiferal species at stations along (A) the  
917 TOC-gradient (%) and (B) the bottom-water dissolved oxygen concentration-gradient (mL  
918 O<sub>2</sub>.L<sup>-1</sup>). Definition of low (<2.7%), medium (2.7-3.4%) and high (>3.4%) level of TOC follows  
919 Molvaer et al. (1997). Definition of normoxia (bottom-water [O<sub>2</sub>] > 2 mL O<sub>2</sub>.L<sup>-1</sup>), hypoxia (0.5  
920 - 2 mL O<sub>2</sub>.L<sup>-1</sup>), severe hypoxia (< 0.5 mL O<sub>2</sub>.L<sup>-1</sup>) follows Diaz & Rosenberg (1995). Please note:  
921 different scales on the x-axes.

922

923 Figure 6: Abundances ( $\text{ind.m}^{-2}$ ) of the main macrofaunal species at stations along (A) the  
924 TOC-gradient (%) and (B) the bottom-water dissolved oxygen concentration-gradient ( $\text{mL}$   
925  $\text{O}_2\cdot\text{L}^{-1}$ ). Definition of low (<2.7%), medium (2.7-3.4%) and high (>3.4%) level of TOC follows  
926 Molvaer et al. (1997). Definition of normoxia (bottom-water  $[\text{O}_2] > 2 \text{ mL O}_2\cdot\text{L}^{-1}$ ), hypoxia ( $0.5$   
927  $- 2 \text{ mL O}_2\cdot\text{L}^{-1}$ ) and severe hypoxia ( $< 0.5 \text{ mL O}_2\cdot\text{L}^{-1}$ ) follows Diaz & Rosenberg (1995). Please  
928 note: different scales on the x-axes.

929

930 Figure 7: Biplot based on predictive co-correspondence analysis (CoCA) of foraminifera (left)  
931 against macrofauna (right) showing 49% and 24% of the total variance in the foraminiferal  
932 and macrofaunal data, respectively.

933

#### 934 **Table captions**

935

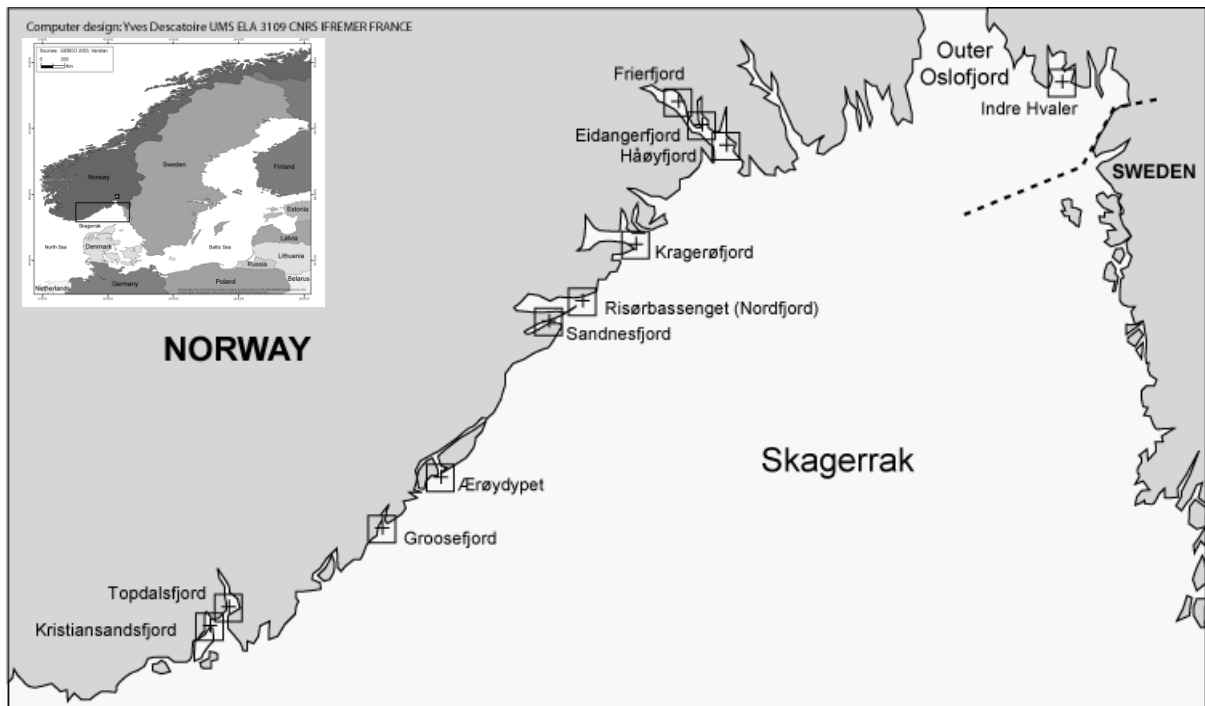
936 Table 1: Characteristics of sampling stations (nd: no data): water depth, depth below  
937 threshold (Norwegian Hydrographic Service), bottom-water dissolved  $\text{O}_2$ -concentration at  
938 the time of sampling ( $[\text{O}_2]_{\text{tos}}$ ,  $\text{mL O}_2\cdot\text{L}^{-1}$ ), sediment grain size ( $\%<63\mu\text{m}$ ), total organic carbon  
939 (TOC, %), Total nitrogen (TN, %) and C/N ratio of sampling stations. For location of areas, see  
940 Fig. 1.

