Accepted Manuscript

This is an Accepted Manuscript of the following article:

Brij Mohan Sharma, Jitka Bečanová, Martin Scheringer, Anežka Sharma, Girija K. Bharat, Paul G. Whitehead, Jana Klánová, Luca Nizzetto. Health and ecological risk assessment of emerging contaminants (pharmaceuticals, personal care products, and artificial sweeteners) in surface and groundwater (drinking water) in the Ganges River Basin, India. Science of The Total Environment. 646, 2019, 1459-1467, ISSN 1879-1026.

The article has been published in final form by Elsevier at http://dx.doi.org/10.1016/j.scitotenv.2018.07.235

© 2018. This manuscript version is made available under the

CC-BY-NC-ND 4.0 license

http://creativecommons.org/licenses/by-nc-nd/4.0/

Health and ecological risk assessment to emerging
 contaminants (pharmaceuticals, personal care products, and
 artificial sweeteners) in surface and groundwater (drinking
 water) in the Ganges River Basin, India

Brij Mohan Sharma ^{a,*}, Jitka Bečanová ^{a,b}, Martin Scheringer ^{a,c},
 Anežka Sharma ^a, Girija K. Bharat ^{d,e}, Paul G. Whitehead ^f, Jana
 Klánová ^a, Luca Nizzetto ^{a,g}

- ^a Research Centre for Toxic Compounds in the Environment (RECETOX), Masaryk
 ^y University, Brno 62500, Czech Republic
- ^b Graduate School of Oceanography, University of Rhode Island, RI 02882, USA
- ¹¹ ^c Institute for Biogeochemistry and Pollutant Dynamics, ETH Zürich, 8092 Zürich,
- 12 Switzerland
- ¹³ ^d Mu Gamma Consultants Pvt. Ltd., Sector-50, Gurgaon, Haryana 122018, India
- ¹⁴ ^e The Energy and Resources Institute (TERI), Darbari Seth Block, India Habitat Centre,
- 15 Lodhi Road, New Delhi 110003, India
- ¹⁶ ^f School of Geography and the Environment, University of Oxford, Oxford OX1 3QY,
- 17 United Kingdom
- ⁹ Norwegian Institute for Water Research (NIVA), Gaustadalleen 21, Oslo 0349, Norway
- 19
- 20 <u>Corresponding author's Email</u>:
- 21 <u>sharma@recetox.muni.cz</u>, <u>brijmsharma05@gmail.com</u> (Brij Mohan Sharma)
- 22
- 23

24 1. Introduction

Several pharmaceuticals and personal care Products (PPCPs) as well as artificial sweeteners 25 (ASWs) are known as contaminants of emerging concerns due to their frequent detection in 26 27 environmental samples (Brack et al., 2012; Petrie et al., 2014). Often, residues of these contaminants are collected in wastewater effluents after their consumption and due to their low 28 29 removal in wastewater treatment plants (WWTPs), these compounds enter freshwater ecosystems. In several cases where WWTPs are not existing, these substances enter the 30 31 aqueous environment directly following human and veterinary excretion and municipal 32 wastewater effluents. Their environmental (i.e. in soils and aquatic environment) occurrence has been pointed as a direct indicator of wastewater-derived pollution (Kümmerer and Henninger, 33 2003), especially some ASWs such as sucralose and acesulfame K that are highly soluble and 34 35 stable in the environment. During the last two decades, numerous studies have documented the occurrence of PPCPs and ASWs in wastewater and recipient water from many regions (Boxall et 36 37 al., 2012; Sui et al., 2015; Y. Yang et al., 2017). Presence of the pharmaceuticals in groundwater could foster the dissemination of antibiotic resistance genes, which may interact with human 38 39 intestinal flora and spread the resistance determinants, potentially impacting the human health 40 (Szekeres et al., 2018). On the other hand, irrigation using water containing pharmaceuticals could imply that crops may take up these compounds, being another route of human exposure 41 42 (Miller et al., 2016).

43 As some of the PPCPs are biologically active at low concentrations and have potential to accumulate in aquatic organisms (Brausch and Rand, 2011; Lillicrap et al., 2011; Tanoue et al., 44 2015), occurrence of these contaminants in drinking and irrigation water may pose health 45 concerns and a water management challenge in many regions. Areas suffering drinking water 46 47 scarcity and poor wastewater management are particularly sensitive to emerging contaminants. This is typically the case of developing countries with transitional economies, growing urban 48 populations, insufficient pollution control infrastructures, and subject to frequent draught 49 periods. 50

51 The pharmaceutical sector in India has seen a large growth in the last few decades and now ranks globally 3rd by volume, accounting 10% of the global pharmaceutical production 52 (Department of Pharmaceuticals. Government of India, 2018). Increasing access to therapeutic 53 54 drugs, use of veterinary drugs in intensive animal farming and industrially-processed food, and 55 feeds represent drivers of high emission of PPCPs and ASWs to recipient waters as well as, potentially, in drinking water resources. Water resources scarcity, poor or absent wastewater 56 57 management and high demand for irrigation water can also result in transferring a significant 58 contaminant load to farms and possibly in the food production chain. Data supporting this 59 hypothesis in developing countries including India are rare as monitoring of contaminants of 60 emerging concern is not implemented on a routine base due to high cost and needs of in-place 61 capacity.

In India, increasing demand for drinking water, poor management of available water resources, 62 63 and unreliable water supply due to both anthropogenic and climate factors cause a virtually chronic water crisis (Natarajan et al., 2016; Thatte, 2018). While the protection of available 64 65 drinking water resources from pollution is crucial, the process of building resilience in safe 66 drinking water supply still requires essential base-line information. Only eight studies have 67 monitored PPCPs in river water and two in groundwater in India, while no study monitored ASWs (Balakrishna et al., 2017; Fick et al., 2009; Philip et al., 2018). Available studies from India 68 describe the results of local surveys, while works with a larger spatial breadth are still missing. 69

In this study, we examined the presence of selected PPCPs and ASWs in the surface and 70 71 groundwater in the largest river basin in India, the Ganges River Basin (GRB). It is home of 72 about 7% of the global human population and several industries are located within the 73 watershed. For the last few decades, several mega-cities and semi-urban areas in the GRB have 74 been experiencing serious water pollution issues and drinking water supply scarcity (Chakraborti et al., 2018; Natarajan et al., 2016). More than 60% of the irrigated agriculture and 85% of the 75 76 drinking water supplies depend on the groundwater resources in India (World Bank, 2010) and 77 contamination of groundwater aguifers may turn into a potential threat to the health of millions 78 of people in India. The specific objectives in this study are to: (1) assess the contamination 79 profile of selected PPCPs and ASWs in the Ganges River water acting as recipient of municipal 80 wastewater (partially or mostly untreated) and groundwater resources in its proximity, and (2) evaluate the human health and ecological risks associated with existence of PPCPs and ASWs in 81 82 drinking water and river water, respectively.

83 **2. Materials and methods**

2.1. Description of study area and sampling

The Ganges River is the largest river in India and along with the Brahmaputra and Meghna Rivers 85 86 the third largest in the world in terms of water discharge. Along the 2 525 km long course in 87 India, the Ganges crosses a steep environmental and socioeconomic gradient. Its average annual discharge is 12 400 m³/s and the hydrological basin covers 861 452 km² (MoWR, 2014; UNESCO, 88 89 1971). Surface water and groundwater resources of the GRB are extensively used to support 90 the livelihood of 43% of the Indian population through irrigation, provision of drinking water and of water for industrial purposes, ultimately contributing to 40% of India's gross domestic 91 product. There are around 764 industries and 36 class I cities (population > 100 000) situated 92 along the Ganges River (Narain, 2014). An estimated 1.4 x 10⁶ m³/day of mostly untreated 93 domestic wastewater and 0.26 x 10⁶ m³/day of industrial sewage are discharged into the Ganges 94 River and its tributaries (Natarajan et al., 2016). Along the Ganges River channel, there are 95 about 27 chemical plants including production of fertilizers, pesticides, and pharmaceuticals 96 97 which generate about 98 x 10³ m³/day of wastewater. Chemical industries along with sugar and 98 pulp industries generate 79% of the total industrial wastewater along the Ganges River (CPCB 99 (Central Pollution Control Board), 2013).

The GRB can be divided into three reaches representing different ecological and socioeconomic 100 101 conditions: the Himalayan Reach (HR, mostly rural and semiurban population), Middle Reach (MR, mostly urban, growing cities and industries), and Lower Reach (LR, mostly semirural and 102 rural, and industries). River water and groundwater samples were collected from 9 locations (4 103 104 in the HR, 2 in MR, and 3 in LR) (figure 1). To capture local contamination patterns, samples upstream and downstream of major cities were collected from 5 of the 9 locations (namely UK, 105 106 KP, VS, PT, and FK, see figure 1 caption for location legend). In total, 14 sampling sites were included. Groundwater was collected from handpumps (used as drinking water sources) by local 107 communities. Sampling locations for groundwater were positioned within 5 km from the Ganges 108 109 River. Sample collection took place during February-April 2014 in dry weather conditions (defined as no rain in the previous 24 h and less than 2 mm in the previous of 48 h (Tran et al., 110 2014). Collected samples were kept in an icebox and transported to The Energy and Resources 111 Institute (TERI) laboratory where they were temporarily stored frozen until they were shipped 112 (cooled at 4 °C) to the Research Centre for Toxic Compounds in the Environment (RECETOX) 113 laboratory for chemical analysis. Further details about the sampling campaign are provided in 114 115 supplementary data (Table S1) and in a previous work (Sharma et al., 2016).



Figure 1. Sampling locations in the GRB. Sampling locations are: UKU/D = Uttarkashi upstream/downstream, DAK = Devprayag along Alaknanda River, DBG = Devprayag along Bhagirathi River, DGR = Devprayag along Ganges River, KPU/D = Kanpur upstream/downstream, VSU/D = Varanasi upstream/downstream, PTU/D = Patna upstream/downstream, FKU/D = Farakka upstream/downstream, GSR = Ganga Sagar.

122 **2.2. Sample preparation and extraction**

Collected water samples were stored in RECETOX laboratories at -25° C until analysis and 123 processed in agreement with previous work (Sharma et al., 2016). All samples and procedural 124 125 blanks were filtered prior to the extraction (70 mm Whatman GF/C filter with pore size 1.2 µm) 126 and spiked with 20 µL of surrogate standard (C13 caffeine; 400 ng/mL, used for recovery control). The extraction was performed using solid phase extraction column (Waters[®] Oasis HLB 127 6cc/150mg). Cartridges were conditioned with 4 mL of 0.1% ammonium hydroxide in methanol, 128 129 4 mL of methanol, and 4 mL of Milli-Q water. Water samples (pH adjusted to 2 – 3) were loaded into the pre-conditioned cartridge at a flow rate ranging between 3 and 6 mL/min under 130 moderate vacuum. After extraction, cartridges were dried for 30 minutes under vacuum in a 131 132 protected atmosphere and washed with acetate buffer (4 mL, 0.025M). Pharmaceuticals and 133 artificial sweeteners were eluted with 4 mL methanol into the falcon tubes and concentrated to 134 about 250 μ L (exactly weighted) using a gentle stream of nitrogen in TurboVap II (Caliper 135 LifeSciences, USA) concentrator. Prior to the analysis, two aliquots of each extract (10 and 50 136 μ L) were diluted with ammonium acetate in water (5mM) to get a final volume of 100 μ L and 137 the content of either 10% (for ASWs and PPCPs ionized with ESI+) or 50% (for PPCPs ionized 138 with ESI-) of methanol in the sample to reach the initial mobile phase content use for different target method. 139

140 **2.3. Target compounds and chemical analysis**

In total, 12 pharmaceuticals (acetaminophen, atenolol, caffeine, carbamazepine, ciprofloxacin, 141 142 clofibric acid, diclofenac, hydrochlorothiazide, ibuprofen, ketoprofen, naproxen, and sulfamethoxazole), three personal care products (diethyltoluamide (DEET), triclocarban, 143 144 triclosan) and five artificial sweeteners (acetsulfamate K, aspartame, cyclamate, saccharine, sucralose) were analyzed in the collected river and groundwater samples. The selection of these 145 146 PPCPs and ASWs in this study was based on their popularity in selected previous studies from 147 India and in other countries, and their availability on the Indian market. In addition, selection of 148 PPCPs and ASWs for analysis was also influenced by the availability of analytical methods used 149 in the trace laboratories of RECETOX where the collected samples were processed and analyzed.

ASWs were separated using an ultra-performance liquid chromatograph (UPLC ACQUITY, 150 151 Waters[®], Milford, MA, USA) equipped with BEH C18 (100x 2.1 mm, 1.7 µm, 130 Å) column (Waters, Milford, MA, USA). Both water and methanol used as mobile phases contained 0.1% 152 formic acid. Gradient elution with an initial content of 10% of methanol was applied and the final 153 154 content of methanol (90%) was reached in 5 minutes. The flow rate of the mobile phase was 155 0.4 mL/min. The injection volume was 10 µL per individual sample. Before the next separation, the column was equilibrated using the initial composition of the mobile phase for 3 minutes. The 156 157 mass spectrometer (Xevo TQS, Waters[®], Milford, MA, USA) was operated in negative ion mode (ESI-). Quantification of analytes was based on an external calibration using freshly prepared 158

standards with a range of 0.05–500 ng/mL (10 points). The mass-labelled standard (sucralosse-d3) was used for matrix effects evaluation and sucralose quantification.

For the analysis of the first fraction of the PPCPs (acetaminophen, atenolol, caffeine, 161 162 carbamazepine, ciprofloxacine, DEET, diclofenac, sulfamethoxazole), the same column as used for the ASWs was used with methanol and water as mobile phase, both containing 0.01% formic 163 acid and 0.1M ammonium acetate. The separation gradient, flow rate of mobile phase and 164 injected volume were as described above for ASWs. The mass spectrometer was operated in 165 166 positive ion mode (ESI+). Quantification of analytes was based on an external calibration using 167 freshly prepared standards (10 points) with a range of 0.02-100 ng/mL (0.002-10 ng/mL for carbamazepine). The mass-labelled standards (paracetamol-d4 and sulfamethoxazole-d4) were 168 used for matrix effects evaluation. 169

The second fraction of PPCPs (clofibric acid, hydrochlorothiazide, ibuprofen, ketoprofen, 170 naproxen, triclocarbon, triclosan) was separated using an Xterra C18 (100 x 2.1 mm, 3,5µm) 171 column (Waters, Milford, MA, USA). Water containing 0.1% acetic acid and 0.1% ammonium 172 acetate and a mixture of methanol and acetonitrile (50:50) were used as mobile phases. The 173 174 initial gradient was set at 40:60 organic:water, in ten minutes the content of the organic mixture 175 was increased up to 100 percent (hold for 2 min). The flow rate of the mobile phase was 0.2 176 mL/min. The injection volume was 10 µL per individual sample. Before the next run, the column was equilibrated using the initial composition of the mobile phase for 4 minutes. The mass 177 178 spectrometer was operated in negative mode (ESI-). Quantification of analytes was based on 179 an external calibration using freshly prepared standards (10 points) with a range of 0.1 – 500 ng/mL. The mass labelled standards (ibuprofen-d3 and 13C6-triclosan) were used for 180 matrix effects evaluation. Mass-labelled standards were added to the extracts prior to the 181 182 instrumental analyses and were used for matrix control. If the response of mass-labelled 183 standards dropped below the threshold (60% of response in calibration) the samples were 184 diluted and re-analyzed to minimalize the matrix effects.