941

942 Table 2: Suggested new assignments into EGs for foraminifera and macrofauna species  
943 according to Alve et al (2016) and Borja et al. (2000), respectively (na: not assigned).

944

945 Figure1

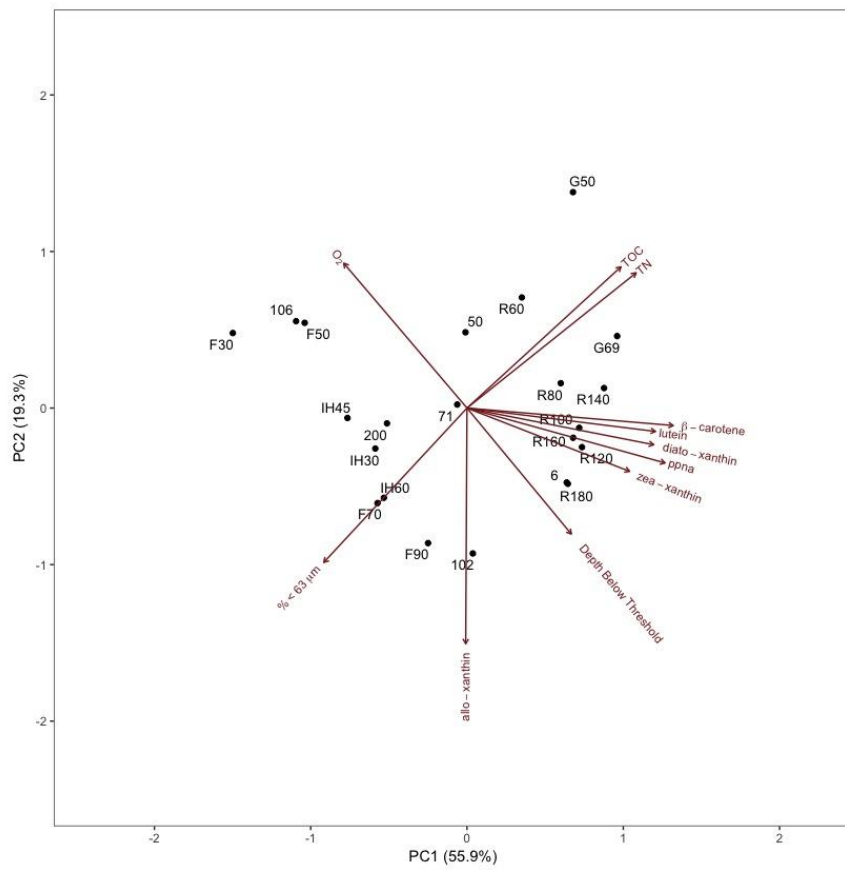