185 **2.4.** Quality assurance/quality control (QA/QC)

To ensure no significant contamination occurred during transport of samples, travel blanks were 186 analyzed during a pilot study prior to the campaign. Travel blanks were obtained from a previous 187 sampling campaign (in which marine water samples originating from pristine open ocean areas 188 189 in the South Atlantic were collected). These were transferred in the same type of bottles and 190 travel conditions used for the field campaign in this study and transported sealed during several 191 days using commercial courier-express services. Concentration of PPCPs and ASWs measured in the travel blanks (n=3) (reported in (Brumovský et al., 2017)) were similar to those measured 192 193 in laboratory procedural blanks, suggesting no significant contamination occurred during 194 transport. Travel blanks results are provided in Table S4.

Procedural blanks (n=3; SPE cartridges without any loaded samples) were processed under identical laboratory conditions as the field samples and used to control potential contamination during analysis. In addition, to control LC-MS instrument sensitivity, a QA/QC sample was analyzed after each batch of 10 samples.

199 Method detection limits (MDLs) were calculated as the average of the individual compounds signals in the procedural blanks plus 3 times their standard deviation (SD). These signals were 200 201 considered significant if they exceeded a threshold of 3 in the signal-to-noise ratio. For analytes which were not detected in blanks, MDLs were calculated as the concentration in samples with 202 a signal-to-noise ratio equal to 3. The MDL values calculated from the procedural blank 203 contamination were then compared with those obtained from the signal to noise ratio in the 204 205 samples and the highest were chosen. The average MDLs (Table S7) were found between 0.06-15.0 ng/L and 0.5–2.0 ng/L for PPCPs and ASWs, respectively. The final results were blank-206 207 corrected using the average concentration of the target compounds in the procedural blanks. Reported concentrations of PPCPs and ASWs were not corrected for recovery. Concentrations of 208 209 target analytes in procedural and field blanks are provided in Table S4. Recovery tests (n=10)210 were performed using spiked Milli-Q water with addition of target analytes at levels 20–200 ng/L 211 (Table S5). Recoveries of most compounds ranged from 54% to 125%. In contrast, low but 212 consistent recoveries were observed for acetsulfamate K $(28\pm6\%)$ and cyclamate $(20\pm4\%)$.

213 Measures for quality assurance and control have been described in detail by Brumovský et al.
214 (Brumovský et al., 2017) and are also reported in the SI.

215 **2.5. Estimation of human health and ecological risk**

We estimated the worst-case scenario of potential health risk of the detected PPCPs in the groundwater (i.e. drinking water) along the Ganges River. We calculated an age-dependent risk quotient (RQ) for each detected PPCP by dividing the maximum measured concentration in the groundwater (MC_{GW}) by the corresponding age-dependent drinking water equivalent level (DWEL) (Eq. 1). Age-specific assessment of exposures has been previously used to reduce uncertainty in risk assessment (de Jesus Gaffney et al., 2015; Leung et al., 2013; Yun Ya Yang et al., 2017).

$$RQ = \frac{MC_{GW}}{DWEL}.$$
 (1)

Often, it is easier to convert the acceptable daily intake (ADI) into a corresponding water concentration, such as DWEL, so that the comparison of chemical concentrations measured in drinking water to ADIs is simpler. The DWEL was estimated for seven age categories (from 1 year to >21 years of age) by using equation 2.

228
$$DWEL = \frac{ADI(or RSD)*BW}{DWI*AB*FOE}$$
.....(2)

229 Where ADI (µg/kg/day) is the acceptable daily intake or risk specific dose (RSD) for 230 noncarcinogenic and carcinogenic effects, respectively. Values of ADI (or RSD) for each detected PPCP and ASW were adopted from the literature (Leung et al., 2013; Yun Ya Yang et al., 2017) 231 232 (Table S9). BW is the median body weight (kg) of age-specific groups (Table S10), DWI is the daily drinking water intake (L/day) of age-specific groups (Table S10), AB is the gastrointestinal 233 234 absorption rate assumed to be 1, and FOE is related to the frequency of exposure (350 days/365 235 days)(de Jesus Gaffney et al., 2015; Yun Ya Yang et al., 2017). A RQ value greater than 1 236 indicated the possibility of human health risk. A RQ value between 0.2 and 1 calls for more detailed assessment, whereas RQ \leq 0.2 is considered of no appreciable concern to human health 237 (Schriks et al., 2010; Yun Ya Yang et al., 2017). 238

The ecological risk assessment was performed by calculating RQ for the detected PPCPs and ASWs in the river water, as described in a previous publication (Yun Ya Yang et al., 2017). It was calculated by dividing the maximum river water concentration (MC_{RW}) for each PPCP and ASW by the corresponding predicted no effect concentration (PNEC) for three classes of aquatic organisms (i.e. algae, *Daphnia Magna*, and Fish) (Eq. 3).

$$RQ = \frac{MC_{RW}}{PNEC}$$
(3)

245 PNEC was calculated as:

246

Where the EC₅₀ (effective concentration, reducing a biological process by 50%) or LC₅₀ (lethal 247 concentration, killing 50% the organisms) was obtained from the literature or by using the 248 US EPA Ecological Structure Activity Relationship (ECOSAR v1.10) model. For PPCPs or ASWs 249 (namely: atenolol, caffeine, ciprofloxacin, hydrochlorothiazide, cyclamate, saccharine, 250 sucralose), more than one toxicity values (EC_{50}/LC_{50} value) were available from ECOSAR model. 251 252 For these contaminants, baseline toxicity values were chosen as a precaution measure. A 253 summary of the EC₅₀/LC₅₀ values is provided in the Table S12. AF, a standard assessment factor 254 with a value of 1000, was introduced to account for extrapolation from intra- as well as interspecies variability in sensitivity (Hernando et al., 2006). Risk to aquatic organisms was 255 subsequently classified into three categories: Low risk (RQ<0.1), moderate risk (0.1<RQ<1), 256 257 and high risk (\geq 1) (de Souza et al., 2009; Hernando et al., 2006).

258 **3. Results**

259 **3.1. Distribution of PPCPs and ASWs in the Ganges River water**

260 Of the 15 target PPCPs, 14 were detected in the Ganges River water at one or more sampling 261 sites. Atenolol and ibuprofen were detected at only one sampling site, while clofibric acid was 262 not detected at any sampling site. Caffeine, DEET, and ketoprofen were detected with 100% 263 detection frequency. The sum of detected PPCPs in the river water (Σ PPCPs_{RW}) at the monitored 264 sampling sites ranged between 54.7–826 ng/L. The highest Σ PPCPs_{RW} was found in the lower reach of the Ganges River. The highest concentration among the PPCPs was found for caffeine (743 ng/L), followed by ketoprofen (107 ng/L). High concentrations of caffeine were found at sampling sites in Himalayan reach. Except caffeine, the other PPCPs were generally higher in the middle and lower reach of the Ganges River compared to the Himalayan reach. At local level, as expected, concentrations of PPCPs were generally higher at the downstream sites of major cities such as Kanpur, Varanasi, and Patna. Concentrations of the frequently detected PPCPs in the Ganges River water are depicted in figure 2A and presented in Table S7.

Of the five ASWs, only three (cyclamate, saccharine, and sucralose) were detected in the 272 Ganges River water at more than one sampling sites. Cyclamate and sucralose were detected 273 with 79% frequency. The highest concentration among these frequently detected ASWs was 274 found for saccharine, followed by sucralose and cyclamate. In river water, concentrations of 275 saccharine and sucralose ranged between 2.4–85 ng/L and 0.5–24 ng/L, respectively. 276 277 Concentrations of saccharine and sucralose clearly peaked in the middle and lower reach of the Ganges River. Concentrations of cyclamate ranged between 0.2–1.2 ng/L with elevated 278 279 concentrations in the Himalayan and lower reach. Acetsulfamate K was detected only at one 280 location in the lower reach. The sum of detected ASWs in river water (ΣASWs_{RW}) at monitored sampling sites ranged between 0.2–102 ng/L. Similar to SPPCPs_{RW}, the highest SASWs_{RW} was 281 282 detected at downstream of Patna in the lower reach. Concentrations of ASWs detected in the 283 Ganges River water are depicted in figure 2B and presented in Table S8.



284

Figure 2. Panels A and C depict concentrations of frequently PPCPs in the Ganges River water and groundwater from the vicinity of the river channel, respectively. Caffeine is displayed by yellow bars corresponding to the secondary y-axis and it is not part of stacked bars in panels A and C. Panel B and D depict concentrations of frequently detected ASWs in Ganges River water and in groundwater from the vicinity of the river channel, respectively.

3.2. Distribution of PPCPs and ASWs in groundwater

291 Thirteen out of the 15 target PPCPs were detected in the groundwater at one or more sampling 292 sites. Atenolol and clofibric acid were not detected at any sampling sites. Similar to river water, 293 caffeine and DEET were detected with 100% frequency. The sum of detected PPCPs in 294 groundwater (Σ PPCPs_{GW}) ranged between 34–293 ng/L, with elevated concentrations observed 295 in the middle and lower reaches of the Ganges River. $\Sigma PPCPs_{GW}$ were about a factor of 2 296 (geometric mean) lower than those found in the river water. However, at few sampling sites, 297 Σ PPCPs_{GW} was found higher than the Σ PPCPs_{RW}. Similar to the river water, highest concentrations 298 in groundwater were detected for caffeine, ranging from 15–262 ng/L. Other elevated PPCPs in 299 groundwater were ibuprofen (<MDL-49.4 ng/L), carbamazapine (<MDL-27.2 ng/L), and 300 ketoprofen (<MDL-23.4 ng/L). Unlike river water, PPCPs in groundwater did not consistently 301 display higher concentrations in wells located downstream the major cities. Concentrations of 302 selected PPCPs in the groundwater along the Ganges River are depicted in figure 2C and 303 presented in Table S7.

304 Similar to river water, cyclamate and sucralose in groundwater were detected at more than one 305 sampling site. Sucralose was detected in groundwater with 100% frequency and ranged between 0.5-25 ng/L. Groundwater concentrations of cyclamate ranged between <MDL-0.26 ng/L. 306 307 Results for cyclamate have however been taken cautiously due to poor recoveries (Table S5). The detection frequency of sucralose in groundwater was higher than that in the river water, 308 309 whereas that of cyclamate was 50% and lower than that observed in river water (78%). 310 Interestingly, similar to river water, acetsulfamate K was detected in groundwater only at one location in the Himalayan reach. The sum of detected ASWs in the groundwater ($\Sigma ASWs_{GW}$) 311 312 ranged between 0.5–27 ng/L. Except for one sampling site (UKD) in the Himalayan reach, levels of detected ASWs in groundwater were elevated in the middle and lower reaches of the Ganges 313 314 River. Concentrations of ASWs in the groundwater are depicted in figure 2D and presented in 315 Table S8.

Generally, it is expected that PPCPs and ASWs with low logK_{ow} would have a tendency to be present in groundwater, but in this study, no clear relationship was observed between logK_{ow} and either the frequency of detection of PPCPs and ASWs or their concentrations in groundwater.

Pairwise/Dependent ANOVA was used for testing differences in concentrations of frequently 319 detected PPCPs and ASWs in both river and groundwater in the three different reaches of the 320 321 Ganges River. These PPCPs and ASWs selected for ANOVA were acetaminophen, DEET, caffeine, 322 carbamazepine, sulfamethoxazole, ketoprofen, hydrochlorothiazide, triclocarban, cyclamate, and sucralose. Pairwise ANOVA was chosen because of possible overlaps in PPCP and ASW 323 324 contamination sources to the river and groundwater. Significant differences (p < 0.05) between 325 concentrations in the three reaches were observed for carbamazepine (p=0.004), sulfamethoxazole (p=0.008), and sucralose (p=0.050). Carbamazepine was significantly higher 326 327 in both river and groundwater in the middle reach of the Ganges River. Similarly,

328 sulfamethoxazole was significantly higher in river water in the middle reach of the Ganges River,

but not in the groundwater. Sucralose was found significantly higher in the river water in the lower reach of the Ganges River, but not in ground water.

331 3.3. Human health and ecological risk

We provide first estimations of age-specific RQs based on the maximum detected groundwater 332 333 concentrations of PPCPs in the GRB as a worst-case scenario of human exposure. For all detected PPCPs, DWELs for all age groups ranged from 4.8 μ g/L (for carbamazepine, 1–2 years age group) 334 to 12.8 mg/L (for acetaminophen and 16–21 years age group) and RQs ranged from 1.5 x 10^{-7} 335 (for acetaminophen, 16–21 years age group) to 0.0021 (for carbamazepine, 16–21 years age 336 337 group) (Table S11). The PPCPs with higher RQs were carbamazepine, ciprofloxacin, ketoprofen, caffeine, ibuprofen, and triclosan (figure 3A, Table S11). Among different age groups, children 338 339 (1-11 years) had higher RQs than adolescents (11-21 years) and adults (>21 years). Overall, 340 RQs of all detected PPCPs were <1, implying that the detected PPCPs in groundwater in the 341 present study do not pose a risk to human health through drinking water consumption.

The RQ values (based on PNECs from Table S12) for ecological risk due to PPCPs and ASWs in 342 river water are presented in Figure 3B and Table S13. For some of the PPCPs and ASWs 343 noticeably high RQs were calculated, mainly for algae. For example, the RQ of caffeine as high 344 as 49.5 was observed for three different aquatic organisms. Similarly, the RQs of triclocarban 345 and triclosan ranged from 0.03 to 0.3 and 0.01 to 3.9, respectively. Among ASWs, the RQ values 346 up to 0.1 were observed for sucralose, particularly for algae. The RQs for freshwater 347 invertebrates (except for daphnia from triclocarbon) and fish were generally lower than 0.1, 348 349 implying negligible risk of acute/chronic toxicity to these aquatic organisms.



Figure 3. Panel A shows the human health life-stage RQs for some of the PPCPs in the drinking water (groundwater).
 Panel B shows the risk quotients (on a logarithmic scale) for PPCPs and ASWs in the river water with respect to acute toxicity for algae, daphnia, and fish.