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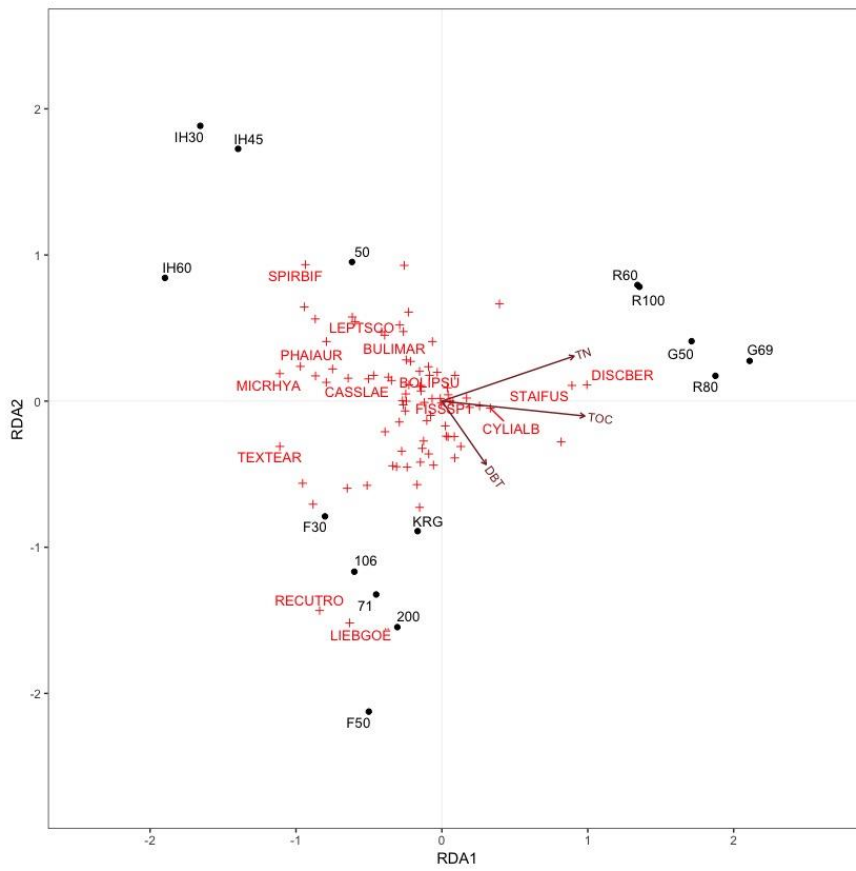
949 Figure2



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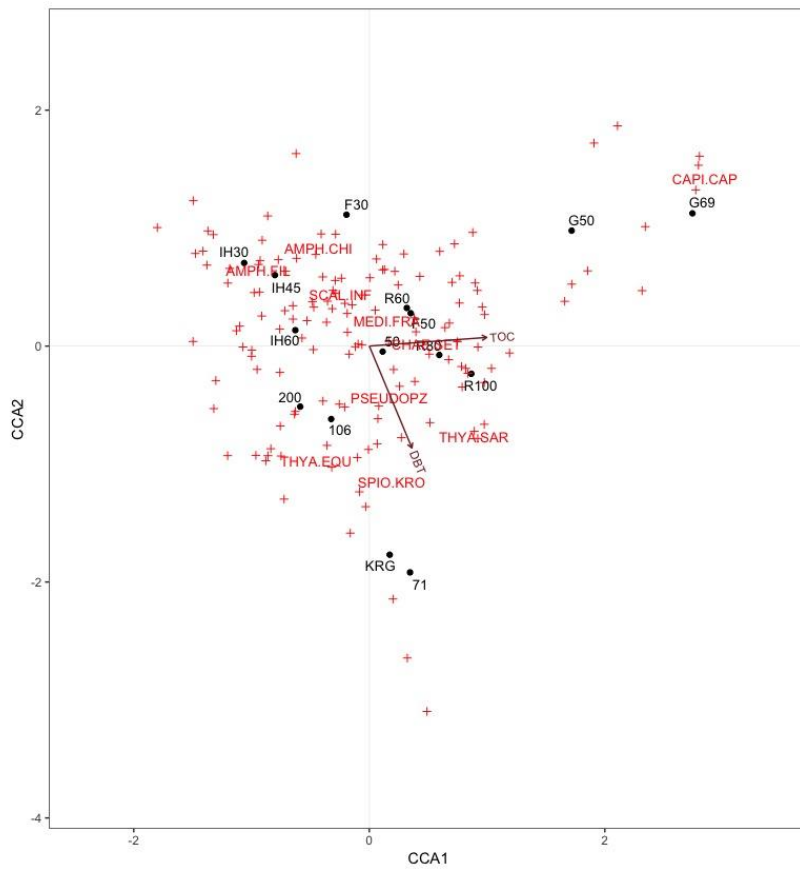
952 Figure3