354 **4. Discussion**

This is the first study reporting concentrations of PPCPs and ASWs in river and groundwater 355 resources in the GRB. Previously, only eight studies have reported data on PPCPs in rivers from 356 India (Balakrishna et al., 2017). Among them, only one study determined the fate and 357 occurrence of some commonly used antibiotics in the Yamuna River in Delhi, a tributary of the 358 359 Ganges (Mutiyar and Mittal, 2014). The other seven studies reported PPCPs concentrations in 360 rivers from southern India. Similarly, studies determining levels of PPCPs in groundwater in India are scarce, only two studies reported concentrations of selected pharmaceuticals in Indian 361 groundwater (Fick et al., 2009; Jindal et al., 2015), despite the fact that groundwater accounts 362 for over 85% of drinking water supplies in India (World Bank, 2010). To the best of our 363 knowledge no previous study focused on ASWs in rivers or groundwater in India. Two earlier 364 studies reported data on ASWs in WWTPs in India (Anumol et al., 2016; Subedi et al., 2015). 365

The present study shows that many PPCPs and ASWs are ubiquitous in both river and groundwater wells in proximity of the main river channel. Concentrations of selected PPCPs (diclofenac, ibuprofen, ketoprofen, naproxen) measured in this study were in similar ranges to

those reported in southern Indian rivers (Shanmugam et al., 2014). However, concentrations of 369 370 ciprofloxacin in the Ganges River were up to 6 orders of magnitude lower than those found in the Isakavagu-Nakkavagu Rivers and in southern India (ciprofloxacin: 10–2500 µg/L) and in the 371 Yamuna River (<1.44 µg/L) (Fick et al., 2009; Mutiyar and Mittal, 2014). In the present study, 372 373 triclosan in river water ranged between <MDL-5.4 ng/L and had concentrations higher than MDL 374 at only three sampling sites, whereas triclosan concentrations were found up to three order of 375 magnitude higher in rivers in southern India (Ramaswamy et al., 2011). Concentrations of 376 ciprofloxacin and diclofenac in the groundwater in the present study were up to 3 orders of magnitude lower than those observed in wells located villages of southern- and northern-India 377 (Fick et al., 2009; Jindal et al., 2015). 378

379 In a global context, various studies have reported levels of PPCPs and ASWs in river and groundwater in North America, Europe, and Asia (Ebele et al., 2017; Kuroda et al., 2012a; Liu 380 381 and Wong, 2013; Sui et al., 2015). River water concentrations of 14 PPCPs in the present study were up to one order of magnitude lower than those detected in the Qing and Liangshui Rivers 382 383 in China (276–6 109 ng/L) (Dai et al., 2016), and in same range of those detected in the Túria 384 River in Spain (average 50 ng/L) (Carmona et al., 2014). The median concentrations of sucralose 385 were 1 140 ng/L in the Pearl River delta (Yuan Yuan Yang et al., 2017a), 2 orders of magnitude 386 higher than those detected in river water in the present study. Similarly, up to 3 orders of 387 magnitude higher concentrations of sucralose were detected in the Haihe River, China (Gan et al., 2013). Noticeably higher levels of cyclamate (0.12 – 0.67 μ g/L) compared to the present 388 389 study were detected in the Haihe River, China (Gan et al., 2013). Similarly, higher levels of cyclamate and sucralose were detected in European rivers (Lange et al., 2012). Although 390 concentrations of PPCPs and ASWs in the present study were lower than those in many other 391 392 regions, mass loadings of some of the PPCPs and ASWs in the Ganges River can be substantially higher or similar to those found in developed countries (Spoelstra et al., 2013) due to the 393 enormous river water discharges of the Ganges. 394

Concentrations of PPCPs in the groundwater along the Ganges River were found to be either 395 lower or in ranges of those detected in various other countries (Sui et al., 2015). For example, 396 groundwater concentrations of sulfamethoxazole (MDL-4.13 ng/L) detected in this study were 397 up to two orders of magnitude lower than those detected in groundwater in vicinity of municipal 398 landfills in Guangzhou, China (29–125 ng/L) (Peng et al., 2014) and in range of those detected 399 400 in groundwater from the Jianghan Plain (<0.8 ng/L) (Tong et al., 2014). Groundwater 401 concentrations of ibuprofen in this study were in the same range as detected in Serbia and Canada (Gottschall et al., 2012; Petrović et al., 2014), while one order of magnitude lower than 402 those reported in Spain, China, and Germany (López-Serna et al., 2013; Peng et al., 2014; Wolf 403 et al., 2012). Groundwater concentrations of ASWs in the present study were up to three orders 404 405 of magnitude lower than those reported in Canadian groundwater along streams (cyclamate<23 ng/L and sucralose<24 µg/L) (Van Stempvoort et al., 2011). Although, groundwater 406

407 concentrations of sucralose found in this study were higher than those detected in Singapore
408 (Tran et al., 2014), China (<9.6 ng/L) (Gan et al., 2013).

Selected PPCPs (ketoprofen, DEET, acetaminophen, and caffeine) and one ASW (cyclamate) in 409 410 the Ganges River water in the pristine Himalayan reach (at Uttarkashi) were found similar to those in the densely populated and industrialized middle and lower reach of the Ganges River. 411 This might be due to the overlap of sampling campaign in Himalayan reach with the tourism 412 413 season. This part of the river also has religious importance and hosts millions of people coming 414 for pilgrimage between April and October. A recent study has elucidated the impact of tourism 415 on levels of PPCPs in Alpine rivers (Mandaric et al., 2017). Moreover, the sampling locations in the Himalayan reach are also the only towns in the region providing health care services and 416 weekly grocery shopping for residents of hundreds of nearby villages. 417

418 There are various possible sources of PPCPs and ASWs in the Ganges River water, including direct discharge of domestic and industrial wastewater in to the river. There are 764 industries 419 420 (including chemical, dairy, food and beverage, and sugar) along the main channel of Ganges and its tributaries, which discharge about 501 million liters per day (MLD) of wastewater. In 421 422 addition, 36 class-I and 14 class-II cities along the Ganges River discharge a total 2 601.3 MLD 423 wastewater into the Ganges River. Total wastewater discharge at the sampling locations in the 424 Himalayan reach is about 3.46 MLD which is 2 orders of magnitude lower than that at the sampling locations in middle and lower reaches (Kanpur: 598 MLD, Varanasi: 410 MLD, and 425 Patna: 233 MLD) (CPCB (Central Pollution Control Board), 2013). In addition to these canalized 426 427 wastewater sources, many household along the river banks discharge wastewater directly. In 428 this study, we did not find any significant correlation of river water concentrations of PPCPs and ASWs with either wastewater discharges volumes into the Ganges River through major drains 429 or population inhibiting within 20 km from sampling locations. 430

Possible sources of PPCPs and ASWs in the groundwater in vicinity of the Ganges River could be 431 bank infiltration, irrigation through Ganges River water followed by leaching to the groundwater, 432 433 leakage from septic tanks (or unpaved septic tanks) and leaching from landfills (e.g. many of which, in India, may illegally receive hospital waste, expired pharmaceuticals, etc.), flaws in 434 435 sewage disposal practices, and unpaved drainage system. The intensity of these sources could vary during the wet and dry seasons depending on the magnitude and direction of infiltration. 436 Several studies have previously used selected PPCP and ASW as markers of wastewater 437 contamination. Caffeine, carbamazepine, acesulfame, sucralose, cyclamate, etc. have been 438 439 indicated as appropriate wastewater indicator substances (Kuroda et al., 2012; Seiler et al., 1999; Yuan Yuan Yang et al., 2017b). In the present study, ketoprofen, DEET, and caffeine 440 441 among PPCPs in river and groundwater and sucralose among ASWs in groundwater were 442 detected with 100% detection frequency. These substances can be considered as appropriate 443 indicators of wastewater contamination in surface and groundwater in the GRB.