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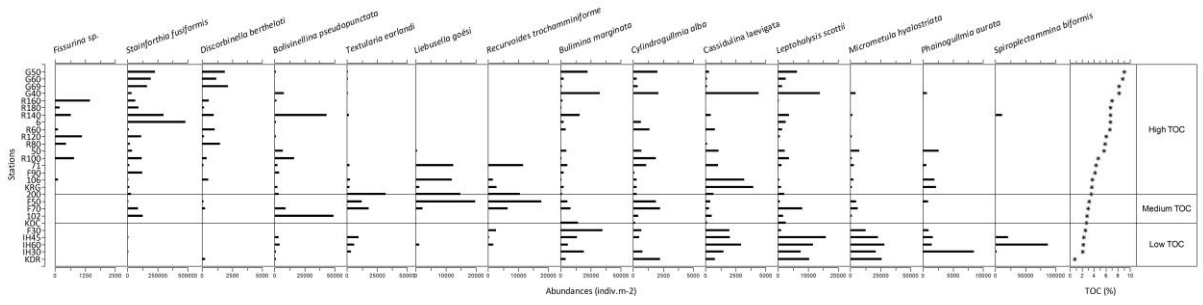
955 Figure4



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958 Figure5A

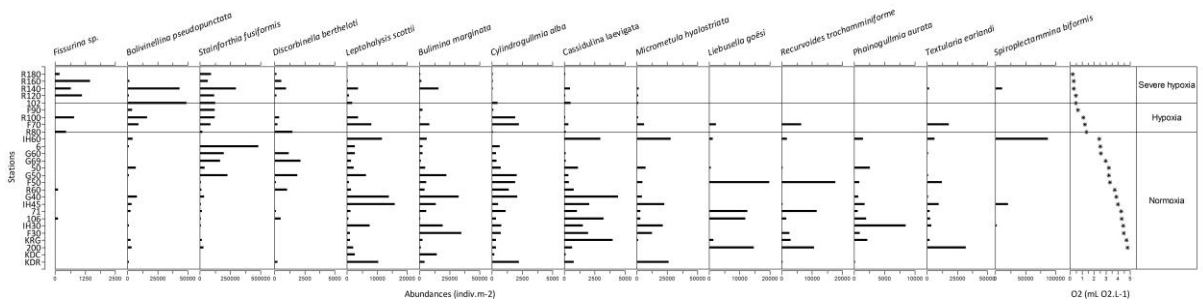


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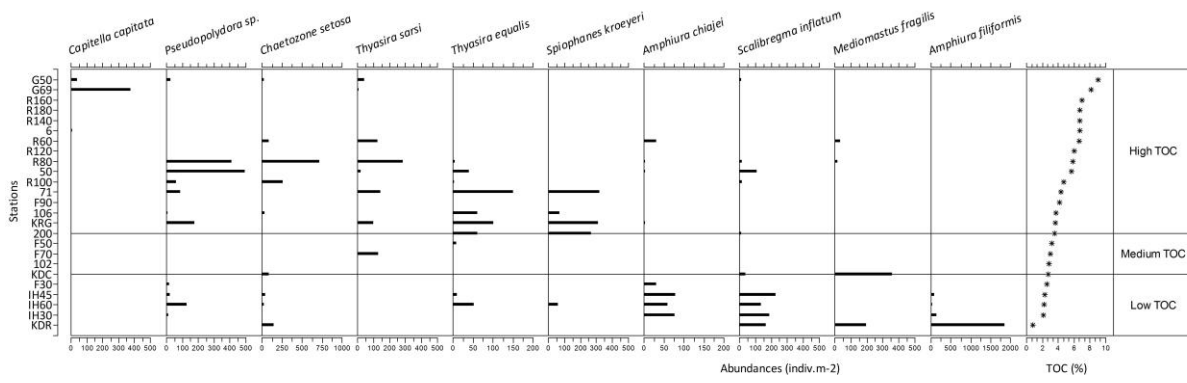
962 Figure5B



963

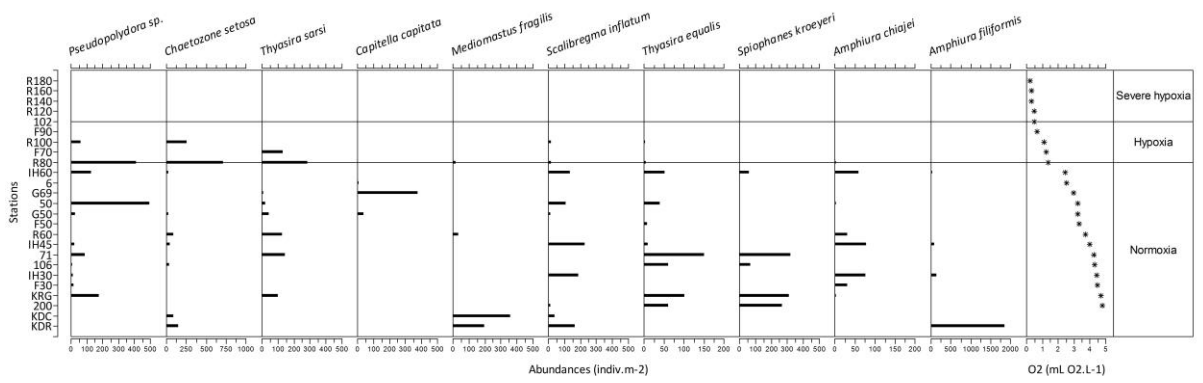
964

965 Figure6A



966

967 Figure6B

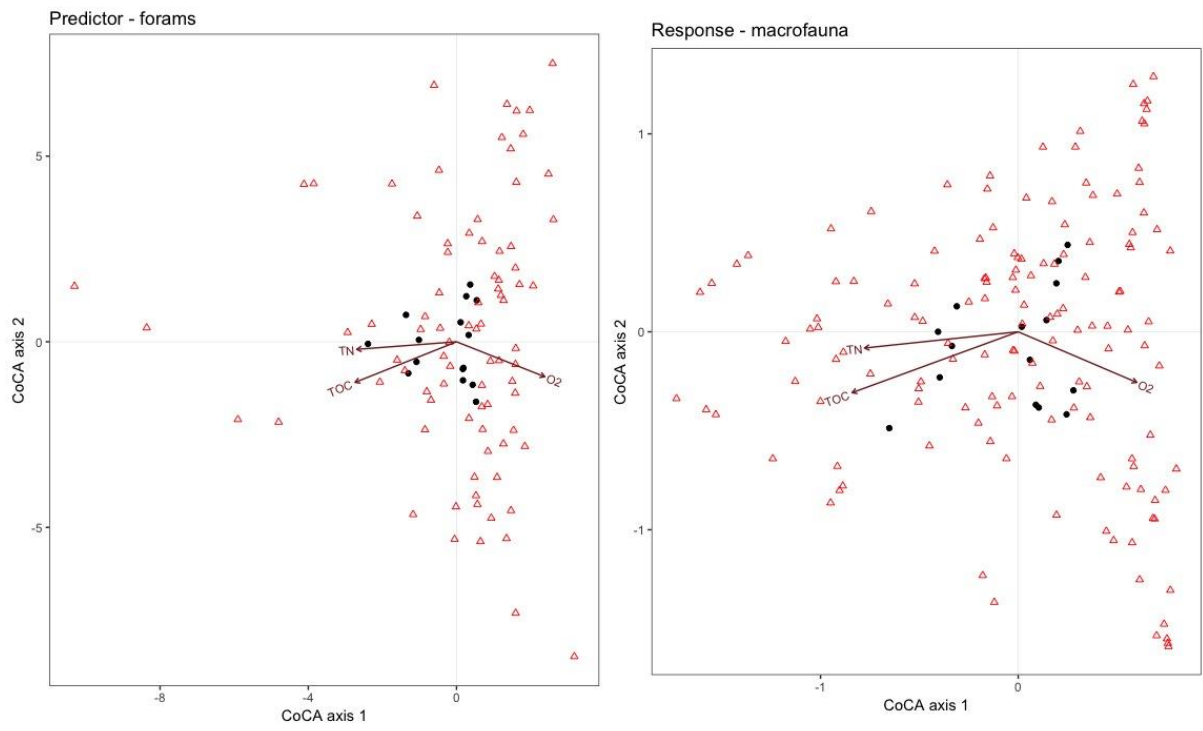


968

969



970 Figure7



971

972

973 Table1

	Station	Water depth (m) at stations	Depth below threshold (m)	[O <sub>2</sub> ] <sub>tos</sub>	Grain size	TOC	TN	C/N
Ærøydypet	200	111	59	4.76	93	3.5	0.4	9.7
Kragerøfjord	71	138	118	4.25	98	4.3	0.5	7.9
	KRG	102	79	4.67	94	3.6	0.5	7.2
Sandnesfjord	50	65	42	3.21	92	5.7	0.4	12.6
Eidangerfjord	106	103	43	4.31	96	3.7	0.2	15.5
Risør bassenget	R60	60	35	3.73	83	6.6	0.9	7.4
	R80	80	56	1.36	89	5.8	0.6	10.1
	R100	104	75	1.09	87	4.7	0.6	8.1
	R120	124	99	0.47	90	6.0	0.6	10.5
	R140	142	118	0.33	83	6.7	0.6	10.7
	R160	157	134	0.29	91	7.0	0.7	10.4
	R180	182	157	0.20	91	6.7	0.7	10.1
Håøyfjord	102	204	169	0.49	98	2.8	0.3	9.7
Topdalsfjord	6	74	42	2.52	94	6.7	0.5	12.5