444 This is the first study which provides estimates of health and ecological risks associated to PPCPs 445 and ASWs in river and groundwater in India. Results of this study show that all detected PPCPs individually posed no considerable human health concern. However, as previously suggested, 446 447 co-exposure to the PPCP cocktail have different implications for risk estimation (Backhaus and 448 Karlsson, 2014). It has also to be noted that this assessment focuses only on a limited number 449 of PPCPs, thus, the presence of other PPCPs and other emerging and legacy contaminants 450 together should be considered in future studies. Compared to no appreciable risk to human 451 health, moderate risks associated with some of the PPCPs were observed for aquatic organisms (i.e. algae and Daphnia magna). Previous studies also highlighted moderate risks associated 452 453 with some of the PPCPs (namely: sulfamethoxazole and triclocarban) for aquatic organisms in other regions of the world (Du et al., 2017; Tamura et al., 2012; Yun Ya Yang et al., 2017). 454

455 **5. Conclusion**

456 This study shows that both surface and groundwater in the GRB are contaminated by PPCPs and 457 ASWs, which are markers of wastewater contamination. Interestingly, some of the PPCPs and ASWs were detected in the river and groundwater at sampling locations in the pristine 458 459 Himalayas. Previous studies have already shown contamination by other emerging and legacy contaminants in surface and groundwater from the Ganges River basin (Sharma et al., 2016, 460 461 2015). In this study, no considerable human health risk and moderate ecological risk associated to PPCPs and ASWs were estimated. However, health and environmental risk from exposure to 462 a large mixture of emerging and legacy pollutants may be of concern, especially because river 463 464 water (from the Ganges River and its tributaries) and groundwater are the important sources of drinking water and agricultural production for 600 million Indians living in the GRB. In recent 465 years, a few studies have reported serious health problems associated to contaminated water in 466 the Ganges River (Chakraborti et al., 2018). Due to increasing population, urbanization and 467 shifting lifestyle standards from traditional to contemporary, we can expect higher exposure 468 levels in the future, unless appropriate water and waste management solutions will take place. 469 Results reported in this study are a useful baseline for planning and assessing efficacy of possible 470 471 future pollution control measures as part of the Indian white paper on Ganges protection and 472 restauration.

474 Acknowledgement

This study was supported by the Norwegian Research Council's NORKLIMA (215975/E10) 475 program through the project Climate Induced Mobilization of Persistent Organic Pollutants 476 (POPs) in Rivers in India (INDNOPOP). The study was also supported by the CETOCOEN UP 477 478 (CZ.1.05/2.1.00/19.0382) and CETOCOEN PLUS projects (CZ.02.1.01/0.0/0.0/15_003/0000469) and the Ministry of Education, Youth, and Sports of the 479 Czech Republic (RECETOX research infrastructure, LM2015051 and 480 CZ.02.1.01/0.0/0.0/16_013/0001761) under the activity Project of Major Infrastructures for 481 Research Development and Innovations. Authors are grateful to Ondřej Sáňka (RECETOX, 482 Masaryk University) for creating map of study area. 483

485 **References**

- Anumol, T., Vijayanandan, A., Park, M., Philip, L., Snyder, S.A., 2016. Occurrence and fate of
 emerging trace organic chemicals in wastewater plants in Chennai, India. Environ. Int.
 92–93, 33–42. doi:10.1016/j.envint.2016.03.022
- Backhaus, T., Karlsson, M., 2014. Screening level mixture risk assessment of pharmaceuticals
 in STP effluents. Water Res. 49, 157–165.
- 491 doi:https://doi.org/10.1016/j.watres.2013.11.005
- Balakrishna, K., Rath, A., Praveenkumarreddy, Y., Guruge, K.S., Subedi, B., 2017. A review of
 the occurrence of pharmaceuticals and personal care products in Indian water bodies.
 Ecotoxicol. Environ. Saf. doi:10.1016/j.ecoenv.2016.11.014
- 495 Boxall, A.B.A., Rudd, M.A., Brooks, B.W., Caldwell, D.J., Choi, K., Hickmann, S., Innes, E., 496 Ostapyk, K., Staveley, J.P., Verslycke, T., Ankley, G.T., Beazley, K.F., Belanger, S.E., 497 Berninger, J.P., Carriquiriborde, P., Coors, A., DeLeo, P.C., Dyer, S.D., Ericson, J.F., 498 Gagné, F., Giesy, J.P., Gouin, T., Hallstrom, L., Karlsson, M. V., Joakim Larsson, D.G., Lazorchak, J.M., Mastrocco, F., McLaughlin, A., McMaster, M.E., Meyerhoff, R.D., Moore, 499 R., Parrott, J.L., Snape, J.R., Murray-Smith, R., Servos, M.R., Sibley, P.K., Straub, J.O., 500 Szabo, N.D., Topp, E., Tetreault, G.R., Trudeau, V.L., Van Der Kraak, G., 2012. 501 Pharmaceuticals and personal care products in the environment: What are the big 502 questions? Environ. Health Perspect. doi:10.1289/ehp.1104477 503
- Brack, W., Dulio, V., Slobodnik, J., 2012. The NORMAN Network and its activities on emerging
 environmental substances with a focus on effect-directed analysis of complex
 environmental contamination. Environ. Sci. Eur. 24, 29. doi:10.1186/2190-4715-24-29
- Brausch, J.M., Rand, G.M., 2011. A review of personal care products in the aquatic
 environment: Environmental concentrations and toxicity. Chemosphere.
 doi:10.1016/j.chemosphere.2010.11.018
- Brumovský, M., Bečanová, J., Kohoutek, J., Borghini, M., Nizzetto, L., 2017. Contaminants of
 emerging concern in the open sea waters of the Western Mediterranean. Environ. Pollut.
 229, 976–983. doi:https://doi.org/10.1016/j.envpol.2017.07.082
- Carmona, E., Andreu, V., Picó, Y., 2014. Occurrence of acidic pharmaceuticals and personal
 care products in Turia River Basin: From waste to drinking water. Sci. Total Environ. 484,
 53–63. doi:10.1016/j.scitotenv.2014.02.085
- Chakraborti, D., Singh, S., Rahman, M., Dutta, R., Mukherjee, S., Pati, S., Kar, P., 2018.
 Groundwater Arsenic Contamination in the Ganga River Basin: A Future Health Danger.
 Int. J. Environ. Res. Public Health 15, 180. doi:10.3390/ijerph15020180
- 519 CPCB (Central Pollution Control Board), 2013. POLLUTION ASSESSMENT : RIVER GANGA. 520 Delhi.
- Dai, G., Wang, B., Fu, C., Dong, R., Huang, J., Deng, S., Wang, Y., Yu, G., 2016.
 Pharmaceuticals and personal care products (PPCPs) in urban and suburban rivers of
 Beijing, China: occurrence, source apportionment and potential ecological risk. Environ.
 Sci. Process. Impacts 18, 445–455. doi:10.1039/C6EM00018E
- de Jesus Gaffney, V., Almeida, C.M.M., Rodrigues, A., Ferreira, E., Benoliel, M.J., Cardoso,
 V.V., 2015. Occurrence of pharmaceuticals in a water supply system and related human
 health risk assessment. Water Res. 72, 199–208.
 doi:https://doi.org/10.1016/j.watres.2014.10.027
- de Souza, S.M.L., de Vasconcelos, E.C., Dziedzic, M., de Oliveira, C.M.R., 2009. Environmental
 risk assessment of antibiotics: An intensive care unit analysis. Chemosphere 77, 962–967.
 doi:https://doi.org/10.1016/j.chemosphere.2009.08.010
- 532 Department of Pharmaceuticals. Government of India, 2018. Pharma Industry Promotion
 533 [WWW Document]. URL http://pharmaceuticals.gov.in/pharma-industry-promotion

- 534 (accessed 4.23.18).
- Du, J., Zhao, H., Liu, S., Xie, H., Wang, Y., Chen, J., 2017. Antibiotics in the coastal water of
 the South Yellow Sea in China: Occurrence, distribution and ecological risks. Sci. Total
 Environ. 595, 521–527. doi:https://doi.org/10.1016/j.scitotenv.2017.03.281
- Ebele, A.J., Abou-Elwafa Abdallah, M., Harrad, S., 2017. Pharmaceuticals and personal care
 products (PPCPs) in the freshwater aquatic environment. Emerg. Contam.
 doi:10.1016/j.emcon.2016.12.004
- Fick, J., Söderström, H., Lindberg, R.H., Phan, C., Tysklind, M., Larsson, D.G.J., 2009.
 Contamination of surface, ground, and drinking water from pharmaceutical production.
 Environ. Toxicol. Chem. 28, 2522. doi:10.1897/09-073.1
- Gan, Z., Sun, H., Feng, B., Wang, R., Zhang, Y., 2013. Occurrence of seven artificial
 sweeteners in the aquatic environment and precipitation of Tianjin, China. Water Res. 47,
 4928–4937. doi:10.1016/j.watres.2013.05.038
- Gottschall, N., Topp, E., Metcalfe, C., Edwards, M., Payne, M., Kleywegt, S., Russell, P., Lapen,
 D.R., 2012. Pharmaceutical and personal care products in groundwater, subsurface
 drainage, soil, and wheat grain, following a high single application of municipal biosolids
 to a field. Chemosphere 87, 194–203. doi:10.1016/j.chemosphere.2011.12.018
- Hernando, M.D., Mezcua, M., Fernández-Alba, A.R., Barceló, D., 2006. Environmental risk
 assessment of pharmaceutical residues in wastewater effluents, surface waters and
 sediments. Talanta 69, 334–342. doi:https://doi.org/10.1016/j.talanta.2005.09.037
- Jindal, K., Narayanam, M., Singh, S., 2015. A systematic strategy for the identification and
 determination of pharmaceuticals in environment using advanced LC-MS tools: Application
 to ground water samples. J. Pharm. Biomed. Anal. 108, 86–96.
 doi:10.1016/j.jpba.2015.02.003
- Kümmerer, K., Henninger, A., 2003. Promoting resistance by the emission of antibiotics from
 hospitals and households into effluent. Clin. Microbiol. Infect. 9, 1203–1214.
 doi:10.1111/j.1469-0691.2003.00739.x
- Kuroda, K., Murakami, M., Oguma, K., Muramatsu, Y., Takada, H., Takizawa, S., 2012.
 Assessment of groundwater pollution in Tokyo using PPCPs as sewage markers. Environ.
 Sci. Technol. 46, 1455–1464. doi:10.1021/es202059g
- Lange, F.T., Scheurer, M., Brauch, H.J., 2012. Artificial sweeteners-A recently recognized class
 of emerging environmental contaminants: A review. Anal. Bioanal. Chem.
 doi:10.1007/s00216-012-5892-z
- Leung, H.W., Jin, L., Wei, S., Tsui, M.M.P., Zhou, B., Jiao, L., Cheung, P.C., Chun, Y.K.,
 Murphy, M.B., Lam, P.K.S., 2013. Pharmaceuticals in Tap Water: Human Health Risk
 Assessment and Proposed Monitoring Framework in China. Environ. Health Perspect. 121,
 839–846. doi:10.1289/ehp.1206244
- Lillicrap, A., Langford, K., Tollefsen, K.E., 2011. Bioconcentration of the intense sweetener
 sucralose in a multitrophic battery of aquatic organisms. Environ. Toxicol. Chem. 30, 673–
 681. doi:10.1002/etc.433
- Liu, J.-L., Wong, M.-H., 2013. Pharmaceuticals and personal care products (PPCPs): A review
 on environmental contamination in China. Environ. Int. 59, 208–224.
 doi:10.1016/j.envint.2013.06.012
- López-Serna, R., Jurado, A., Vázquez-Suñé, E., Carrera, J., Petrović, M., Barceló, D., 2013.
 Occurrence of 95 pharmaceuticals and transformation products in urban groundwaters
 underlying the metropolis of Barcelona, Spain. Environ. Pollut. 174, 305–315.
 doi:10.1016/j.envpol.2012.11.022
- Mandaric, L., Diamantini, E., Stella, E., Cano-Paoli, K., Valle-Sistac, J., Molins-Delgado, D.,
 Bellin, A., Chiogna, G., Majone, B., Diaz-Cruz, M.S., Sabater, S., Barcelo, D., Petrovic, M.,

- 2017. Contamination sources and distribution patterns of pharmaceuticals and personal
 care products in Alpine rivers strongly affected by tourism. Sci. Total Environ. 590–591,
 484–494. doi:10.1016/j.scitotenv.2017.02.185
- Miller, E.L., Nason, S.L., Karthikeyan, K.G., Pedersen, J.A., 2016. Root Uptake of
 Pharmaceuticals and Personal Care Product Ingredients. Environ. Sci. Technol. 50, 525–
 541. doi:10.1021/acs.est.5b01546
- 589 MoWR, 2014. Ganga Basin Report. New Delhi.
- Mutiyar, P.K., Mittal, A.K., 2014. Occurrences and fate of selected human antibiotics in
 influents and effluents of sewage treatment plant and effluent-receiving river Yamuna in
 Delhi (India). Environ. Monit. Assess. 186, 541–557. doi:10.1007/s10661-013-3398-6
- Narain, S., 2014. GANGA: The river, its pollution and what we can do to clean it. New Delhi.
- Natarajan, P.M., Kallolikar, S., Ganesh, S., 2016. Transforming Ganges to be a Living River
 through Waste Water Management. Int. J. Environ. Chem. Ecol. Geol. Geophys. Eng. 10,
 233–242.
- Peng, X., Ou, W., Wang, C., Wang, Z., Huang, Q., Jin, J., Tan, J., 2014. Occurrence and
 ecological potential of pharmaceuticals and personal care products in groundwater and
 reservoirs in the vicinity of municipal landfills in China. Sci. Total Environ. 490, 889–898.
 doi:10.1016/j.scitotenv.2014.05.068
- Petrie, B., Barden, R., Kasprzyk-Hordern, B., 2014. A review on emerging contaminants in
 wastewaters and the environment: Current knowledge, understudied areas and
 recommendations for future monitoring. Water Res. 72, 3–27.
 doi:10.1016/j.watres.2014.08.053
- Petrović, M., Škrbić, B., Živančev, J., Ferrando-Climent, L., Barcelo, D., 2014. Determination
 of 81 pharmaceutical drugs by high performance liquid chromatography coupled to mass
 spectrometry with hybrid triple quadrupole-linear ion trap in different types of water in
 Serbia. Sci. Total Environ. 468–469, 415–428. doi:10.1016/j.scitotenv.2013.08.079
- Philip, J.M., Aravind, U.K., Aravindakumar, C.T., 2018. Emerging contaminants in Indian
 environmental matrices-A review. Chemosphere 190, 307–326.
 doi:10.1016/j.chemosphere.2017.09.120
- Ramaswamy, B.R., Shanmugam, G., Velu, G., Rengarajan, B., Larsson, D.G.G.J., 2011. GC-MS
 analysis and ecotoxicological risk assessment of triclosan, carbamazepine and parabens in
 Indian rivers. J. Hazard. Mater. 186, 1586–1593. doi:10.1016/j.jhazmat.2010.12.037
- Schriks, M., Heringa, M.B., van der Kooi, M.M.E., de Voogt, P., van Wezel, A.P., 2010.
 Toxicological relevance of emerging contaminants for drinking water quality. Water Res.
 44, 461–476. doi:https://doi.org/10.1016/j.watres.2009.08.023
- Seiler, R.L., Zaugg, S.D., Thomas, J.M., Howcroft, D.L., 1999. Caffeine and Pharmaceuticals as
 Indicators of Waste Water Contamination in Wells. Ground Water 37, 405–410.
 doi:10.1111/j.1745-6584.1999.tb01118.x
- Shanmugam, G., Sampath, S., Selvaraj, K.K., Larsson, D.G.J., Ramaswamy, B.R., 2014. Non steroidal anti-inflammatory drugs in Indian rivers. Environ. Sci. Pollut. Res. 21, 921–931.
 doi:10.1007/s11356-013-1957-6
- Sharma, B.M., Bharat, G.K., Tayal, S., Larssen, T., Bečanová, J., Karásková, P., Whitehead,
 P.G., Futter, M.N., Butterfield, D., Nizzetto, L., 2016. Perfluoroalkyl substances (PFAS) in
 river and ground/drinking water of the Ganges River basin: Emissions and implications for
 human exposure. Environ. Pollut. 208, 704–713. doi:10.1016/j.envpol.2015.10.050
- Sharma, B.M., Nizzetto, L., Bharat, G.K., Tayal, S., Melymuk, L., Sáňka, O., Přibylová, P.,
 Audy, O., Larssen, T., 2015. Melting Himalayan glaciers contaminated by legacy
 atmospheric depositions are important sources of PCBs and high-molecular-weight PAHs
 for the Ganges floodplain during dry periods. Environ. Pollut. 206, 588–596.