Groosefjord	G40	40	17	3.85	nd	8.1	0.6	12.9
	G50	54	31	3.25	84	9.0	0.7	12.5
	G60	60	37	2.58	nd	8.8	0.7	11.7
	G69	69	47	2.95	93	8.1	0.7	10.7
Frierfjord	F30	28	8	4.48	98	2.6	0.1	32.2
	F50	52	32	3.30	92	3.2	0.2	13.3
	F70	70	50	1.25	98	3.0	0.1	21.1
	F90	91	70	0.67	98	4.2	0.2	17.8
Indre Hvaler	IH30	30	6	4.44	98	2.1	0.1	14.2
	IH45	45	20	3.98	98	2.3	0.2	14.3
	IH60	62	37	2.42	98	2.2	0.2	13.2
Kristiansandsfjord	KDR	23	nd	nd	58	0.8	0.1	14.6
	KDC	31		nd	52	2.7	0.2	14.6

974

975 Table2

FORAMINIFERA	This study EG	Foram-AMBI EG
<i>Micrometula hyalostriata</i>	1	na
<i>Phainogullmia aurata</i>	1	na
<i>Cylindrogullmia alba</i>	2	na
<i>Leptohalysis scottii</i>	4	na
<i>Cassidulina laevigata</i>	2	1
<i>Liebusella goësi</i>	3	2
<i>Bolivinellina pseudopunctata</i>	4	2
MACROFAUNA	This study EG	AMBI-list EG
<i>Scalibregma inflatum</i>	1	3
<i>Thyasira cf. sarsii</i>	4 or 5	3

976