- 632 doi:10.1016/j.envpol.2015.08.012
- Spoelstra, J., Schiff, S.L., Brown, S.J., 2013. Artificial sweeteners in a large Canadian river
 reflect human consumption in the watershed. PLoS One 8.
 doi:10.1371/journal.pone.0082706
- Subedi, B., Balakrishna, K., Sinha, R.K., Yamashita, N., Balasubramanian, V.G., Kannan, K.,
 2015. Mass loading and removal of pharmaceuticals and personal care products, including
 psychoactive and illicit drugs and artificial sweeteners, in five sewage treatment plants in
 India. J. Environ. Chem. Eng. 3, 2882–2891. doi:10.1016/j.jece.2015.09.031
- Sui, Q., Cao, X., Lu, S., Zhao, W., Qiu, Z., Yu, G., 2015. Occurrence, sources and fate of
 pharmaceuticals and personal care products in the groundwater: A review. Emerg.
 Contam. doi:10.1016/j.emcon.2015.07.001
- Szekeres, E., Chiriac, C.M., Baricz, A., Szőke-Nagy, T., Lung, I., Soran, M.-L., Rudi, K.,
 Dragos, N., Coman, C., 2018. Investigating antibiotics, antibiotic resistance genes, and
 microbial contaminants in groundwater in relation to the proximity of urban areas.
 Environ. Pollut. 236, 734–744. doi:https://doi.org/10.1016/j.envpol.2018.01.107
- Tamura, I., Kagota, K., Yasuda, Y., Yoneda, S., Morita, J., Nakada, N., Kameda, Y., Kimura,
 K., Tatarazako, N., Yamamoto, H., 2012. Ecotoxicity and screening level ecotoxicological
 risk assessment of five antimicrobial agents: triclosan, triclocarban, resorcinol,
 phenoxyethanol and p-thymol. J. Appl. Toxicol. 33, 1222–1229. doi:10.1002/jat.2771
- Tanoue, R., Nomiyama, K., Nakamura, H., Kim, J.W., Isobe, T., Shinohara, R., Kunisue, T.,
 Tanabe, S., 2015. Uptake and Tissue Distribution of Pharmaceuticals and Personal Care
 Products in Wild Fish from Treated-Wastewater-Impacted Streams. Environ. Sci. Technol.
 49, 11649–11658. doi:10.1021/acs.est.5b02478
- Thatte, C.D., 2018. Water resources development in India. Int. J. Water Resour. Dev. 34, 16–
 27. doi:10.1080/07900627.2017.1364987
- Tong, L., Huang, S., Wang, Y., Liu, H., Li, M., 2014. Occurrence of antibiotics in the aquatic
 environment of Jianghan Plain, central China. Sci. Total Environ. 497–498, 180–187.
 doi:10.1016/j.scitotenv.2014.07.068
- Tran, N.H., Hu, J., Li, J., Ong, S.L., 2014. Suitability of artificial sweeteners as indicators of
 raw wastewater contamination in surface water and groundwater. Water Res. 48, 443–
 456. doi:10.1016/j.watres.2013.09.053
- 663 UNESCO, 1971. Discharges of selected rivers of the world. Monthly and annual discharges 664 recorded at various selected stations II.
- Van Stempvoort, D.R., Roy, J.W., Brown, S.J., Bickerton, G., 2011. Artificial sweeteners as
 potential tracers in groundwater in urban environments. J. Hydrol. 401, 126–133.
 doi:10.1016/j.jhydrol.2011.02.013
- Wolf, L., Zwiener, C., Zemann, M., 2012. Tracking artificial sweeteners and pharmaceuticals
 introduced into urban groundwater by leaking sewer networks. Sci. Total Environ. 430, 8–
 doi:10.1016/j.scitotenv.2012.04.059
- World Bank, 2010. Deep wells and prudence : towards pragmatic action for addressinggroundwater overexploitation in India.
- Yang, Y., Ok, Y.S., Kim, K.-H., Kwon, E.E., Tsang, Y.F., 2017. Occurrences and removal of
 pharmaceuticals and personal care products (PPCPs) in drinking water and water/sewage
 treatment plants: A review. Sci. Total Environ. 596–597, 303–320.
 doi:https://doi.org/10.1016/j.scitotenv.2017.04.102
- Yang, Y.Y., Liu, W.R., Liu, Y.S., Zhao, J.L., Zhang, Q.Q., Zhang, M., Zhang, J.N., Jiang, Y.X.,
 Zhang, L.J., Ying, G.G., 2017a. Suitability of pharmaceuticals and personal care products
 (PPCPs) and artificial sweeteners (ASs) as wastewater indicators in the Pearl River Delta,
 South China. Sci. Total Environ. 590–591, 611–619. doi:10.1016/j.scitotenv.2017.03.001

Yang, Y.Y., Toor, G.S., Wilson, P.C., Williams, C.F., 2017. Micropollutants in groundwater from
septic systems: Transformations, transport mechanisms, and human health risk
assessment. Water Res. 123, 258–267. doi:10.1016/j.watres.2017.06.054

Yang, Y.Y., Zhao, J.L., Liu, Y.S., Liu, W.R., Zhang, Q.Q., Yao, L., Hu, L.X., Zhang, J.N., Jiang,
Y.X., Ying, G.G., 2017b. Pharmaceuticals and personal care products (PPCPs) and artificial
sweeteners (ASs) in surface and ground waters and their application as indication of
wastewater contamination. Sci. Total Environ. doi:10.1016/j.scitotenv.2017.10.241